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A quantitative weight of evidence assessment of confidence in modesof-action and their human relevance



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ABSTRACT

A quantitative weight of evidence (QWoE) methodology was developed to assess confidence in postulated mode(s) of action for adverse effects in animal toxicity studies. The QWoE is appropriate for assessing adverse effects as relevant endpoints for classification and labeling purposes. The methodology involves definition of mode of actions and scoring supporting data for all key steps using predefined criteria for quality and relevance/strength of effects. Scores for all key steps are summarized, and the summary score is compared to the maximal achievable score for the mode of action. The ratio of the summary score to the maximal achievable scores gives an indication of confidence in a specific mode of action in animals. The mode of action in animals with highest confidence is then taken forward to assess appropriateness to humans. If one of the key steps cannot occur in humans, the mode of action is not relevant to humans. The methodology developed is applied to four case studies.

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1. Introduction

The assessment of potential human health risks from exposure to chemicals requires the evaluation of many datasets providing information of widely differing nature. Toxicity testing in experimental animals remains the most common basis for human health risk characterization. Results from toxicity testing are the major basis for classification and labeling (C&L) of chemicals under the Globally Harmonized System of Classification, Labeling and Packaging (GHS) regarding specific toxicities (Dekant and Bridges, 2016b). Clear adverse effects, as defined by WHO/IPCS, in appropriately performed toxicity studies usually trigger classification for specific hazards with consequences such as restriction in use for chemicals classified as toxic to reproduction or as carcinogenic. However, the EU guidance (EC-Regulation, 2008) states that even in the presence of adverse effects, classification may not be appropriate if the adverse effect is only observed in the presence of marked differences in toxicokinetics and/or toxicodynamics between experimental animals and humans, e.g non-linear toxicokinetics (Saghir, 2015). This provision acknowledges the issues

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of animal to human extrapolation and the presence of a number of animal-specific modes of action without human relevance (Corton et al., 2014; Swenberg and Lehman-McKeeman, 1999). The EU guidance does not elaborate the approaches to assessing human relevance except to state that expert judgment and "weight of evidence" should be used.

Weight of evidence provides a more transparent communication of scientific judgments that should be less susceptible to bias (Lutter et al., 2015; US-OSHA, 2016). However, applied weight of evidence evaluations vary widely in their scope. Recently, quantitative weight-of-evidence (QWoE) methods have been developed to evaluate inconsistent databases on the toxicity of chemicals with the aim of generating support for decision-making in classification and labeling (Dekant and Bridges, 2016b) and assessing persistent, bioaccumulative and toxic organic pollutant properties (Bridges and Solomon, 2016). This approach relies on scoring aspects of study quality and reported effects, including weighting of effects depending on the level of biological organization that is influenced or relevance of the endpoint evaluated (Bridges and Solomon, 2016; Dekant and Bridges, 2016b; Van Der Kraak et al., 2014).

Assessment of the human relevance of an observed adverse effect in experimental animals requires information on the mode of action in animals that produces the adverse effect (Borgert et al., 2015). In the past, evaluations of the mode of action for a specific

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chemical and its human relevance relied on narrative descriptions, which may not provide the necessary transparency. Therefore, QWoE, based on predetermined scores for how well the data support a mode of action and the absence/presence of human relevance of the effects in animals, may offer a less bias-prone and more transparent procedure.

To better structure information on mode of action, the World Health Organization (WHO/IPCS) developed a mode-of-action and human relevance framework based on an understanding of toxicity pathways (termed adverse outcome pathways) leading to disease development (Meek et al., 2013, 2014a, 2014b; OECD, 2016). The interaction of a chemical with biological macromolecules, the molecular initiating event, is a fundamental concept (Ankley et al., 2010). Within a mode of action, this molecular initiating event is followed by one or more key events that are connected by a key event relationship that describes the toxicodynamic relationships between individual key events (Becker et al., 2015; Hill, 1965; Meek et al., 2013, 2014a, 2014b; OECD, 2016; Patlewicz et al., 2013). Mode of action needs to be biologically plausible and a number of modes of action for adverse effects have found their way into textbook knowledge (Klaassen, 2013). In many cases, mode of action has been defined based on case studies where plausibility and empirical support for key effect relationships and essentiality of key effects for a disease process have been developed based on test results from a range of chemicals (OECD, 2016). Mode of action may also be based on chemical-specific information.

However, in practice, several different modes of action with potential widely differing relevance to humans may result in the observed adverse effect in experimental animals, and information to support a specific mode of action may be highly variable. Therefore, a comparative evaluation of biological plausibility and experimental support for a specific mode of action and its human relevance is required. While concepts for a systematic comparison of different possible adverse outcome pathways have been developed (Becker et al., 2017), systematic analysis of the confidence in a specific pathway (as compared to other possible pathways), applying quantitative approaches is lacking. Since basic approaches in the application of mode of action analysis have already been incorporated into guidance for regulatory approaches to chemical safety (EC-Regulation, 2008; US-EPA, 2005), a quantitative evaluation of the level of evidence is desirable and should be included in regulatory guidance for risk characterization. The approach presented here may also strengthen conclusions made in systematic reviews.

This manuscript describes a QWoE methodology to compare support for different modes of actions that may cause an adverse effect and their human relevance. The highest score supports the most reliable dataset and supports decisions on human relevance (Fig. 1). To illustrate the utility of the approach, four case studies are evaluated by these criteria. These case studies include a well-recognized male rat-specific mode of action (α_{2u} -globulin nephropathy) as case 1. The 2nd and 3rd cases involve octame-thylcyclotetrasiloxane (D4) for which a narrative human relevance assessment of the animal toxicity data both on reproductive toxicity and tumorigenicity was recently published (Dekant et al., 2017). Case 4 is the developmental toxicity of diethylhexyl phthalate in male rats where a mode of action has been developed to demonstrate human relevance.

2. Methodological approach

2.1. Development of QWoE

A weight of evidence analysis includes definition of the causal question (termed problem formulation by the US EPA),

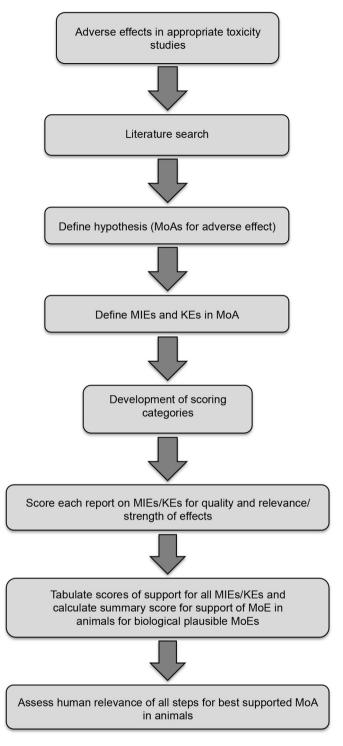


Fig. 1. Basic process of QWoE developed to support hypothesized mode of action for induction of adverse effects by a chemical. MoA = Mode of action, MIE = molecular initiating event, KE = key event. For details of study evaluation, see text.

development and application of criteria for review, evaluation and integration of evidence, and conclusions based on inference (Rhomberg et al., 2013). In this context, the causal questions are i) to what extent is the mode of action for the adverse effects biologically plausible, ii) what is the confidence that a specific mode of action causes the toxic effects, and iii) how relevant to humans is the plausible mode of action considering species differences in

anatomy, physiology, toxicokinetics, and biochemistry. Responses to these questions require development of a quantitative scoring process and criteria for the application of the scores to the question in a stepwise approach (Fig. 1).

2.2. Development of mode of action

The process starts after identification of an adverse effect as defined by WHO/IPCS. The first step is a search for information applying strategies as outlined previously (Dekant and Bridges, 2016b) followed by the development of hypothesized mode of actions including identification of key events and key event relationships. The hypothesized mode of action needs to provide an explanation for the adverse effect regarding target-organ specificity, type of functional change, and induced pathology. Amenable modes of action have well defined sequential key events that are either experimentally measurable or result in measurable biomarkers, because only such molecular initiating/key events can be scored regarding data quality and relevance/strength of effects. While all possible individual steps fulfilling these criteria should be integrated in a mode of action, in practice, most modes of actions consist of four to six individual steps (Becker et al., 2015). Each component of the mode of action needs to be justified by biological plausibility and, if available, analogy of the proposed mode of action with established examples.

Several modes of actions may result in a common adverse effect. For example, tumors may be induced by a genotoxic mode of action involving DNA-damage as the molecular initiating event but may also involve a receptor interaction as the molecular initiating event or cytotoxicity-induced cell proliferation as a key event. Therefore, alternative mode of action need to be developed and comparatively evaluated to assess weight of evidence in support of a specific mode of action.

2.3. Scoring individual studies for quality

QWoE assessments must consider the study design, experimental systems, endpoints addressed, and changes reported (Bridges and Solomon, 2016; Dekant and Bridges, 2016a; Van Der Kraak et al., 2014). Scoring sheets to best cover all quality aspects of the different experimental approaches need to be developed. Quality should be based on best practice for the different endpoints. OECD/EU/US EPA toxicity testing guidelines are useful in identifying best practice for toxicity studies in experimental animals or the assessment of the potential genotoxicity. However, many of the study types required to investigate key steps in a mode of action rely on best practices derived from the scientific literature. These are, at best, only partially covered by basic principles used for guideline development.

Data to support a mode of action may be generated by experiments in intact animals, in cultured tissues or cells, or in subcellular fraction or other cell-free systems. Based on approaches developed previously (Dekant and Bridges, 2016b), quality/reliability criteria were developed for these different study types. These criteria can be considered in an assessment of support for a mode of action. The quality/reliability scoring sheets were developed to assess mechanistic studies in intact animals (Table 1), mechanistic studies in vitro (Table 2), and genotoxicity studies (Table 3).

The scoring criteria for studies in intact animals are based on previously presented tables developed to assess quality of experimental animal studies with some modifications (Dekant and Bridges, 2016b). Scoring criteria for *in vitro* studies are new. Besides potential issues due to inadequately defined identity and purity of the chemical of interest, the quality/reliability criteria for *in vitro* studies also address issues of study design such as the

number of independent repeats of an assay, inclusion of appropriate positive and negative controls, relevance of timing of the application and sampling of the test material, extent of quality assurance, and procedures used for statistical evaluation. Scores also need to reflect the extent of characterization of stability of the test chemical in the application medium. An important issue in in vitro toxicity studies is dosimetry since concentrations of the chemical of interest in the medium may differ from those reaching the target system in an intact animal due to factors such as solubility, loss of material due to volatility, and absorption to the surface of glass or plastic materials used in the experiments. Studies on the genotoxicity of a chemical also require specific scoring sheets, which were developed based on the available guidelines and some of the aspects also considered in scoring the in vitro studies. Scores from 0 to +4 were assigned for all of the study types considered with a score of +4 representing highest quality/reliability (Dekant and Bridges, 2016a).

2.4. Scoring system of individual studies for relevance/strength of adverse effects

Three criteria are proposed for scoring of individual studies for relevance/strength of effects: i) relevance of the model system to the assessment of the molecular initiating/key event or a biomarker of the key event, ii) relevance of the exposure conditions, e.g. concentrations at which there is a change in the key event or biomarker in the model system compared to exposure conditions where an adverse outcome relevant to C&L is produced in an experimental animal, and iii) strength and consistency of effects. Scores from 0 (no relevance) to three (highly relevant) apply to these criteria. Justification for the scores is given in Table 4.

Regarding "relevance of model system," experiments using the specific species and strain of animals in the toxicity studies under exposure conditions relevant to those inducing the adverse effect of interest are considered most appropriate since both toxicokinetics and toxicodynamics are represented (+3). However, well-justified studies in other model systems including genetically modified animals also may be appropriate. Many of the molecular initiating and early key events in a mode of action are difficult to assess in intact animals. Therefore, these events are often assessed in vitro using different systems ranging from freshly isolated cells from the target organ in experimental animals to tissue fractions and homogenates. When scoring for relevance/strength of effects, there is a need to assess whether the system is fit for the purpose of generating adequate data to support molecular initiating and early key events. Such an assessment needs to consider relevance of the in vitro system to target organ anatomy and physiology.

Relevance/strength of evidence scoring also needs to assess exposure conditions that result in an observable molecular initiating and early key events (Becker et al., 2017). Concentrations applied in an experiment to assess key steps may be many orders of magnitude above or below those in the target organ in an intact animal. Such experimental conditions have no relevance in supporting a step in a mode of action and receive a score of 0. Again, exposure of experimental animals under dosing conditions that have induced the adverse effect are most relevant here (+3), but there is a need to consider toxicokinetics regarding duration of the short-term exposure with respect to absorption, time to reach peak blood/tissue levels, and conclusions regarding time points of sampling. For many chemicals, early biomarkers for an effect may be observable after short-term exposures; the only exceptions to be considered here are bioaccumulating chemicals. In this case, shortterm exposures may not result in tissue concentrations sufficient to trigger molecular initiating or early key events. Such cases will require specific considerations regarding time of exposure and

 Table 1

 Score sheet for quality assessment of data from mechanistic studies in intact animals.

| Criterion | Score of 4 | Score of 3 | Score of 2 | Score of 1 | 0 |
|--|---|--|---|---|--|
| Chemical characterization including presence contaminants that may result in confounding. Avoidance of contamination from equipment/feed/dosing solutions; | Fully characterized by performing laboratory or analysis certificate available, source specified, CAS given, impurity analysis conducted, high purity (>99.8%), identified contaminants highly unlikely to interfere with assay. Suitable procedures in place and measurements to ensure compliance, methods well evaluated | contaminants identified and | Compliance for only one of | Not considered, source not or poorly defined, e.g. synthesis in lab and not adequately characterized, limited information on purity and contaminants Contamination appears likely and not considered in the design. | Not described |
| 2. General experimental design (number of animals per dose group, controls, suitability of study duration, housing conditions) | Well designed for purpose including use of adequate positive and negative controls including information on historic | potentially significant limitations identified. Lower number of animals/group or no positive or historic control data available. Some aspects of study duration and | | methodology, such as low numbers of animals (3)/ group, inappropriate controls, inadequate sampling plan | Not suitable |
| 3. Assessment of possible interference from stress due to restraint, toxicity. | well established exposure and sampling system, experienced facility and staff regarding animal handling, toxicity endpoints regarding cytotoxicity and irritation for test chemical well described, controls sham exposed | Experienced staff and appropriate exposure conditions, possible interference by slight irritation or cytotoxicity due | Exposure conditions involve stressful handling, little experience in performing such exposures in performing laboratory, exposure conditions remain in an area of slight irritation | Significant stress due to exposure conditions such as evident irritation/cytotoxicity | No consideration of interference in study design and conduct |
| * * | Checks made on levels of test item in the feed/stability in vehicle if gavage used/intake | duration appear appropriate | No measurements but mode, duration and/or route raises no specific issues | | Unsuitable route of administration, insufficient reporting |
| 5. Appropriate animal species and strain selection and historic data for effects incidences in controls | Appropriate species and strain used, assignment to controls and test groups randomized. Historic | Some deficiencies identified such as not Sprague Dawley rats for reproductive toxicity studies. Full historic data available. | Insufficient information to judge fully or some deficiencies identified | Source not well described | Not mentioned |
| Suitability of sampling method, sampling times and procedures. | Complies with best practice for all sampling including adequate intermediary sampling times to test the hypothesis; sampling times included at several intervals | Some doubts regarding suitability of study design for picking up relevant effects | Substantial doubts about suitability of sampling scheme, e.g. single time point only | Major issues with experimental descriptions, but information provided remains interpretable | Not described or inappropriate |
| | Complies with best practice for all measurements Study blinded to assessor and replicates run, appropriate quality control | Selection not complying fully with best practice limiting inter parameter consistency to be assessed | considerable difficulty in | Limited description for informed conclusions, e.g. no description of QA procedures | |
| 8. Suitability of pathological/ functional assessment | 1 2 | with best practice; e.g., | Limited number of endpoints, considerable difficulty in understanding how methods were validated. No replicate samples or blinding | Only one endpoint assessed, methods not well described | Not suitable or insufficient for purpose |
| 9. Accessibility of raw data | Complete access to all raw data | Limitations in access to data to identify details of methodology used or results. | Difficult to identify important methodological details | Summary data only reported | Data provided very limited |
| 10. Statistical analysis | Appropriate statistical method suitable for analysis of endpoint. Checked for normal distribution | Statistical methodology not optimal but acceptable | Statistical methodology not appropriate although usable for some purposes | Findings too variable to be useful except for qualitative purposes | Not amenable to interpretation |

achieved tissue concentrations. In cultured cells or tissue homogenates, the chemical of interest should be present in concentrations within an order of magnitude of those that have been measured or predicted in the target tissue of the adversely affected experimental animals to achieve a high score for "relevance of concentrations"

used." Effects elicited by unrealistic concentrations of the chemical of interest or nonspecific toxicity in an *in vitro* system may have little relevance for confirming a molecular initiating event in an intact animal. Exceptions are for very simple assays such as receptor binding or solely hazard-related endpoints such as

 Table 2

 Score sheet for quality assessment of data from in vitro toxicity studies.

| Criterion | Score of 4 | Score of 3 | Score of 2 | Score of 1 | 0 |
|---|---|--|---|---|--------------------------------------|
| causing interference, appropriate caution to | Fully characterized by performing laboratory or analysis certificate available, source specified, CAS given, impurity analysis conducted All media from certified sources, well characterized, concentrations applied below limit of solubility, specific design to avoid loss of volatiles from sampling to measurements | Material adequately characterized, major contaminants identified and quantified Well characterized media, design to avoid or limit loss of volatiles | | | Not described |
| 2. General experimental design such as number of assays per dose/concentration, controls, suitability of exposure duration, | Well-designed including use of adequate positive and negative controls and information on historic controls. Adequate number of independent repeats (n > 6), well justified sampling plan with adequate study duration, dose-response assessed in detail | potentially significant limitations identified. Lower number of repeats or no positive or historic control data available. Some aspects of study duration and | Appears to be well described, but no consideration why design selected. Low number of repeats, lack of positive and historic controls, sampling and study duration not well justified, only one concentration used | methodology such as only three determinations of | Not suitable |
| 3. Mode of application of test item system (stability, vehicle used, route of application, dosing intervals, estimation of actual concentration of chemical of interest in medium) | Concentrations of chemical of interest in medium over time determined by analytical procedures, stability of chemical of interest well assessed and solubility well characterized, vehicle controls | Concentrations only assessed | Concentrations in media only calculated based on amount added to system | | Not suitable |
| 4. All assessments include determination of toxicity to model organism | Detailed toxicity assessment by appropriate methods in controls and exposed system, cytotoxicity of chemical of interest in system well defined and reported | assessment based on methodology or limited | Toxicity assessment limited to few measurements under assay conditions, but toxicity not evident from other parameters | Very limited information on cytotoxicity | Not suitable |
| Suitability of sampling method, sampling times and procedures. | Sampling procedures well justified based on analogy/ previous experience and time course of response assessed in positive and negative controls, deviations well justified | | Only few samples collected and evaluated, limited justification for sampling plan | Only few samples collected, no justification for sampling plan | Not suitable |
| 6. Suitability of biochemical measurements including quality control | Complies with best practice for all measurements. Study blinded to assessor and replicates run, appropriate quality control | Selection not complying fully with best practice limiting inter parameter consistency to be assessed | Limited number of endpoints, considerable difficulty in understanding how methods applied | Limited description for informed conclusions, e.g. no description of QA procedures | |
| 7. System used for biotransformation/cells have capacity to simulate relevant reactions that occur with chemical of interest in animals | functional biotransformation system integrated, functionality assessed with adequate positive control, well characterized cell type with enzyme activities determined in conducting laboratory, biotransformation pathways for chemical of interest characterized | biotransformation to induce effects included, but only limited information on biotransformation of chemical of interest. Capacity for biotransformation | effect included, but no information on biotransformation of | Little experience regarding biotransformation capacities of system, only taken from literature | No relevant information |
| 8. Accessibility of raw data | Complete access to all raw data | Limitations in access to data to identify details of methodology used or results. | Difficult to identify important methodological details | Summary data only reported | Data provided very limited |
| 9. Statistical analysis | Appropriate statistical method suitable for analysis of endpoint. Checked for normal distribution | Statistical methodology not optimal but normal distribution checked | Statistical methodology not appropriate although usable for some purposes | Findings too variable to be useful except for qualitative purposes | Not amenable to interpretation |

genotoxicity, where, in the absence of cytotoxicity, limited solubility or pronounced changes in osmolality may limit maximal concentrations applied. For genotoxicity, specific guidance from OECD is available regarding concentrations to be applied, and a score of +3 can be given when these requirements are fulfilled.

The 3rd criterion, "strength of effects," assesses the magnitude

of change in a molecular initiating and early key events. A score of +3 (strong support) applies when the measured changes are consistent over time and highly significant, scores of +2 (moderate support) may be selected if the effect is less pronounced and has a lower statistical significance, and scores of +1 (weak support) can be given for effects that are measurable, but have limited

Table 3Score sheet for quality assessment of genotoxicity studies.

| Criterion | Score of 4 | Score of 3 | Score of 2 | Score of 1 | Score of 0 |
|---|---|---|--|--|--|
| Chemical well characterized including presence contaminants | Fully characterized, analysis certificate available, source specified, CAS given, impurity analysis conducted | Material adequately characterized, major contaminants identified and quantified | Reliance on supplier for information on identity and purity | Not considered, source not or poorly defined, e.g. synthesis in lab and not adequately characterized | information |
| of replicates per concentration, and | Well-designed including use of adequate positive and negative controls and information on historic controls. Study design consistent with guideline including recommended concentrations of test chemical | limitations identified. Lower number of repeats or no historic control data | Appears to be well described, but no consideration why design selected. Low number of repeats, lack of historic controls, sampling and study duration not well justified, | Potentially flaws in the methodology such as inappropriate controls, inadequate sampling plan and/or study design | Not useful |
| 4. Mode of application of test item to system and appropriate test system (stability, vehicle used, route of administration) | Recommended solvent, test system from established supplier as recommended by guideline | Some limitations such as application in less widely used solvent, | Use of rarely applied solvent, some uncertainty regarding addition of chemical of interest and resulting concentrations | appropriate, insufficient | Unsuitable, insufficient reporting |
| 5. Appropriate metabolic activation system when required | According to guideline and obtained from established supplier or specifically justified due to known pathways of bioactivation | Standard activation procedures applied, system generated in performing laboratory | Limitations regarding activation system | Possible issues with metabolism not considered | No information |
| 7. Suitability of the procedures used to assess genotoxicity | Procedures consistent with respective guidelines | System used not covered by available testing guidelines, but significant experience with performance available | System not covered by guideline and limited information on performance available | Method and QA description has significant limitations | Not described |
| 8. Accessibility of raw data | Full access | Only limited access to raw data | Raw data not accessible, but detailed description of results | Summary data only reported, questions if all generated data were reported | Data provided very limited |
| 9. Statistical analysis | Fully appropriate | Some significant variations between observations, but appropriate statistical tests | Substantial variation between observations, limitations regarding statistical treatment of data | Findings too variable to | Not amenable to interpretation |

Table 4 Scoring criteria for relevance/strength of effects.

| Score | Weak, 1 | Moderate, 2 | Strong, 3 |
|--|---|---|--|
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | Concentrations required to induce effect are at least two orders of magnitude above concentrations of the chemical of interest reasonably expected in tissue under exposure conditions causing the adverse effect | Concentrations required to induce effect are one order of magnitude above concentrations of the chemical of interest reasonably expected in tissue under exposure conditions causing the adverse effect | Concentrations required to induce effect are in the range of concentrations of the chemical of interest reasonably expected in tissue under exposure conditions causing the adverse effect |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | Uncertainty regarding suitability of endpoint or biomarker to reflect critical endpoint in vivo, limitations of model system | Established model system, but some limitations regarding relevance of endpoint determined for sequence of events resulting in adverse effect in vivo | Endpoint or biomarker is clearly compatible with key event in vivo in mode of action, model system applied is highly relevant |
| Strength of effects | Changes in endpoint or biomarker observed, but no dose or time dependence and limited statistical significance (only $p \geq 0.05$) | Changes in endpoint or biomarker | Consistent and time- and dose-related change in assessed endpoints, several measurements show significant changes $(p < 0.05)$ |

significance or are observed only at a single time point.

2.4.1. Deriving a score to support an individual step in a mode of action

Overall support for a molecular initiating/key event is obtained by multiplying the mean score for quality (0 to +4) with the mean overall score for relevance/strength of effects. Studies with adequate study design and detailed reporting usually give quality/reliability scores well above three, while average quality/reliability scores between two and three indicate some significant limitations. Quality/reliability scores below two indicate inadequate quality likely insufficient to come to valid conclusions based on study outcome. For transparency, the approach should integrate all studies, even when quality is well below an average of two. However, exclusion criteria for low quality studies may be applied if

defined before conducting the QWoE.

The relevance/strength of effects score is obtained by multiplication of the scores obtained for relevance of exposure conditions, relevance of model system, and magnitude of effect (maximum of 27). Multiplication is used here because studies that use irrelevant exposure conditions, an irrelevant model system, or produce no response (scores of 0) do not support a molecular initiating/key event. In case several studies address a specific molecular initiating/key event with the chemical of interest, the mean of the overall study scores is used to calculate the scores for overall support of the mode of action.

2.4.2. Calculation of overall support for a mode of action in experimental animals

To obtain a summary score to assess experimental support for a

mode of action, weighting factors for molecular initiating/key events are needed. Usually, molecular initiating/key events are supportive for a specific mode of action, while late kev events represent markers of a disease process that may have been initiated by upstream mechanisms (Becker et al., 2017). Therefore, late key events receive a relative weight of 0.33 in the final calculation of an overall support score for a mode of action whereas molecular initiating/early key events receive a weight of 1. The individual scores for the molecular initiating event and all weighted key events are added to give an overall score reflecting support for a mode of action. This procedure integrates supporting information and data that do not support a specific step, i.e., those that received a score of zero. Mode of action with good experimental support will receive scores close to the maximum score possible when high quality studies were performed in relevant model systems, applying relevant concentrations, and obtaining clear results. In contrast, datasets giving little support will receive a low overall score indicating that the mode of action is unlikely to account for the adverse effect induced by the chemical of interest. The overall confidence score of the dataset is compared to the maximum score achievable assuming that all studies have a quality score of +4 and all individual steps receive a score of 27 (3 \times 3 \times 3). A summary score for the mode of action of >75% of the maximal achievable score (quality scores of +4 for all studies and use of relevant models and concentrations with clear effects) is considered to provide very good support a specific mode of action in the test species since a quality rating of +4 for a study is not often obtained, especially for mechanism-oriented studies with many complex interferences and sometimes less pronounced changes in early endpoints. Summary scores between 50 and 75% of the maximum score achievable present moderate support indicating that data support for key events is only partially available, whereas summary score below 50 indicate only weak support for the hypothesized mode of action. Summary scores below 50 will results for datasets where several key events have limited support or where there is no support for one key event. In practice, scores for a mode of action below 30% of the maximal achievable score indicate absence of support. Low scores are mainly driven by the scores received for the late key events that are common to several modes of action and usually do not include mechanistic information. Since late key events are integrated in the calculation of summary scores, albeit with a reduced weight, summary scores of 0 for a mode of action cannot be achieved. Therefore, support scores up to the maximal score achievable for the late key events cannot be considered as support for a mode of action. The cut-offs of 30, 50, and 75% of the maximum score for the determination of support are arbitrary; an extension of the application of the QWoE-methodology to cover a larger dataset of model compounds may be needed to identify values that reliably identify the strength of support.

2.4.3. Identification of the best-supported mode of action in an animal model based on overall scores

Where there are two or more biologically plausible modes of actions that may account for an adverse effect, these are scored using identical criteria. The overall score and its relation to the maximal score achievable for a specific mode of action gives an indication of confidence and thus permits comparison of confidence in the different modes of actions. For the assessment of human relevance, the mode of action with best support in experimental animals (i.e. highest score) is selected for assessment of human relevance. Because the scoring system manipulates ranks (1, 2, 3) arithmetically as numbers and assumes equal strength of similar ranks in different domains, a comparison of scores will be most reliable when differences in scores are high. Small differences in scores may not be meaningful given the underlying assumptions

about the ranks representing numerical quantities.

2.4.4. Score human relevance for best-supported mode of action in experimental animals

For scoring human relevance of the best supported mode of action in experimental animals, we propose using a simple scoring system with a score of zero for an individual step that does not occur in humans due to basic differences in anatomy, physiology, or biochemistry. A score of 1 is assigned if this step may occur in humans or if there is no relevant information to support a conclusion on human relevance. A score of zero breaks the keyevent relationship. Thus, the mode of action for the resulting adverse effect induced by the chemical in experimental animals cannot be propagated to the apical endpoint, and the overall mode of action is not supported in humans (Bridges and Solomon, 2016).

3. Case studies

To demonstrate the applicability of the QWoE approach for assessment of human relevance, the methodology is applied in four case studies. Case 1, renal tumors in male rats induced by inhalation of MTBE due to accumulation of α_{2u} -globulin in the kidney, is a well described example for a mode of action that is recognized to be without human relevance (Swenberg et al., 1989; US-EPA, 1991). Cases 2 and 3 describe the application of the QWoE-methodology to octamethylcyclotetrasiloxane (D4) and to two separate effects induced by inhalation of D4, impaired female fertility and induction of benign uterine tumors observed in rats. Narrative assessments regarding the potential human relevance of these D4-induced have been published (Dekant et al., 2017). The mode of action for these two cases involving D4 are similar, and the cases are used here to demonstrate the differences in weighting of the same studies when considering different apical end-points. Case 4 represents diethylhexyl phthalate, a data-rich chemical that induces genital malformations in male offspring of rats with a specific mode of action for which human relevance is discussed (Boberg et al., 2011; Furr et al., 2014; Johnson et al., 2012; van den Driesche et al., 2015; Wilson et al., 2008).

3.1. Application of the QWoE methodology to assess human relevance of renal tumors in male rats induced by long-term inhalation of methyl tert butyl ether (MTBE)

3.1.1. Summary of the relevant toxicology of MTBE

MTBE is used as an additive in gasoline at concentrations between 2 and 15% and human exposures are likely (Stern and Kneiss, 1997; Vainiotalo et al., 1999). The toxicology of MTBE has been intensively investigated, and only studies relevant for a proof of concept regarding human relevance of a mode of action are reviewed here. Several studies have implicated the kidney as a target organ for toxicity after repeated exposure of male rats to MTBE. MTBE is consistently negative in genotoxicity testing (McGregor, 2006). In an inhalation study, Fischer 344 rats were exposed to 0, 400, 3 000, or 8000 ppm MTBE for 24 months (Bird et al., 1997). The kidney was the main target of MTBE toxicity, and the incidence of renal tubular cell tumors was increased in male rats at the intermediate dose but not at the highest dose. The lack of dose-dependency may be due to decreased survival in the high-dose group due to early death from progressive nephropathy.

Renal toxicity and renal tumor induction were not seen in female rats. MTBE was also tested for carcinogenicity in mice exposed to the same MTBE concentrations in air as rats, without effects on the kidney (Bird et al., 1997). Species differences in toxicokinetics and biotransformation of MTBE do not account for the specific effects of MTBE on the kidney of male rats (Hutcheon et al., 1996).

The developed QWoE-methodology involves a) defining mode of actions to account for the adverse effect of concern; b) scoring the quality of the experimental support for individual steps in the mode of action, c) comparing summary confidence scores for the mode of actions, d) selecting of the best supported mode of action based on the scores obtained, and e) scoring human relevance of the best supported mode of action for the adverse effect in animals. For renal tumors induced by MTBE, two possible modes of action were developed based on mechanisms of tumorigenicity of chemicals in the rodent kidney (Dekant and Vamvakas, 1992; Hard, 1998; Lock and Hard, 2004).

The first hypothetical mode of action is that the tumor induction by MTBE is related to the ability of MTBE to impair the degradation of the male rat -specific protein α_{2u} -globulin and induce a sequence of changes in renal pathology, finally ending in renal tumors. The sequence of individual key steps is shown in Table 5. This mode of action has been demonstrated with a variety of other chemicals that cause male rat-specific kidney tumors and impair the degradation of α_{2u} -globulin (Borghoff et al., 1990; Swenberg and Lehman-McKeeman, 1999; Swenberg et al., 1989). The alternative mode of action proposes tumor induction by DNA-damage and induction of mutations by MTBE. A genotoxic mode of actions has been implicated for some other chemicals that induce renal tumors in rodents (Dekant and Vamvakas, 1992; Hard, 1998; Lock and Hard, 2004).

Scoring of the role of α_{2u} -globulin-induced male rat-specific nephropathy gives a high overall score based on conclusive experimental support for the identified individual key steps based on targeted studies with high quality. The mode of action consists of a molecular initiating event, binding of MTBE to α_{2u} -globulin and a series of early and late key events (Table 5). In this case study, support for the individual key events is from high quality studies (quality score of 3.7) that were specifically targeted to the key events in the hypothesized mode of action in highly relevant system (intact rats) applying concentrations identical to those inducing the adverse effect. Therefore, scores of 3 for all aspects of relevance/strength of effects are obtained resulting in an overall

relevance score of 27 (3 \times 3 \times 3) for all key steps. Multiplication of the relevance/strength of effects score by the quality scores then translates to a high weighted score for the individual key events. The summary score for the α_{2u} -globulin mode of action for MTBE is close to the maximal score achievable (90%, Table 5). Several other appropriately performed repeated dose studies also have described the characteristic pathology of α_{2u} -globulin nephropathy after MTBE exposures (Cruzan et al., 2007) and the available information on MTBE fulfills the US EPA criteria (Swenberg and Lehman-McKeeman, 1999) for establishing the role of α_{2u} -globulin nephropathy in male rat renal carcinogenesis. The very high score indicates strong support.

A genotoxic mode of action requires MTBE or its metabolites to interact with cellular DNA in the target tissues and cause damage to the genome. Inaccurate repair of DNA-damage causes mutations that may alter the course of cell differentiation The genotoxicity of MTBE has been intensively investigated (for overviews, see (Cruzan et al., 2007; McGregor, 2006; McGregor et al., 2005)). MTBE was not mutagenic in bacteria, did not induce mitotic gene conversion in a yeast, and did not induce chromosome damage, gene mutation or DNA-damage in mammalian cells or in intact animals (IARC-Monographs, 1999). The few positive studies (Cruzan et al., 2007) "were conducted with inappropriate methods, published only as abstracts, or not confirmed by other adequate studies".

Scoring of experimental support for a mutagenic mode of action for MTBE shows that a mutagenic mode of action has no support due to the predominantly negative genotoxicity database on MTBE and its known metabolites (11.4%, Table 6). Due to the many studies available and the time consuming scoring of quality, an average quality of 3.3 (expected quality score for a database consisting both of genotoxicity studies performed according to OECD guidelines and other laboratory studies; most of the MTBE studies were performed following testing guidelines) was assumed for the database on the genotoxicity of MTBE.

Table 5 Scoring for quality and relevance/strength for the sequence of individual steps in the pathogenesis of α_{2u} -globulin nephropathy induced by methyl tert.butyl ether (MTBE) in male rats. MIE = molecular initiating event, KE = key event.

| Key step in mode of action | Data support | | Relevance/strength of evidence score evidence | Weighted total score for MIE/ KE |
|--|--|-----|---|--|
| Binding of MTBE/MTBE- metabolite to α _{2u} -globulin | Binding of MTBE to α_{2u} -globulin in vivo, displacement of MTBE-derived radioactivity from renal protein by more potent ligand (Prescott-Mathews et al., 1997; Williams and Borghoff, 2001) | | Model 3 Concentrations 3 Strength of effect 3 | 99.9 |
| Impaired lysosomal degradation of α_{2u} -globulin with bound chemical | Supported by accumulation of α_{2u} -globulin in male rat kidney after MTBE-exposure (Prescott-Mathews et al., 1997) | 3.7 | Model 3 Concentration 3 Effect 3 | 99.9 |
| Accumulation of α _{2u} -globulin in proximal tubular epithelial cells | Demonstrated by observation of protein droplets after short term MTBE exposure (Prescott-Mathews et al., 1997), | 3.7 | Model 3 Concentration 3 Effect 3 | 99.9 |
| Lysosomal swelling/cytotoxicity | Demonstrated in (Prescott-Mathews et al., 1997), and other studies summarized in (Cruzan et al., 2007) | 3.7 | Model 3 Concentration 3 Effect 3 | 99.9 |
| Cell death observed by histopathology | Observed with MTBE after repeated exposure (Prescott-Mathews et al., 1997), | 3.7 | Model 3 Concentration 3 Effect 3 | Relative weight 0.33 33.3 |
| Regenerative cell proliferation, specific histopathology | Observed in several studies (Cruzan et al., 2007) | 3.7 | Model system 3 Concentration 3 Effect 3 | Relative weight 0.33 33.3 |
| Male rat specific renal tumors | Yes, but limited dose response due to early death in male rats (Bird et al., 1997) | 3.8 | Model system 3 Concentration 3 Effect 2 | Relative weight 0.33 22.2 |
| | Total score for mode of action Max. score achievable Percent of maximum | | | 488.4 540 90.4 |

Table 6Scoring for quality and relevance/strength for the sequence of individual steps in the pathogenesis of renal tumors by MTBE in male rats induced by a mode of action involving DNA-damage and induction of mutations. MIE = molecular initiating event, KE = key event.

| Key step in mode of action | Data support | Quality score(s) | Relevance/strength of evidence score evidence | Weighted total score for MIE/KE |
|--|---|---------------------|---|------------------------------------|
| DNA reactivity leading to mutation | All high-quality genotoxicity studies are negative (Cruzan et al., 2007; McGregor et al., 2005) | 3.3 | Model system 3 concentrations applied 3 strength of effect 0 | 0 |
| Insufficient repair of mutations | No DNA-repair induced (Cruzan et al., 2007; McGregor et al., 2005 |) 3.3 | Model system 3 Concentrations applied 3 Strength of effects 0 | 0 |
| Perturbation of cell growth and survival | Yes, (Prescott-Mathews et al., 1997), also supported by other studies | s 3.7 | Model system 3 Concentrations applied 3 Strength of effects 1 * | 33 × 0.33 (late KE) = 11 |
| Male rat specific renal tumors | Yes, but limited dose response due to early death in male rats (Bird et al., 1997) | 1 3.8 | Model system 3 concentrations applied 3 strength of effect 2** | 66 × 0.33 (late KE) = 22 |
| Total score for mode of action | on | | | 33 |
| Max. score achievable | | | | 287.3 |
| Percent of maximum | | | | 11.4 |

^{*} low score for strength of effect due to highly specific pathology selectively observed in male rats, distinct from other pathologies resulting in kidney tumors; ** lower score due to non-linear increase in tumor incidence due to high mortality.

3.1.2. Scoring of the best-supported mode of action for MTBE regarding human relevance

The mode of action involving binding of MTBE to α_{2u} -globulin received very high support in the scoring of individual steps and in support of the overall mode of action since it scored much higher than the alternative mechanism. This mode of action is taken forward to human relevance assessment. Table 7 summarizes the results of the human relevance assessment and an analysis of concordance between renal tumor induction and the presence of α_{2u} -globulin in species where the tumorigenicity of MTBE has been assessed. The available information shows that α_{2u} -globulin is not biosynthesized in female rats and in mice that are insensitive to renal tumor induction by MTBE (IARC-Monographs, 1999). In addition, α_{2u} -globulin is not present in humans and proteins in human kidney do not interact with other chemicals that induce

 α_{2u} -globulin nephropathy (Cruzan et al., 2007; McGregor, 2006). Due to these basic differences in biochemistry and physiology regarding the molecular initiating event, the chain of key events operative in male rats with MTBE is not operative and thus cannot progress to the apical endpoint in humans. Therefore, the overall mode of action identified for male rats is not relevant to humans.

3.2. Female rat-specific reproductive toxicity of octamethyltetracyclosiloxane (D4)

Octamethylcyclotetrasiloxane (D4) is a cyclic siloxane primarily used as a monomer or intermediate in the production of silicone polymers resulting in potential exposure of workers and potential low level inhalation or dermal exposure for the general public. D4 is an odorless liquid that is highly volatile. Human exposure occurs by

Table 7

Human relevance scoring of the α_{2u} -globulin mode of action for MTBE-induced renal tumors in male rats (0, not relevant since specific step in mode of action is not possible in humans due to species differences in biochemistry/physiology/anatomy; 1, step possible based on human biochemistry/physiology/anatomy). If an early step in a mode of action is scored as 0, further downstream steps are not possible and the mode of action is therefore not relevant in humans.

| Essential step in mode of | Data support | | Possible in Humans |
|--|--|--|---|
| action | Male rats | Female rats, male and female mice | |
| Binding of MTBE/MTBE-metabolite to α_{2u} -globulin, | Binding of MTBE to α_{2u} - globulin in systemic circulation | Not possible since α_{2u} -globulin not expressed, structurally similar proteins do not bind chemicals that induce α_{2u} -globulin nephropathy in male rats 0 | Not possible since α_{2u} -globulin not expressed, α_{2u} -globulin structurally similar proteins do not bind chemicals that induce α_{2u} -globulin nephropathy in male rats α_{2u} -globulin nephropathy in α_{2u} -globulin ne |
| Impaired lysosomal degradation of α _{2u} - globulin with bound chemical, | Accumulation of α_{2u} -globulin in kidney due to impaired renal degradation, 1 | 0 | 0 |
| Accumulation of α _{2u} - globulin in proximal tubular epithelial cells | Protein droplets after short term MTBE exposure, 1 | 0 | 0 |
| Lysosomal swelling/ cytotoxicity | Demonstrated after repeated exposure 1 | 0 | 0 |
| Cell death observed by histopathology | Demonstrated after repeated exposure | 0 | 0 |
| Regenerative cell proliferation, specific hiostopathology | Demonstrated after repeated exposure | 0 | 0 |
| Male rat specific renal tumors | Demonstrated after repeated exposure 1 | 0 | 0 |
| Total | 1 | 0 | 0 |

inhalation and dermal contact, although the volatility of this compound makes inhalation the most important potential route of exposure. A detailed toxicity database for D4 is available including a number of mechanistic studies performed to address mode of actions for relevant adverse effects observed in the toxicity testing (for an overview, see Dekant et al., 2017; Franzen et al., 2017).

In a reproduction toxicity study, rats were exposed by inhalation to D4 concentrations of up to the maximum achievable vapor concentration, 700 ppm. Exposure of female rats by inhalation of D4 at 700 ppm resulted in a decrease in the number of corpora lutea, the number of uterine implantation sites, and litter size (Siddiqui et al., 2007). A more detailed reproduction study in female rats exposed by inhalation to D4 was reported (Kaufman, 1998) and subsequently published (Meeks et al., 2007). The sensitive time period for the decrease in implantations and litter size was isolated to the peri-fertilization phase, from three days prior to mating to gestation day three. There was a decrease in fertility with exposure for six hours on the day prior to mating. These data point to an inhibition of ovulation or corpus luteum function as a key event in the reduction of female fertility after exposure to D4. This mechanism was confirmed (Quinn et al., 2007a) when D4 was shown to inhibit the pre-ovulatory LH surge causing a delay in ovulation, persistent follicles, and a prolonged exposure to elevated estrogen in the adult Sprague-Dawley rat. The QWoE methodology was applied to two possible modes of action scenarios to assess their experimental support and to evaluate the human relevance. The competing scenarios propose molecular initiating events based either on dopamine activity by D4 or estrogenicity of D4. The chain of key events for these competing scenarios and their scores are shown in Tables 8 and 9. The quality assessments of all underlying studies are shown in the Supplemental Material. There are other conceivable mode of actions that could be proposed to explain ovulatory disturbance. For example, alterations by D4 in noradrenergic activity or receptor binding have been investigated in vitro (Elias, 2009; McMullin, 2009). The evidence for an adrenergic mechanism of ovulatory disturbance is scanty, and this mechanism was therefore not considered in this QWoE analysis. Because the QWoE is data-driven, it is limited to considerations of mode of actions for which there are adequate data.

3.2.1. Dopamine activity mode of action

This mode of action involves interaction of D4 with the dopamine system causing increased dopamine activity. Increased dopaminergic activity may result in decreased prolactin and impairment in ovulation and corpus luteum function in rats (Bachelot and Binart, 2007). Inhibition/delay of ovulation and/or inadequate corpus luteum formation results in decreased mating and decreased fertility. There is inadequate evidence for a direct interaction of D4 with dopamine receptor(s) suggesting that postreceptor events are more likely (Dekant et al., 2017; Franzen et al., 2017; Jean and Plotzke, 2017). However, a mode of action based on dopamine activity is supported by studies showing dopamine-like effects of D4 in in vitro systems (Dekant et al., 2017; Franzen et al., 2017; Jean, 2005; Jean and Plotzke, 2017) and an observed decrease in prolactin, secretion of which is inhibited by dopamine, in in vivo experiments (Dekant et al., 2017; Franzen et al., 2017; Jean, 2005; Jean and Plotzke, 2017; Quinn et al., 2007a). The downstream key events (decreased prolactin and LH surge) in this mode of action have been well established for D4 using in vivo studies. However, one of the available datasets on the prolactin decrease and/or the decreased LH surge did not demonstrate an effect (Dekant et al., 2017; Elias, 2010; Jean and Plotzke, 2017) resulting in reduced scores for some of the key events due to inconsistent data.

The QWoE scoring uses the quality and strength/relevance

scores of the underlying studies in the calculation of a mean score for each key event. Table 8 shows the scoring for the dopamine agonist mode of action for female reproductive toxicity of D4 based on the quality and relevance/strength of effects scoring (see Supplemental Material). For several of the key events, different datasets are available and the scores integrated into calculated overall score for the key event are means of the scores for the individual studies and their results. The last two steps received a weighting of 0.33 because they are late events and appear in both mode of action tables. This mode of action received a score of 44% of the maximum score achievable, which is considered weak support.

3.2.2. Estrogenic activity mode of action

Interaction of D4 with the estrogen receptor and downstream consequences of this interaction can be a basis for adverse effects on female rat fertility (Table 9). With regard to experimental support for this mode of action, binding of D4 to estrogen receptor- α as molecular initiating event was demonstrated in cell-free systems (Quinn et al., 2007b); binding activated the receptor and resulted in estrogenic activity (He et al., 2003; McKim et al., 2001; Quinn et al., 2007b). However, the estrogenic activity of D4 is many orders of magnitude less than the reference estrogens ethinyl estradiol and diethylstilbestrol and about two orders of magnitude less than the common food estrogen coumestrol. Therefore, low scores were given for "relevance of concentrations" in the studies supporting the molecular initiating event and the 1st key step. Estrogens at certain exposure levels trigger release of LH from the pituitary, but high or prolonged estrogen exposure is expected to suppress pituitary LH by altering gonadotropin-releasing hormone (GnRH) production from the hypothalamus (Tng, 2015).

There is no data support for an effect of D4 on gonadotropinreleasing hormone production by rat hypothalamic explants (Meeker, 2009). The last two key events are identical in both a dopamine activity and an estrogenic mode of action for D4-induced effects on female rat fertility. In addition to the very low scores for experimental support, the estrogen mode of action pathway cannot be supported based on the break in the chain of key events. Even if the broken chain is ignored, this mode of action scored only 18.7% of the possible maximum, clearly inferior to the dopaminergic activity mode of action.

3.2.3. Human relevance

The next step is an evaluation of the human relevance of the dopamine activity mode of action, which is best supported (Table 10). While binding of D4 to the dopamine receptor may be considered possible in humans the available data do not support a direct interaction of D4 with the dopamine receptor (Dekant et al., 2017; Franzen et al., 2017; Jean and Plotzke, 2017). Regardless of the molecular initiating event, an increase in dopamine activity that decreases prolactin in humans is not relevant to human ovulation or corpus luteum maintenance, because prolactin is not important in ovulation in primates. Prolactin null mice have irregular estrous cycles and do not conceive (Bachelot and Binart, 2007). When these mice ovulate, the corpus luteum does not form normally and if conception occurs, pregnancy does not continue. By contrast, prolactin is not important in primate ovulation and, indeed, excessive prolactin interferes with ovulation, even if the excess is transient and clinically unapparent (Suginami et al., 1986). Dopamine agonist medications are used to treat ovulatory disturbances attributed to prolactin excess in women (Anon, 2004). Because there are no data suggesting that D4 binds to the dopamine receptor and because dopamine agonism does not interfere with ovulation in women, the species differences in this key event break the chain. Therefore, the mode of action that best explains the adverse effects of D4 on fertility in female rats is not relevant to humans.

Table 8 Scoring for quality and relevance/strength for the sequence of individual steps for a dopamine agonism mode of action for inhibition of ovulation in female rats exposed by inhalation to D4 at 700 ppm. MIE = molecular initiating event, KE = key event.

| Key steps in mode of action | Data support | Quality scores (from supplemental tables) | Relevance/ strength of evidence score | Weighted score for MIE/ KE |
|---|---|---|---|---|
| Increased dopamine activity | MMQ cells (a rat pituitary tumor cell line) produced less prolactin after exposure to D4 (Jean, 2005) and had decreased forskolin-stimulated cyclic AMP at D4 concentrations \geq 25 μ M without cytotoxicity (Domoradzki, 2011). Not mediated by | 2.3 | Concentrations: 2 Effect: 2 | 18.4 |
| | D2 receptor (not blocked by raclopiride) or G-protein (not blocked by pertussin toxin) (Domoradzki, 2011). | (Jean, 2005) 2.2 | Model: 2 Concentrations: 1 Effect: 1 | 4.4 |
| | | | Mean | 11.4 |
| Decreased prolactin | Sprague-Dawley rats exposed to D4 700 or 900 ppm on diestrus 1–2 and proestrus had decreased prolactin at 1400 h on proestrus (Quinn et al., 2007a). This effect was attributable to the non-ovulatory animals. In another study, prolactin was | | Model: 3 Concentrations: 3 Effect: 2 | 64.8 |
| | decreased 18 h but not 8 h after inhalation of D4 700 ppm in Fischer 344 rats that had been dopamine-depleted by the administration of reserpine (Llames, 2010). However, (Elias, 2010) did not show a decrease in prolactin in 20-month-old female | , | Model: 1 Concentrations: 3 Effect: 0 | 0 |
| | Fischer 344 rats exposed by nose-only inhalation to D4 700 ppm. In ovariectomized Sprague-Dawley rats, prolactin was decreased in response to estradiol implant | (Llames, 2010) 3.4 | Model: 2 Concentrations: 3 | 61.2 |
| | (Stump, 2001). | (Stump, 2001) 3.8 | Effect: 3 Model: 3 Concentrations: 3 | 68.4 |
| | | | Effect: 2 | |
| | | | Mean | 48.6 |
| Decreased LH surge | Ovariectomized Sprague-Dawley rats exposed to 700 or 900 ppm D4 for 6 h had a downward shift in distribution of LH values, although mean values were not changed from control (Stump, 2001). Sprague-Dawley rats exposed to D4 700 or | (Quinn et al., 2007a) 3.6 | Model: 3 Concentrations: 3 Effect: 2 | 64.8 |
| | 900 ppm on diestrus 1–2 and proestrus had decreased height of LH surge at 1800 h on proestrus (Quinn et al., 2007a). There was a 3–4% decrease in terminal body weight in the treated groups. | (Stump, 2001) 3.8 | Model: 3 Concentrations: 3 Effect: 2 | 68.4 |
| | • | | Mean | 66.6 |
| Inhibition/delay of ovulation and/or inadequate corpus luteum | In Sprague-Dawley rats, 700 or 900 ppm exposure on diestrus 1 and 2 and proestrus reduced the proportion of animals that ovulated (chi-squared statistically significant by us) and the number of oocytes in the oviducts on estrus (Quinn et al., | (Meeks et al., 2007) 3.5 | Model: 3 Concentrations: 3 Effect: 3 | 94.5 |
| ruccum | 2007a). When exposure was restricted to a 6-h window, the periovulatory period was uniquely sensitive (Meeks et al., 2007). However, a decrease in oocytes in the oviducts was not seen (Quinn, 2006), although there were fewer animals. There was | | | 64.8 |
| | an increase in the number of animals with 5-day cycles (more time in diestrus) after 35 days of D4 inhalation treatment at 700 ppm. Also, an increase in resorptions seen in (Meeks et al., 2007) with exposure form 3 days before until 3 days after mating is | (Quinn, 2006) | Model: 3 Concentrations: 2 Effect: 0 | 0 |
| | not explained and might be spurious given lack of confirmation in longer exposures that also included this time period. | | Mean | 53.1 × 0.33 (late KE) = 17.7 |
| Decreased mating, fertility | Decrease in number of pups born in 2-generation study (Siddiqui et al., 2007), decrease in fertility in rats exposure for 6 h on the day prior to mating (Meeks et al., 2007). | (Meeks et al., 2007) 3.5 | Model: 3 Concentrations: 3 Effect: 3 | 94.5 |
| | | (Siddiqui et al., 2007) 3.7 | Model: 3 Concentrations: 3 Effect: 3 | 99.9 |
| Total mean scores for mode | of action | | Mean | 97.2 × 0.33 (late KE) = 32.1 176.4 |
| Maximum score achievable Percent of maximum | or action | | | 396 44% |

The exposure level at which there are adverse effects of D4 treatment in female reproduction in rats (500 and 700 ppm) could also be used to assess the relevance of this finding for human risk assessment. However, effective exposures inducing effects and their dose-response are often ignored in simplistic hazard identification schemes such as C&L. ECHA guidance (ECHA, 2015) and recent OSHA (US-OSHA, 2016) guidance for assessing carcinogenicity for C&L does, however, provide a list of factors that can be viewed as either increasing or decreasing the level of concern for human reproductive toxicity and carcinogenicity. One of these factors is "the possibility of a confounding effect of excessive toxicity at test doses". The reproductive effects following D4 exposure were only seen at the two highest dose levels (500 and 700 ppm). It is possible that these doses may have exceeded the rat

physiological capacity to handle the chemical thereby calling into question the relevance of this effect in humans at dose levels so unrealistic compared to human exposures.

3.3. Benign uterine tumors after D4 treatment

In a two-year bioassay with Fischer344 rats, inhalation of D4 at 700 ppm was associated with an increase in cystic endometrial hyperplasia and uterine adenomas. While these are benign findings, the observation of such changes might still be considered for human health risk assessments and/or C&L. An evaluation of potential mode of actions for uterine tumor formation by D4 was conducted previously but without application of a QWoE methodology (Dekant et al., 2017). The previous evaluation identified

Table 9Scoring for quality and relevance/strength for the sequence of individual steps for an estrogenic mode of action for inhibition of ovulation in female rats exposed by inhalation to D4 at 700 ppm. MIE = molecular initiating event, KE = key event.

| Key steps in mode of action | Data support | Quality scores | Relevance/ strength of effects scores | Weighted scores for MIE/KE |
|---|--|---|---|------------------------------------|
| Binding of D4 to an estrogen receptor | Cell-free systems demonstrate displacement of 17 β -estradiol from estrogen receptor- α by D4 (He et al., 2003; Quinn et al., 2007b) | (He et al., 2003) 0.9 | Concentrations: 1 | 1.8 |
| | | (Quinn et al., 2007b) 2.33 | Effect: 2 Model: 3 Concentrations: 1 Effect: 2 | 13.8 |
| | | | Mean | 7.8 |
| Activation of downstream elements | D4 shows estrogenic activity in transactivation assay and uterotrophic assays (He et al., 2003; McKim et al., 2001; Quinn et al., 2007b; Turck, 1999). Potency is several orders of magnitude lower than ethinyl estradiol or | 2007b) | Model: 3 Concentrations: 1 | 28.8 |
| | diethylstilbestrol. Uterotrophic assay negative in one study of inadequate quality (Lee et al., 2015). | 2.33 (He et al., 2003) 2.2 | Effect: 3 Model: 3 Concentrations: 1 | 6.6 |
| | | | Effect: 1 | |
| | | (McKim et al., 2001) | Model: 3 Concentrations: 1 | 30.6 |
| | | 3.4 | Effect: 3 Model: 3 | 0 |
| | | (Lee et al., 2015) | Concentrations: 1 | U |
| | | 2.1 | Effect:0 Mean | 16.5 |
| Negative estrogenic feedback on hypothalamic kisspeptin or GnRH systems and/or on pituitary gonadotropins | D4 does not suppress rat hypothalamic GnRH in vitro (Meeker, 2009). | 2.8 | Model: 1 Concentrations: | 0 |
| nhibition/delay of ovulation | In SD rats, 700 or 900 ppm exposure on diestrus 1 and 2 and proestrus reduced the proportion of animals that ovulated and the number of oocytes in the oviducts on estrus (Quinn et al., 2007a). When exposure was restricted | | Effect: 0 Model: 3 Concentrations: 3 | 94.5 |
| | to a 6-h window, the periovulatory period was uniquely sensitive (Meeks et al., 2007). However, a decrease in oocytes in the oviducts was not seen, although there were fewer animals. There was an increase in the number of | 3.5 (Quinn et al., | Effect: 3 Model: 3 Concentrations: | 64.8 |
| | animals with 5-day cycles (more time in diestrus) after 35 days of D4 inhalation treatment at 700 ppm. Also, an increase in resorptions seen in (Meeks et al., 2007) with exposure form 3 days before until 3 days after mating is not explained and might be spurious given lack of confirmation in longer exposures that also included this time period. | 2007a) 3.6 | 3 Effect: 2 | |
| | | (Quinn, 2006) | Model: 3 Concentrations: 2 | 0 |
| | | | Effect: 0 Mean | 53.1 × 0.33 (late |
| Decreased mating, fertility | Decrease in number of pups born in 2-generation study (Siddiqui et al., 2007), decrease in fertility in rats exposure for 6 h on the day prior to mating | | Model: 3 Concentrations: | KE) = 17.2 94.5 |
| | (Meeks et al., 2007). | 2007) 3.5 (Siddiqui et al., 2007) | Effect: 3 | 99.9 |
| | | 3.7 | Effect: 3 Mean | 97.2 × 0.33 |
| Total score for mode of action Max. score achievable | | | | (late KE) = 32.1 73.9 396 |
| Percent of maximum | | | | 18.7% |

three possible modes of action for which there were adequate data. The experimental support for these mode of actions, including dopaminergic agonism or dopamine agonist-like activity of D4, D4 estrogenicity, and D4 mutagenicity, is now subjected to the QWoE analysis. The first two of these mode of action are similar but not identical to the two mode of actions considered for female reproductive effects of D4.

The three competing possible mode of actions are presented in

Tables 11—13 with the quality and relevance/strength of effects scoring of the underlying studies detailed in the Supplemental Material. Some of the scores allocated here differ from scores used for the modes of action for female reproduction due to the use of different strains of rats; the reproduction studies were performed with Sprague-Dawley rats and the two-year bioassay was performed with Fischer-344 rats. Because rat strains may differ in the sensitivity of response to D4, use of the strain in which the apical

Table 10

Human relevance scoring for a dopamine mode of action regarding female fertility for D4. 0, not relevant since specific step in mode of action is not possible in humans due to species differences in biochemistry/physiology/anatomy; 1, step possible based on human biochemistry/physiology/anatomy. If an early step in a mode of action is scored as 0, further downstream steps are not possible and the mode of action is therefore not relevant in humans.

| Key steps in mode of action | Data support | Possible in humans |
|---|---|--------------------|
| Increased dopamine activity | There are no data on D4 and dopamine in humans, but it is theoretically possible that a chemical exposure could increase dopamine activity, as do some pharmaceutical products. | 1 |
| Decreased prolactin | An increase in dopaminergic activity will decrease prolactin in humans | 1 |
| Decreased LH surge | A decrease in prolactin is not associated with a decrease in LH surge in humans. | 0 |
| Inhibition/delay of ovulation and/or inadequate corpus luteum | A sufficient decrease in LH surge will inhibit or delay ovulation in humans. However, control of LH is very different in rodents compared to primates and humans. | 1 |
| Decreased mating, fertility | Inhibition of ovulation will decrease fertility in humans. | 1 |
| Total | | 0 |

endpoint was identified (fertility in Sprague-Dawley, uterine tumors in Fischer-344) is considered to provide the most relevant information for support of a mode of action. In the estrogenicity assays, the strain of rat used would be expected to be less important, and scores are largely unchanged (OECD, 2007).

Although the percentage of the maximum possible score was comparatively low for all modes of action evaluated, the dopamine activity mode of action is the best supported of the three proposed reaching a score of 48% of the maximal score achievable (Table 11). The scores indicating only weak to moderate support are mainly driven by inconsistent datasets for the 2nd key step, decreased prolactin, and a reduced weight for an important study assigned to a late key event (Jean and Plotzke, 2017; Sloter, 2015).

An estrogen mode of action for induction of uterine tumors by D4 (Table 12) received considerably less support (22% of max. achievable score). The low score is mainly due to the very low estrogenic potency of D4 resulting in low scores for concentrations applied in the relevance/strength of effects scoring.

For a mode of action involving DNA-damage by D4 and induction of mutations to account for the induction of uterine tumors, the consistent absence of genotoxicity of D4 clearly drives the low score received (Table 13). The consistently negative genotoxicity studies with D4 result in a score of 0 for the molecular initiating even and the 1st key event. The overall score is due to late key events that are common to all three modes of actions.

Because the dopamine activity mode of action is best supported for development of uterine lesions after D4 inhalation in rats, it is taken forward to the assessment of human relevance (Table 14). When evaluating human relevance of the molecular initiating/key events, the chain of key steps is again broken at key step #3, decreased LH surge, due to the absence of an association between a decrease in prolactin and the LH surge in humans. Therefore, the dopamine activity mode of action for proliferative endometrial lesions is not relevant to humans, based again on lack of a role for prolactin in human ovulatory function. As in the discussion of the relevance of the rat reproductive effect, the exposure level at which there are adverse effects of D4 treatment in female reproduction in rats (700 ppm) could also be used to assess the relevance of this finding for human risk assessment. However, effective exposures inducing effects and their dose-response are ignored in simplistic hazard identification schemes such as C&L and we will not further discuss exposure level here.

3.4. Male developmental toxicity of diethylhexyl phthalate

In case 4, to demonstrate a quantitative WoE that supports human risk assessment, the impairment of male genital development by di-(2-ethylhexyl) phthalate (DEHP) in fetal and neonatal rats is subjected to the QWoE-procedure. Although there is evidence for other developmental alterations with DEHP treatment,

and species other than rats are also sensitive, we have restricted our discussion due to the very large literature on the reproductive and developmental effects of this compound. Even within this restricted data set, we were selective in the papers that were included, bringing forward the studies that most clearly delineated the mode of action.

Euling et al. (2013) presented a qualitative WoE evaluation of the related phthalate diester dibutyl phthalate for which there are similar toxicogenomic considerations. Although the Euling et al. presentation was not quantitative, a quantitative approach could be supported by their analysis, and the proposed mode of action is similar to that proposed for DEHP in rats. Table 15 summarizes the mode of action for the anti-androgenic effects of DEHP on fetal and very young postnatal rats. A quantitative assessment of the underlying literature is summarized in the supplemental tables. Only weak support for this mode of action can be derived from the analysis, likely due to the complexity of the studies addressing the different endpoints and the large number of studies available.

Table 16 presents the human relevance scoring for this mode of action. Although the second step in the mode of action has not been evaluated in humans, it remains possible and the mode of action therefore is considered relevant to human risk assessment. However, a risk assessment must consider human exposures compared to the exposures in the experimental animal studies which are orders of magnitude above measured human exposures. Because the mode of action in rats (and other species) relies on conversion to the monoester, the kinetics of human compared to rat intestinal lipases is another factor to be considered in conducting the human risk assessment for the oral route of exposure. The fact that we have not proceeded beyond a determination that the rat data are relevant to human risk assessment does not mean that there is not considerable additional work to do in completing the risk assessment.

4. Discussion

4.1. General aspects

Criteria for C&L are hazard-based and unjustified from a scientific viewpoint (Barlow, 2016), but hazard assessment is a regulatory requirement for most uses of chemicals. Hazard assessment may be hampered by inconsistent explanations of the results of animal toxicity studies between regulators and disagreements on the relevance for humans of adverse effects in experimental animals (Golden et al., 2003; Ruden, 2001a, 2001b). Thus, a more harmonized framework based on current scientific understanding for hazard assessment and issues of extrapolation from animals to humans is needed (Schreider et al., 2010). While weight of evidence approaches are increasingly recognized, the advantage of QWoE methodology is that it provides a transparent numerical

Table 11Scoring for quality and relevance/strength for the sequence of individual steps for a dopamine agonism mode of action for uterine tumors in female rats exposed by inhalation to D4 at 700 ppm for 24 months. MIE = molecular initiating event, KE = key event.

| Key steps in mode of action | Data support | Quality scores (from supplemental tables) | Relevance/strength of evidence score | Weighted score for MIE/KE |
|--------------------------------------|---|---|--------------------------------------|-------------------------------------|
| Increased dopamine activity | MMQ cells (a rat pituitary tumor cell line) | (Domoradzki, 2011) | Model: 2 | 18.4 |
| | produced less prolactin after exposure to D4 (Jean, | 2.3 | Concentrations: 2 | |
| | 2005) and had decreased forskolin-stimulated | | Effect: 2 | |
| | cyclic AMP at D4 concentrations \geq 25 μ M without | (Jean, 2005) | Model: 2 | 4.4 |
| | cytotoxicity (Domoradzki, 2011). Not mediated by | 2.2 | Concentrations: 1 | |
| | D2 receptor (not blocked by raclopiride) or G- | | Effect: 1 | |
| | protein (not blocked by pertussin toxin) | | Mean | 11.4 |
| | (Domoradzki, 2011). | | | |
| Decreased prolactin | Sprague-Dawley rats exposed to D4 700 or | (Quinn et al., 2007a) | Model: 3 | 64.8 |
| | 900 ppm on diestrus 1-2 and proestrus had | 3.6 | Concentrations: 3 | |
| | decreased prolactin at 1400 h on proestrus (Quinn | | Effect: 2 | |
| | et al., 2007a). This effect was attributable to the | (Elias, 2010) | Model: 3 | 0 |
| | non-ovulatory animals. In another study, prolactin | 3.6 | Concentrations: 3 | |
| | was decreased 18 h but not 8 h after inhalation of | | Effect: 0 | |
| | D4 700 ppm in Fischer 344 rats that had been | (Llames, 2010) | Model: 2 | 21.6 |
| | dopamine-depleted by the administration of | 3.6 | Concentrations: 3 | |
| | reserpine (Llames, 2010). In ovariectomized | | Effect: 1 | |
| | Sprague-Dawley rats, prolactin was decreased in | (Stump, 2001) | Model:2 | 44.4 |
| | response to estradiol implant (Stump, 2001). | 3.7 | Concentrations: 3 | |
| | However, (Elias, 2010) did not show a decrease in | 3.7 | Effect: 2 | |
| | prolactin in 20-month-old female Fischer 344 rats | (Sloter, 2015) | Model: 3 | 0 |
| | exposed by nose-only inhalation to D4 700 ppm | 3.8 | Concentrations: 3 | o . |
| | and (Sloter, 2015) did not show a decrease in | 5.0 | Effect: 0 | |
| | prolactin in aged Fischer-344 rats exposed to D4. | | Mean | 32.7 |
| Decreased LH surge | Ovariectomized Sprague-Dawley rats exposed to | (Quinn et al., 2007a) | Model: 3 | 64.8 |
| Decreased LH Surge | 700 or 900 ppm D4 for 6 h had a downward shift in | 3.6 | Concentrations: 3 | 04.8 |
| | distribution of LH values, although mean values | 5.0 | Effect: 2 | |
| | were not changed from control (Stump, 2001). | (Stump, 2001) | Model: 2 | 44.4 |
| | , , , | (Stump, 2001) 3.7 | | 44.4 |
| | Sprague-Dawley rats exposed to D4 700 or | 3.7 | Concentrations: 3 | |
| | 900 ppm on diestrus 1–2 and proestrus had | | Effect: 2 | E4.C |
| | decreased height of LH surge at 1800 h on | | Mean | 54.6 |
| Totallate a / delena of annulation | proestrus (Quinn et al., 2007a). | (Maralan et al. 2007) | NA - 4-1- 2 | 01.0 |
| Inhibition/delay of ovulation, | In Sprague-Dawley rats, 700 or 900 ppm exposure | (Meeks et al., 2007) | Model: 3 | 91.8 |
| inadequate corpus luteum | on diestrus 1 and 2 and proestrus reduced the | 3.4 | Concentrations: 3 | |
| | proportion of animals that ovulated (chi-squared | (0.1 | Effect: 3 | |
| | statistically significant by us) and the number of | (Quinn et al., 2007a) | Model: 3 | 64.8 |
| | oocytes in the oviducts on estrus (Quinn et al., | 3.6 | Concentrations: 3 | |
| | 2007a). When exposure was restricted to a 6-h | | Effect: 2 | |
| | window, the periovulatory period was uniquely | | Mean | 78.3×0.33 (late KE) = 25.8 |
| | sensitive (Meeks et al., 2007). | | | |
| Increase in estrogen-dominant | There was an increase in the number of animals | (Quinn, 2006) | Model: 3 | 32.4 |
| cycle phase/Increase in circulating | with 5-day cycles (more time in diestrus) after 35 | 3.6 | Concentrations: 3 | |
| estradiol | days of D4 inhalation treatment at 700 ppm | | Effect: 1 | |
| | (Quinn, 2006). In aged Fischer-344 rats, exposure | (Sloter, 2015) | Model: 3 | 102.6 |
| | to D4 700 ppm produced an increase in number of | 3.8 | Concentrations: 3 | |
| | days with estrogenic vaginal lavage, an increase in | | Effect: 3 | |
| | serum estradiol, and an increase in estrogen/ | | Mean | 67.5×0.33 (late KE) = 22.3 |
| | progesterone ratio (Sloter, 2015). | | | |
| Increase in | A 24-month inhalation study showed an increase | (Batelle-Lee, 2004) | Model: 3 | $63 \times 0.33 (late KE) = 20.8$ |
| estrogen-dependent | in endometrial hyperplasia at 700 ppm and a | 3.5 | Concentrations: 3 | |
| endometrial lesions | statistically significant trend for endometrial | | Effect: 2 | |
| | adenoma, | | | |
| Total mean scores for mode of action | | | | 167.6 |
| Maximum score achievable | | | | 432 |
| Percent of maximum | | | | 38.8% |

Table 12Scoring for quality and relevance/strength for the sequence of individual steps for an estrogenic mode of action for uterine tumors in female rats exposed by inhalation to D4 at 700 ppm for 24 months. MIE = molecular initiating event, KE = key event.

| Key steps in mode of action | Data support | Quality scores (from supplemental tables) | , | Weighted score for MIE/KE |
|--|--|---|--|--|
| Binding of D4 to an estrogen receptor | Cell-free systems demonstrate displacement of 17β -estradiol from estrogen receptor- α by (He et al., 2003; Quinn et al., 2007b) | (He et al., 2003) 0.9 | Model: 1 Concentrations: 1 Effect: 2 | 1.8 |
| | | (Quinn et al., 2007b) 2.33 | Model: 3 Concentrations: 1 Effect: 2 | 13.8 |
| | | | Mean | 7.8 |
| Activation of downstrear elements | n D4 shows estrogenic activity in transactivation assay and uterotrophic assays (He et al., 2003; McKim et al., 2001; Quinn et al., 2007b; Turck, 1999). Potency is several orders of magnitude lower than ethinyl estradiol or diethylstilbestrol. Uterotrophic assay | | Model: 3 Concentrations: 1 Effect: 3 | 28.8 |
| | negative in one study of inadequate quality (Lee et al., 2015). | (He et al., 2003) 2.2 | Model: 3 Concentrations: 1 Effect: 1 | 6.6 |
| | | (McKim et al., 2001) 3.4 | Model: 3 Concentrations: 1 Effect: 3 | 30.6 |
| | | (Lee et al., 2015) 2.1 | Model: 3 Concentrations: 1 Effect: 0 | 0 |
| Increase in estrogen- dependent endometrial lesions | A 24-month inhalation study showed an increase in endometrial hyperplasia at 700 ppm and a statistically significant trend for endometrial adenoma, | (Batelle-Lee, 2004) 3.5 | Mean Model: 3 Concentrations: 3 Effect: 2 | 16.5 63 × 0.33 (late KE) = 20.8 |
| Total mean scores for n Maximum score achieve Percent of maximum | | | Litett, Z | 45.1 252 17.9% |

Table 13Scoring for quality and relevance/strength for the sequence of individual steps for a mutagenic mode of action for uterine tumors in female rats exposed by inhalation to D4 at 700 ppm for 24 months. MIE = molecular initiating event, KE = key event.

| Essential step in mode of action | Data support | Quality scores for key studies | Relevance/strength of evidence score evidence | Weighted total score for MIE/KE |
|--|---|--------------------------------|---|---------------------------------|
| DNA reactivity leading to mutation | All genotoxicity studies are consistently negative, | 3.8 | Model system 3 concentrations applied 3 strength of effect: 0 | 0 |
| Insufficient repair of mutations | No data | | | 0 |
| Perturbation of cell growth and survival | No data | | | 0 |
| Cell proliferation and clonal expansion of neoplastic foci | Uterine hyperplasia in 24-month oncogenicity study | 4 | Model system 3 concentrations applied 3 Strength of effects 2 | 72 × 0.33 (late KE) = 23.76 |
| Uterine tumors | Tumors at 700 ppm in 24-month oncogenicity study | 4 | Model system 3 concentrations applied 3 strength of effect 2 | 72 × 0.33 (late KE) = 23.76 |
| Total mean scores for Mode of action Maximum score achievable Percent of maximum | | | | 47.52 395.3 12% |

Table 14

Human relevance scoring for a dopamine mode of action regarding uterine tumors in rats after inhalation of D4. (0, not relevant since specific step in mode of action is not possible in humans due to species differences in biochemistry/physiology/anatomy; 1, step possible based on human biochemistry/physiology/anatomy). If an early step is scored as 0, further downstream steps are not possible and the mode of action is therefore not relevant in humans.

| Key steps in mode of action | Data support | Possible in humans |
|---|---|-----------------------|
| Increased dopamine activity | There are no data on D4 and dopamine in humans, but it is theoretically possible that a chemical exposure could increase dopamine activity, as do some pharmaceutical products. | 1 |
| Decreased prolactin | An increase in dopaminergic activity will decrease prolactin in humans | 1 |
| Decreased LH surge | A decrease in prolactin is not associated with a decrease in LH surge in humans. | 0 |
| Inhibition/delay of ovulation, inadequate corpus luteum | A sufficient decrease in LH surge will inhibit or delay ovulation in humans. | 1 |
| Increase in estrogen-dominant cycle phase/ Increase in circulating estradiol | Inhibition or delay of ovulation in humans will increase exposure to endogenous estrogens. | 1 |
| Increase in estrogen-dependent endometrial lesions | Increase in exposure to endogenous estrogens in humans will increase estrogen-dependent endometrial lesions. | 1 |
| Total | | 0 |

Table 15
Scoring for quality and relevance/strength for the sequence of individual steps for the anti-androgenic mode of action for developmental toxicity in male rats exposed to DEHP.
MIE = molecular initiating event, KE = key event.

| Key steps in mode of action | Data support | Quality scores | Relevance/ strength of effects scores | Weighted scores for MIE/KE |
|---|---|-------------------------------------|--|------------------------------------|
| Conversion to the monoester (MEHP) | Pregnant Sprague-Dawley rats convert DEHP to MEHP (Kessler et al., 2004) | (Kessler et al., 2004) 2.9 | Model: 3 Concentrations: 3 | 26.1 |
| Decreased activity of steroidogenic acute regulatory (StAR) protein and 5α-reductase activity | (Borch et al., 2006) reported treatment of pregnant Wistar rats with DEHP at 300 mg/kg/day to decrease mRNA and protein for StAR and other steroidogenic enzymes in fetal Leydig cells. | (Borch et al., | Effect: N/A | 57.6 |
| reductase activity | (Svechnikov et al., 2008) showed decreased activity of StAR in Leydig cells cultured from mature and immature rats and decreased 5α -reductase activity in Leydig cells cultured from immature but not mature rats. | (Svechnikov et al., 2008) | Effect: 2 Model: 1 | 19.2 |
| | (Kariyazono et al., 2015) treated gestation-day 15 Wistar rats by gavage with DEHP and showed a decrease of StAR mRNA in fetal testes at a maternal dose level of 100 mg/kg. | (Kariyazono et al., 2015) | Effect: 3 Model: 3 | 18.9 |
| Decreased transport of cholesterol across the mitochondrial membrane | (Svechnikov et al., 2008) showed decreased cholesterol transport associated with the decrease in StAR in rat Leydig cells | (Svechnikov et al., 2008) 2.2 | | 31.9 19.2 |
| Decreased synthesis of testosterone and dihydrotestosterone | (Akingbemi et al., 2001) showed a decrease in serum testosterone in juvenile Long-Evans rats after treatment during pregnancy of their dams with DEHP 100 mg/kg/day. | (Akingbemi et al., 2001) 2.9 | Effect: 3 Model: 3 Concentrations: 3 | 54.8 |
| | Borch et al., 2006 reported a decrease in fetal testis testosterone concentration and testosterone production in Wistar rats after treatment of their dams with DEHP 300 mg/gk/day. (Svechnikov et al., 2008) showed a decrease in hCG-stimulated testosterone | Borch et al., 2006 3.2 | Effect: 3 Model: 3 Concentrations: | 57.6 |
| | production by cultured Leydig cells from adult and immature rats. (Culty et al., 2008) reported a decrease in fetal testis basal production of testerosterone and dihydrotestosterone after treatment of pregnant Sprague-Daley rats with 234 mg/kg/day (testosterone) or 117 mg/kg/day (DHT). | (Svechnikov et al., 2008) 2.2 | Effect: 2 Model:1 Concentrations: | 19.2 |
| | | (Culty et al., 2008) 2.6 | Effect: 3 Model: 3 Concentrations: 3 Effect:: 3 | 23.4 |
| Abnormal genital development | (Moore et al., 2001) reported that treating pregnant and lactating Sprague- Dawley rats with DEHP produced male offspring with reduced anogenital distance, retained nipples and areolae, agenesis of the anterior prostate, | (Moore et al., 2001) 3.1 | Mean Model: 3 Concentrations: 3 | 38.8 27.9 |
| | undescended testes, incomplete preputial separation, and reduced weights of testes and testosterone-dependent sex organs beginning at 375 mg/kg/day. A multigeneration continuous breeding study by NTP (NTP, 2005) using Sprague-Dawley rats reported reduced male anogenital distance and delayed | (NTP, 2005) 3.9 | Concentrations: 3 | 35.1 |
| | preputial separation and testis descent. Epididymides and testes were smaller. Effects were seen at a dietary level of 7500 ppm and higher, corresponding to a dose level of about 400–600 mg/kg/day. | | Effect: 3 Mean | 31.5 × 0.33 (late KE) = 10.5 |
| Total score for mode of action Max. score achievable Percent of maximum | | | | 126.5 467.64 27% |

assessment of study quality and of reported information that is transparent, consistent, and scientifically robust.

The QWoE proposed uses the criteria in the CLP Regulation (CLP Regulation 1272/2008 1272/2008 part 3.7) as a basis for conclusions regarding C&L with a specific focus on the aspect "mode of action differences are so marked that it is certain that hazardous effects seen in the animal model will not be seen in man" (3.7.2.3.3). Even in the presence of adverse effects of a chemical of interest in an appropriate animal study, classification is not mandated under such circumstances. However, there are widely different opinions on human relevance even for well-characterized mode of actions that have good support for the absence of human relevance (Johnson et al., 2012; Mehlman, 2000; Melnick et al., 2013; van den

Driesche et al., 2015). The QWoE methodology developed here may help to reduce controversial discussions in this area due to the transparent approach. A transparent QWoE-approach may also be used to reduce disagreements on the outputs of hazard assessment and risk characterization.

4.2. Advantages of QWoE

One of the challenges in the development of the QWoE methodology is that each of the endpoints addressed and the effects reported need to be integrated. The approach described here offers several advantages. Scores for each mode of action can be compared based on well-defined criteria to define experimental

Table 16Human relevance scoring for interference with testosterone and dihydrotestosterone synthesis as the mode of action regarding developmental toxicity of DEHP.

| Key steps in mode of action | Data support | Possible in human |
|---|--|----------------------|
| Conversion to the monoester (MEHP) | After ingestion of labelled DEHP or intravenous infusion of DEHP, MEHP and other metabolites were present in urine (Koch et al., 2005a,b). | 1 |
| Decreased activity of steroidogenic acute regulatory (StAR) protein and 5α-reductase activity | There are no data on this step in humans; however, it remains possible. In such cases, human relevance has to be assumed | 1 |
| Decreased transport of cholesterol across the mitochondrial membrane | The activity of StAR in humans is similar to that in rats in facilitating transport of cholesterol across the mitochondrial membrane as an early step in steroid biosynthesis. | 1 |
| Decreased synthesis of testosterone and dihydrotestosterone | A decrease in cholesterol transport across the mitochondrial membrane will decrease steroid biosynthesis in humans, and a decrease in 5α -reductase activity will decrease dihydrotestosterone. | 1 |
| Abnormal genital development | The human embryo relies on adequate exposure to testosterone and dihydrotestosterone to complete development of the male urethra and external genitalia. | 1 |
| Total | | 1 |

support for all steps. Additionally, quality assessment covers all aspects that need to be considered and is based on best practice thus giving a more objective assessment. Quality criteria may help design of experiments optimized to assess molecular initiating/key events, and the detailed evaluation of support for molecular initiating/key events may indicate missing links in a mode of action that need to be addressed by specific experiments. The basis for scoring is clearly defined, can be adapted to changes in scientific understanding, and is broadly applicable. Scoring for relevance/effects provides a transparent approach to integrating complex and contradictory observations into a numerical score that can be used as a basis for conclusions on experimental support for a mode of action. As shown by the outcome of case study #1, targeted experiments to support a well-defined mode of action in a relevant system will results in a high confidence in the mode of action in animals.

4.3. Challenges

Scoring for widely different experimental approaches (specifically when assessing the ever-increasing number of in vitro systems), requires complex knowledge of advantages and pitfalls of such systems and may need input from specialized experts familiar with limitations of the systems and issues with dosimetry. Assignment of experimental data to support a key event is clear when a target experiment on one specific molecular initiating/key events is conducted and a simple system used. However, some experiments assess outcomes under more complex circumstances and use endpoints that may be considered as early or late key events. An example is the study of Jean and Plotzke, 2017, which assessed estrogen-dominant days during the rat estrous cycle and estrogen:progesterone ratio in aged Fischer 344 rats, important evidence to support a dopaminergic mode of action in the uterine tumor evaluation (Table 11). This experiment was carefully designed to assess a key step in the proposed mode of action, and it was given a high score for quality and relevance. Because it addressed a later step in the mode of action, however, its importance was reduced by 0.33, which reduced its contribution to the evaluation. This indicates that a lower score for late events may not be always appropriate and expert judgement may be required to justify a deviation from the general approach.

In this QWoE, only positive scores were applied. Negative scores for strength of effects may need to be included to balance positive and negative studies (Fenner-Crisp and Dellarco, 2016) since larger data sets on potential key events often contain some contradictory findings. Negative scores distinguish evidence of the absence of a response from the absence of evidence (score of 0). In case of negative scores for the criterion "strength of evidence," a negative overall score (clear evidence that the endpoint is not affected) in the model for a molecular initiating/key event will be obtained and

the final calculation will generate a negative number suggesting that the mode of action is unlikely. However, the QWoE here applies scores of zero for both inconclusive experiments and for databases that can be interpreted to support absence of an effect. The exclusion of negative scores reflects our lack of confidence that insufficient sensitivity or model limitations may prevent detection of an effect. In this QWoE, scores of zero for molecular initiating/key events should be interpreted as absence of support.

The criteria applied focus on the plausibility of results to support a key event and consistency with a hypothesis. This is a challenge for QWoE since the rules for the scoring process may not adequately capture the complexities of the interpretation of evidence. This will still require significant expertise and further development of such schemes based on experience will be required. A possible solution for data-rich chemicals may be to perform a QWoE only on high-quality studies with clear results and little confounding to avoid "dilution" of a score for a key event by low quality studies in systems with limited relevance.

While the use of ranking systems for the quality and strength/ relevance elements is reasonable, the treatment of these ranks as equivalent across domains and the arithmetic manipulation (addition, multiplication) of the ranks may be suboptimal. These ranks highlight the difference between high and low quality datasets, and studies that received high scores appear to have high scores in multiple domains. The comparison of mode of action with small differences in overall scores might, at least in theory, produce results different from those reached through scientific judgment.

Scoring of human relevance of a specific mode of action also is challenging. The procedure selected, for reasons of simplicity, applies a simple yes/no response regarding plausibility of a key step of the mode of action in humans without directly considering data support. An expansion of the assessment integrating data support for presence/absence of a key step in humans is may be developed applying a range of scores. However, this may represent a very tedious process that requires scoring of a large number of studies including those assessing basic human physiology performed decades ago often using simple experimental designs.

Quantitative aspects such as area-under-the curve (AUC) and non-linear toxicokinetics that may be important for the expression of toxicity after long term treatment also may require specific considerations in a QWoE, specifically if adverse effects occur after compensatory mechanisms are overwhelmed or only at very high doses may also need specific approaches.

4.4. Results of the case studies

The case study for MTBE yielded a very clear outcome due to the availability of a well characterized mode of action with molecular initiating/key events that can be experimentally addressed with

high precision and that are based on very well-evaluated outcomes that have few confounders. The QWoE of the available datasets shows that the α_{2u} -globulin mode of action receives a very high confidence score based on high quality experiments with consistent outcomes in relevant systems applying relevant concentrations and addressing all plausible key events. The outcome is consistent with regulatory practice that states that when these key events are adequately supported by data, renal tumors induced by this mode of action have no human relevance and should not be used as endpoints for assessing human risk or for classification and labelling.

The D4 studies are more complicated since some endpoints may be confounded by experimental conditions and good in vitro models for some molecular initiating/key events are not available. The data set is further complicated by use of Sprague-Dawley rats in the reproductive studies and F344 rats in the carcinogenicity study. Dekant et al. (2017) proposed that the uterine effects seen in F344 rats following D4 exposure were due to alteration of pituitary control of the estrous cycle as a result of dopamine agonist-like activity. Although cycle disruption was demonstrated in F344 rats, it was not possible to demonstrate LH modulation as a key event because the F344 rat is highly sensitive to stress that can be induced with such a complex study. If the key event of LH modulation accounts for the reproductive effects and altered cycles in F344 rats leading to the uterine effects, it is possible that the effects in the two studies are linked to the same molecular initiating event. Lastly, there may be kinetic differences between Sprague-Dawley and F344 rats in how D4 is processed following exposure in these two strains of rats. The uterine effects were only seen following exposure to the highest exposure concentration of D4 (700 ppm) and the reproductive effects were only seen at the top two dose levels (500 and 700 ppm). At air concentrations of D4 greater than ~300 ppm (Sarangapani et al., 2002) there was an apparent saturation of liver enzymes with subsequent decreasing liver metabolism suggesting that the high doses of D4 may exceed the physiological ability of the rat to handle the chemical. A similar assessment of the kinetics in Sprague Dawley rats could be done to assess if this kinetically maximum tolerated dose is in the same range or lower. Understanding this possible influence of strain would add to the overall weight of evidence for assessing relevance of the observed effects.

The narrative assessment concluded that there is adequate support for a dopamine-like mode of action regarding benign uterine lesions induced by D4. The QWoE produced only low to moderate support, albeit better than the competitive mode of actions involving estrogenicity and genotoxicity. Low to moderate support is due to the presence of inconsistent datasets for some key events in the dopamine mode of action. The evaluation might have been clearer had the Jean and Plotzke, 2017 study been conducted to evaluate an early key event, raising the question of whether the position of the key event in the mode of action should be characterized as early or late as opposed to specific or nonspecific. Late key events are generally applicable to more than one mode of action and are, therefore, regarded as nonspecific. In the dopaminergic mode of action for uterine tumors, the Jean and Plotzke, 2017 study supports a key event that, although late in the process, is highly specific to the mode of action and absent from the competing mode of actions. Fully weighting of this key event might be appropriate based on its specificity. Dekant et al. (2017) acknowledged that it is likely that cycle disruption occurred over time in F344 females exposed to D4 due to either an inhibition by D4 of pituitary prolactin production (via dopamine agonist like activity and/or through modulation of the LH surge by another nonspecified molecular initiating event) leading to an increased endogenous estrogen signal to the uterus. However, they concluded that neither mechanism would be relevant to human risk due to differences between rat and human in pituitary control of the female reproductive cycle (Klaunig et al., 2016; Plant, 2012).

The DEHP case study was included to demonstrate a mode of action that is relevant to human risk assessment, although differences in effective dose levels in rats and human exposures remain to be evaluated in the risk assessment. There is a large literature on DEHP effects on the development of androgen-responsive tissues in a number of species. Our selective citation of some of this literature and restriction to the fetal and neonatal rat was intended only to illustrate the application of the process to a data set with human relevance.

The case studies demonstrate that such a simple approach may be sufficient to come to a conclusion on human relevance. For methyl tert-butyl ether (MTBE), the molecular initiating event of the best-supported mode of action in the test species does not occur in humans, mice, or female rats since neither humans nor female rats nor mice express α_{2u} -globulin or similar proteins. Thus, the molecular initiating event represented by the binding of MTBE to α_{2u} -globulin cannot occur and the overall mode of action is not applicable in humans. In cases where chemicals induce liver tumors in rats by interaction with peroxisome proliferator activated receptor alpha (PPAR α) or the constitutive androstane receptor (CAR), both humans and rodents express the receptor indicating that the molecular initiating event of the mode of action may occur in humans (Braeuning, 2014; Corton et al., 2014; Klaunig et al., 2003; LeBaron et al., 2014). Thus, the molecular initiating event will receive a score of one in the human relevance assessment. However, there are several datasets that show that key events downstream from the molecular initiating event in both the PPARα and the CAR mode of action do not occur in human tissues resulting in a break in the mode of action chain, supporting the conclusion that both the PPAR α and the CAR mode of action for liver tumors have no relevance in humans (Braeuning, 2014; Corton et al., 2014; Klaunig et al., 2003; LeBaron et al., 2014). However, this view has been challenged (Guyton et al., 2009).

5. Conclusion

The three case studies demonstrate the utility of the developed QWoE methodology presented here to 1) assess confidence in evaluating potential mode of action (MoAs) for adverse effects observed in animal toxicity studies and 2) assess the appropriateness of the adverse effects as relevant endpoints in human health risk assessments and for classification and labeling. The method can be applied to a range of different endpoints of common concern. Moreover, it is transparent and scientifically sound and therefore less likely that it will be used as a political tool. The major challenge is to define more fully the basis for which relevance of a mode of action to humans can be discounted. OWoE-approaches should be included as part of the guidance documents for C&L and risk characterization procedures to improve the credibility of the outcome of these processes and many regulations regarding chemical safety such as EUs Equivalent Concern Regulation, the Lautenberg Chemical Safety Act, IARCs assessment of carcinogenic hazard, and risk evaluations by EFSA, ECHA and other regulatory bodies.

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Appendix A. Supplementary data

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Transparency document

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Supplemental Material

Table 1. Quality assessment of studies supporting D4 effect mediated by dopamine agonism

A. In vitro

| A. In vitro | | | | | | | | | | |
|--|--|--|--------------------------|-----------------------------------|--|-----------------|--|--|--|--|
| Study | Jean PA | 2010. Effect of cyclic siloxanes on o | dopamine receptor regula | tion of prolactin release from ra | t pituitary tumor-derived transformed cell line | es. Dow Corning | | | | |
| identification | HES Stud | HES Study Number: 9872-102 | | | | | | | | |
| Design | Model syccyclases. Positive of Concurre | Material studied: D4 99.4% purity Model system: MMQ cells, derived from a rat pituitary tumor; this cell line secretes prolactin and expresses functional dopamine D2 receptors and adenylate cyclases. Positive controls: Dopamine Concurrent controls: Not stated Treatment: D4 added to RPMI medium ay 10 μM Endpoints assessed: Blockage of maitotoxin stimulated prolactin secretion | | | | | | | | |
| Effects | | | | | e and by D4. D4 did not affect cell viability. | | | | | |
| Quality of metho | ods | 4 | 3 | 2 | 1 | 0 | | | | |
| 1. Chemical well including present contaminants Avecontamination fro equipment/dosing Control of adsorp glassware causin interference, approaution to avoid levolatiles, limit of secontrols, suitabilit duration. | ce pidance of om g solutions. tion onto g ropriate oss of solubility) imental umber of concentration, ty of exposure | D4 was well-characterized | | | Only 1 D4 concentration used. Number of replicates not given | | | | | |
| 3. Mode of applic item system (stak used, route of app dosing intervals, actual concentrat chemical of intere | oility, vehicle plication, estimation of est in medium | | | | No information on fate of chemical, no measurement of D4 concentration in medium | | | | | |
| All assessment determination of the model organism | | Cell viability assessed. | | | | | | | | |
| Suitability of sa method, sampling procedures. | g times and | | | | No justification of sampling plan | | | | | |
| 6. Suitability of bi | ochemical | | Commercial kit | | | | | | | |

| managements in alcoling | | | | alita a a satural | | | |
|---|--------------------|------------|--------------------|---|------------------------------------|-----------------------------|-------------------|
| measurements including | | | measurements, qua | ality control | | | |
| quality control | | | | not addressed | | | |
| 7. System used for | | | Well characterized | cell type | | | |
| biotransformation/cells have | | | | | | | |
| capacity to simulate relevant | | | | | | | |
| reactions that occur with | | | | | | | |
| chemical of interest in animals | | | | | | | |
| Accessibility of raw data | | | | | | Summary data reported | |
| Statistical analysis | | | | | Statistical methods not | | |
| | | | | | discussed in detail, no | | |
| | | | | | evaluation of normality of | | |
| | | | | | distributions | | |
| Relevance and strength of effects | 3 | 3 | 2 | | 1 | | |
| Concentrations applied and their rele | evance | | | | | Concentration in culture r | not characterized |
| to dose/tissue concentrations of che | mical of | | | | | with respect to in anticipa | ated in vivo |
| interest resulting in adverse effects i | in | | | | | concentrations | |
| animals | | | | | | | |
| Relevance of model system and end | dpoint | | | Adequate model but it is not clear that | | | |
| assessed to key events occurring in | | | | | mor cell represents in vivo events | | |
| animals | | | | | • | | |
| Strength of effects | | | | | | Changes observed but do | ose- and time- |
| 3 | Strongth or oncote | | | | | dependence not evaluate | |
| Scoring | | | | | | | |
| Quality of methods total | | 20 | | | | | |
| Score/domain | | 20/9 = 2.2 | | | | | |
| Strength and relevance total | | 2 | | | | | |
| Total score | | 4.4 | | | | | |

| Otrodo | D | i IV 0044 to vitro MMO coll become | | | | 1\ | |
|---|---|---|--|---|---|-----------------|--|
| Study | | ki JY. 2011. <i>In vitro</i> MMQ cell-based | | amine receptor activation by octan | netnylcyclotetrasiloxane (dź | i) and | |
| identification Design | | ylcyclopentasiloxane (D5). Study 1125 studied: D4 of ~98% purity | 000-102. | | | | |
| Design | Model system: MMQ cells, derived from a rat pituitary tumor; this cell line secretes prolactin and expresses functional dopamine D2 receptors and adecyclases. Positive controls: Forskolin (activates adylate cyclase resulting in increased cellular cAMP), quinpirole (D2-receptor agonist, attenuates forskolin-inductional dopamine D2 receptors and adecyclases. | | | | | | |
| | | in cellular cAMP), raclopiride (D2-rece | | | | | |
| | | nt controls: Dimethylsulfoxide carrier | ptor anatagornot, brooks the dollar | ny or quiripinolo, pormitting the force | skemi maadda mordadd m c | ondial of tivil | |
| | | at: D4 added to RPMI medium at 0, 25 | . 50. or 100 uM. | | | | |
| | | s assessed: Forskolin-induced increas | | | | | |
| Effects | D4 at 25, | 50, and 100 μM decreased forskolin-i | nduced intracellular accumulation | | | | |
| | | cumulation was not altered by treatme | | | | | |
| | | he forskolin dose-response curve was | | | suggesting competitive inh | ibition by D4. | |
| | | toxin did not block the activity of D4 s | | | 1 | T | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | |
| 1. Chemical well char including presence contaminants Avoidar contamination from equipment/dosing sol Control of adsorption glassware causing interference, appropri caution to avoid loss a volatiles, limit of solute. 2. General experiment design such as numb assays per dose/cond controls, suitability of duration. | nce of lutions. onto liate of collity) ntal er of centration, | | Suitable for purpose but some potentially significant limitations identified. Lower number of repeats or no positive or historic control data available. Some aspects of study duration and sampling plan are questionable. Only limited dose-response | | Reliance on supplier for information on identity and purity. Concentration of chemical of interest in medium not considered, use of media from established supplier | | |
| 3. Mode of application item system (stability, used, route of applica dosing intervals, estin actual concentration ochemical of interest in 4. All assessments in | , vehicle ation, mation of of n medium | Detailed toxicity assessment by | innited dose response | | Limited information on concentration and fate of chemical applied | | |
| determination of toxic model organism | sity to | appropriate methods in controls and exposed system, cytotoxicity of chemical of interest in system well defined and reported | | | | | |
| Suitability of sampl | ling | | | | Only few samples | | |

| method, sampling times and procedures. | | | | | | collected, no justification for sampling plan | |
|---|---|---|--|---|---|---|-------------------------|
| 6. Suitability of biochemical measurements including quality control | | | Selection not comp with best practice lin parameter consistent assessed | miting inter | | | |
| 7. System used for biotransformation/cells have capacity to simulate relevant reactions that occur with chemical of interest in animals | | | | | | | No relevant information |
| 8. Accessibility of raw data | Complete a | ccess to all raw data | | | | | |
| 9. Statistical analysis | suitable for | e statistical method analysis of endpoint. or normal distribution | | | | | |
| Relevance and strength of effe | ects | 3 | 2 | | 1 | | |
| Concentrations applied and their to dose/tissue concentrations of interest resulting in adverse effect animals | chemical of | | | Concentratione order of concentration reasonably exposure of effect | cinadequate information. cions required to induce effect are of magnitude above tons of the chemical of interest expected in tissue under conditions causing the adverse | | |
| | Relevance of model system and endpoint assessed to key events occurring in intact animals | | | limitations determined | d model system, but some regarding relevance of endpoint I for sequence of events resulting effect in vivo | | |
| Strength of effects | | | | observed, I | n endpoint or biomarker but significant changes have e or time-dependency | | |
| Scoring | | | | | | | |
| Quality of methods total | | 21 | | | | | |
| Score/domain | | 12/9 = 2.3 | | | | | |
| Strength and relevance total | | 8 | | | | | |
| Total score | | 18.4 | | | | | |

B. In vivo

| B. In VIVO | | | | | | | | | |
|--------------------------|---|---|---|-------------------------------------|-----------------------------|--------------------|--|--|--|
| Study | | Effect of octamethylcyclotetrasiloxa State | | | CAS No. 541-02-6) on circ | culating prolactin | | | |
| Identification | | levels in the aged female Fischer 344 rat. Study 11360-102. Dow Corning Report No. 2010-10000-62617 Material studied: D4 of 99.77 % purity, monitoring of air concentrations | | | | | | | |
| Design | | | | | | | | | |
| | Type/Strain of animals: female Fischer 344 rats from Taconic Farms Positive controls: Pergolide 0.2 mg/kg by gavage on days 1 and 5 Concurrent controls: air only | | | | | | | | |
| | | | | | | | | | |
| | | <i>nt controls:</i> air only <i>tes:</i> 31 animal cannulated animals, n ra | and from 0 to 10 per group, addi | tional animals avacand to DE are | not considered here | | | | |
| | | | | | | | | | |
| | Feed: Pur | nose-only inhalation exposure to 0 or 7 | oo ppin, 6 nours/day for 5 days, ve | enicle for pergolide was ethanol in | COITI OII. | | | | |
| | | nna 5002 nd age at dosing: Animals were ≥20 m | onths old at experiment start. Tree | tmont was by noso only inhalation | for D4 and by gayage for | oorgolido | | | |
| | | s assessed: Body weight, daily vaginal | | | | | | | |
| | | posure, after at least 6 hours of expos | | | osterone. Blood was sampl | ed the alternoon | | | |
| Effects | | ve control (pergolide) increased prolac | | | not altared by progesterens | was deerseed | | | |
| Ellecis | | There was an increase in estrogenic v | | | | | | | |
| | | control was all increase in estrogenic values to D4. Venous prolactin conce | | | | ation after 1 of 5 | | | |
| Quality of methods | days or ca | A | 3 | 2 | 1 1 | 0 | | | |
| Chemical well char | acterized | Highly purified, well characterized, | | | | | | | |
| including presence | 401011204 | same batch used for all studies | | | | | | | |
| contaminants | | | | | | | | | |
| 2. General experimer | ntal | | Adequate number of animals | | | | | | |
| design (number of an | | | with positive and negative | | | | | | |
| dose group, controls, | | | controls. Inhalation exposure | | | | | | |
| of study duration, hou | | | duration not well justified, | | | | | | |
| conditions) | ionig | | sampling intervals not well | | | | | | |
| oonalions) | | | justified. | | | | | | |
| 3. Assessment of pos | sible | Appropriate handling of control | | | | | | | |
| interference from stre | ss due to | animals to reproduce possible | | | | | | | |
| restraint, toxicity | | stressful conditions | | | | | | | |
| 4. Mode of application | | Used effective concentration from | | | | | | | |
| item to animals (stabi | | other reproductive studies, 700 | | | | | | | |
| vehicle used, route of | | ppm, air concentrations monitored | | | | | | | |
| administration, dosing | 9 | | | | | | | | |
| intervals) | | | | | | | | | |
| 5. Appropriate animal | l model | | Fischer 344 rats were used in | | | | | | |
| and strain selection | | | other studies, including the 2- | | | | | | |
| | | | year studies, but Sprague | | | | | | |
| | Dawley rats were used in the | | | | | | | | |
| | reproductive studies. The use | | | | | | | | |
| | | | of the aged animal makes it | | | | | | |
| | | | difficult to apply results to | | | | | | |
| 6. Suitability of sample | ina | | reproductive studies. Sampling intervals not well | | | | | | |
| measurements, samp | | | justified. | | | | | | |
| times, and procedure | | | Justineu. | | | | | | |
| umes, and procedure | ১ | | | | 1 | 1 | | | |

| 7. Suitability of biochemical | | | Based on manufact | urers | | | |
|--|-----------------------------------|--|-----------------------------------|-------|----------|---|---------------------------------|
| measurements including | | | specifications. | | | | |
| quality control | | | | | | | |
| 8. Suitability of pathological/ | | | Blinding not reporte | d | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | Raw data a | vailable in report | | | | | |
| 10. Statistical analysis | Appropriate for normal of | e methods with checks distribution | | | | | |
| Relevance and strength of effe | cts | 3 | | 2 | | 1 | |
| Concentrations applied and their to dose/tissue concentrations of cinterest resulting in adverse effect animals Relevance of model system and cassessed to key events occurring animals | chemical of tts in endpoint | Concentrations required in the range of concentrations chemical of interest reastissue under exposure control the adverse effect | ations of the sonably expected in | | | Uncertainty regarding so animal to reflect critical limitations of model syst | endpoint in vivo, |
| Strength of effects | | | | | | Changes in endpoint or observed, but no dose of and limited statistical significant significant control of the | biomarker or time dependence |
| Scoring | | | | | | | |
| Quality of methods total 35 | | <u>-</u> | | - | <u> </u> | | |
| Score/domain | Score/domain 35/10 = 3.5 | | | | | | |
| Strength and relevance total | | 3 | | | | | |
| Total score | | 10.5 | | | | | |

| Study | Llamas I 7 | Γ. 2010. <i>In vivo</i> evaluation of the impac | et of exposure/endpoint evaluation | timing on the notantial for estame | athylovelototracilovano and | 1 | | |
|--|---|--|-------------------------------------|--|------------------------------|--------------------|--|--|
| Identification | | ylcyclopentasiloxane to affect circulating | | | | | | |
| identification | | Technical Report. Study number 1125 | | -treated ferriale i isoriei i 344 fat. | bow corning ricality and b | Invironmental | | |
| Design | | tudied: D4 of 99.77 % purity, monitorin | | | | | | |
| 2 00.g.: | Type/Strain of animals: female Fischer 344 rats from Charles River Positive controls: Pergolide 0.2 mg/kg by oral gavage | | | | | | | |
| | | | | | | | | |
| | | nt controls: air only | 3. | | | | | |
| | | es: 32 ovariectomized animals and 72 | intact animals in negative control | and treatment groups; 8 animals i | n positive control group an | d 32 animals in | | |
| | | e treatment groups. | • | | | | | |
| | Vehicle: n | ose-only inhalation exposure to 0 or 70 | 00 ppm for 6 days in 4 groups of fe | emale rats (a D5-exposed group is | s not included in this summ | nary). The vehicle | | |
| | for pergoli | ide was a solution containing 10% etha | anol and 90% corn oil, and the veh | icle for reserpine was 37% DMSC |), 7% glacial acetic acid, a | and 56% reverse | | |
| | osmosis v | | | | | | | |
| | Feed: Pur | | | | | | | |
| | | nd age at dosing: Animals were 9 week | | | | | | |
| | | 8 am, repeated 24 hours later. Negati | | | | | | |
| | | ose of reserpine or vehicle, the positive | | | | de. Immediately | | |
| | | olide or vehicle administration, animals | | | | | | |
| Effects | | s assessed: Body weight gain, mortality is no mortality in the negative control gr | | | | | | |
| Ellecis | treatment | in two of the groups receiving reserpin | oup and the positive (pergolide) of | reated animals was 17.76 ug/g tr | unk blood: there was no Do | to the inhalation | | |
| | | ot exposed to D4. Prolactin concentrati | | | | | | |
| | | rolactin as expected. Reserpine treatm | | | | | | |
| | | ad a blood prolactin concentration simi | | | | | | |
| | | n prolactin. Eight hours after the end of | | | | | | |
| | | nours after the end of the inhalation pe | | | | | | |
| | lower than | n in air-treated animals. | , | · · · · · · · · · · · · · · · · · · · | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | |
| Chemical well char | racterized | Highly purified, well characterized, | | | | | | |
| including presence | | same batch used for all studies | | | | | | |
| contaminants | | | | | | | | |
| 2. General experimer | ntal | | Adequate number of animals | | | | | |
| design (number of an | | | with positive and negative | | | | | |
| dose group, controls, | | | controls. Inhalation exposure | | | | | |
| of study duration, hou | using | | duration not well justified, | | | | | |
| conditions) | | | sampling intervals not well | | | | | |
| | | | justified | | | | | |
| 3. Assessment of pos | | Appropriate use of controls to | | | | | | |
| interference from stre | ess due to | neutralize possible effects of stress | | | | | | |
| restraint, toxicity | n of toot | Ctandard avacrimental design for | | | | | | |
| 4. Mode of application item to animals (stab.) | | Standard experimental design for nose-only exposure, sampling | | | | | | |
| vehicle used, route of | | documented lack of systemic | | | | | | |
| administration, dosing | | exposure of unexposed groups | | | | | | |
| intervals) | 9 | CAPOSCIE OI GITEAPOSEG GIOGPS | | | | | | |
| 5. Appropriate anima | l model | | Fischer 344 rats were used in | | | | | |
| o., appropriate ariiria | | | 1 1001101 0 1 1 1ato word about III | l . | 1 | 1 | | |

| and strain selection | | | other studies, inclusive studies, but Sprag rats were used in reproductive studi | ue Dawley the | | | |
|---|-------------|------------------------|--|------------------|--------------------------------|---|----------------|
| 6. Suitability of sampling | | | | | Serial sampling intervals were | | |
| measurements, sampling | | | | | not justified. | | |
| times, and procedures | | | | | | | |
| 7. Suitability of biochemical | | | Based on manufa | cturers | | | |
| measurements including | | | specifications | | | | |
| quality control | | | | | | | |
| 8. Suitability of pathological/ | | | Blinding was not r | eported. | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | Report ava | ilable with raw data | | | | | |
| 10. Statistical analysis | Appropriate | e methods were used | | | | | |
| Relevance and strength of effe | | 3 | | 2 | | 1 | |
| Concentrations applied and their | r relevance | Exposure level was bas | sed on previous | | | | |
| to dose/tissue concentrations of | chemical of | findings and was appro | priate. | | | | |
| interest resulting in adverse effection animals | cts in | | | | | | |
| Relevance of model system and | endpoint | | | Use of res | erpinized animals is of | | |
| assessed to key events occurrin | g in intact | | | questional | ole relevance to human | | |
| animals | | | | assessme | nt. | | |
| Strength of effects | | | | | | Effects of treatment doo 18 hours after exposure directions, and not at 0 | e, in opposite |
| Scoring | | | | | | | |
| Quality of methods total | | 34 | | | | | |
| Score/domain | | 3.4/10 = 3.4 | | | | | |
| Strength and relevance total | | 6 | | | | | |
| Total score | | 20.4 | | _ | | | |

| | 1 | | | | | | | | |
|---|--|--|-------------------------------------|---|------------------------------|----------------------------------|--|--|--|
| Study | | G, Stump DG, Siddiqui WH, Holson JF | | | study of octamethylcyclote | trasiloxane (D ₄) in | | | |
| Identification | | s using multiple and single day exposi | ure regimens. Reprod Toxicol 23:1 | 92–201. | | | | | |
| | Also: | : fman LE. 1998. An inhalation reproductive toxicity study of octamethylcyclotetrasiloxane (D4) in female rats using multiple exposure regimens. Dow Corning | | | | | | | |
| | | | | | g multiple exposure regime | ens. Dow Corning | | | |
| Daniere | | on Health and Environmental Sciences | | J-4449U. | | | | | |
| Design | Material studied: D4 of >99% purity, monitoring of air concentrations | | | | | | | | |
| | Type/Strain of animals: SD rats from Charles River Positive controls: none | | | | | | | | |
| | | ontrois. None nt controls: air only | | | | | | | |
| | | <i>n controls.</i> all only es: 24 females per dose group expose | nd 29 days prior to mating during r | mating and until gostation day 10 | An additional 60, 60, and | 24 animale | | | |
| | | es, 24 females per dose group expose ely, were exposed to the highest dose | | | | | | | |
| | | day 3, and only on gestation days 2–5 | | | | | | | |
| | | at 700 ppm. | . Additional experiments were per | ionned to further define the sensit | ve intervals for reproductiv | e ellects of D4 | | | |
| | | whole body inhalation exposure to 0, 70 | 300, 500, or 700 ppm, 6 hours/d | ay for 5 days | | | | | |
| | Feed: Pur | | 5, 000, 000, 01 700 ppm, 0 modro, a | ay for a days | | | | | |
| | | nd age at dosing: Animals were 59–71 | days old on receipt and were accl | imated for 14 days prior to the sta | t of the experiment. Treatn | nent was by whole- | | | |
| | body inha | | aayo o.a oooo,p. aa ac | aca ici i i aayo piici ic iiic cia | | | | | |
| | | assessed: Body weight, gravid utering | e weight, number of corpora lutea, | number and location of fetuses, in | mplantation sites. | | | | |
| Effects | | d mean body weight in 700 ppm dams | | | | creased | | | |
| | | tation loss, and decreased viable fetus | | | | | | | |
| | | tation loss was from 3 days before ma | | | • | | | | |
| Quality of methods | <u> </u> | 4 | 3 | 2 | 1 | 0 | | | |
| 1. Chemical well cha | racterized | Highly purified, well characterized, | | | | - | | | |
| including presence | | same batch used for all studies | | | | | | | |
| contaminants | | | | | | | | | |
| 2. General experime | ntal | | Adequate number of animals, | | | | | | |
| design (number of ar | | | study well designed to answer | | | | | | |
| dose group, controls | | | specific question about | | | | | | |
| of study duration, ho | | | sensitive exposure period. No | | | | | | |
| conditions) | g | | positive or historical control | | | | | | |
| , | | | data | | | | | | |
| 3. Assessment of pos | ssible | Stress was evaluated using animal | | | | | | | |
| interference from stre | | weight changes and adrenal | | | | | | | |
| restraint, toxicity | | weights; appropriate controls used. | | | | | | | |
| 4. Mode of applicatio | n of test | | | D4 contamination was not | | | | | |
| item to animals (stability, | | | | | | | | | |
| | | | | assessed, possible effects of | | | | | |
| vehicle used, route o | f | | | assessed, possible effects of estrogen feed, bedding not | | | | | |
| | f | | | estrogen feed, bedding not assessed. Dose level and | | | | | |
| vehicle used, route o | f | | | estrogen feed, bedding not assessed. Dose level and route (whole body inhalation) | | | | | |
| vehicle used, route o administration, dosin intervals) | f g | | | estrogen feed, bedding not assessed. Dose level and | | | | | |
| vehicle used, route o administration, dosin intervals) 5. Appropriate anima | f g | Use of Sprague Dawley rats as in | | estrogen feed, bedding not assessed. Dose level and route (whole body inhalation) | | | | | |
| vehicle used, route o administration, dosin intervals) | f g | Use of Sprague Dawley rats as in reproductive studies was | | estrogen feed, bedding not assessed. Dose level and route (whole body inhalation) | | | | | |
| vehicle used, route o administration, dosin intervals) 5. Appropriate anima and strain selection | f g Il model | reproductive studies was appropriate. | | estrogen feed, bedding not assessed. Dose level and route (whole body inhalation) | | | | | |
| vehicle used, route o administration, dosin intervals) 5. Appropriate anima | f g Il model | reproductive studies was | | estrogen feed, bedding not assessed. Dose level and route (whole body inhalation) | | | | | |

| times, and procedures | adequate. | | | | | | |
|--------------------------------------|--------------|--|--|-----------|--|---|--|
| | | nanufacturer's | | | | | |
| measurements including specification | | ns | | | | | |
| quality control | | | | | | | |
| 8. Suitability of pathological/ | | | Blinding was not rep | oorted. | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | Accessible | in Kaufman (1998) | | | | | |
| 10. Statistical analysis | | | Adequate methods | | | | |
| | | | mention of evaluation | | | | |
| | | | normality of distribut parametric testing. | tions for | | | |
| Relevance and strength of effe | cte | 3 | parametric testing. | 2 | | 1 | |
| Concentrations applied and their | | • | I to induce offect are | | | | |
| to dose/tissue concentrations of | | Concentrations required to induce effect are in the range of concentrations of the | | | | | |
| interest resulting in adverse effect | | chemical of interest reasonably expected in | | | | | |
| animals | | tissue under exposure conditions causing | | | | | |
| | | the adverse effect | | | | | |
| Relevance of model system and | endpoint | Endpoint or biomarker is clearly compatible | | | | | |
| assessed to key events occurring | g in intact | with key event in vivo in mode-of-action, | | | | | |
| animals | | model system applied is highly relevant | | | | | |
| Strength of effects | | Consistent time- and dose-related change | | | | | |
| | | in assessed endpoints, several | | | | | |
| 0 | | measurements show sig | gnificant changes. | | | | |
| Scoring | | | | | | | |
| Quality of methods total | | 35 35/10 = 3.5 | | | | | |
| | Score/domain | | | | | | |
| Strength and relevance total | | 27 | | | | | |
| Total score | | 94.5 | | | | | |

| - C. I | 10 | 2005 5" | " (5.1) | | | 011 . 5 | | | | |
|-------------------------|--|---|-------------------------------------|-------------------------------|----------------------------------|-----------------------|--|--|--|--|
| Study | Quinn AL. 2005. Effects of octamethylcyclotetrasiloxane (D4) on estrous cyclicity, estradiol levels and ovarian endpoints in the female Fischer 344 rats. Dow Corning Corporation Health and Environmental Sciences Technical Report Number 10045-102. | | | | | | | | | |
| Identification | | | | er 10045-102. | | | | | | |
| Design | Material studied: D4 of 99.77 % purity, monitoring of air concentrations | | | | | | | | | |
| | Type/Strain of animals: female Fischer 344 rats | | | | | | | | | |
| | Positive controls: None | | | | | | | | | |
| | Concurrent controls: air only | | | | | | | | | |
| | Group sizes: 20 female rats per dose group | | | | | | | | | |
| | Vehicle: Air Feed: Purina Certified Rodent Chow #5002 | | | | | | | | | |
| | | | | | | | | | | |
| | Dosing and age at dosing: Animals were13 weeks old at delivery and approximately 16 weeks old at the start of exposure. Treatment was by whole-body inhalation of D4 at 0 or 700 ppm. The duration of exposure was 35 days. | | | | | | | | | |
| | Endnoints | assessed: Body weight, daily vaginal | lavage trunk blood estradiol ova | counts ovarian histonathology | animals were killed in estru | s when it was | | | | |
| | | predict cycle phase. | iavage, trank blood estraction, eva | counts, ovarian mstopathology | , ariimais were kiilea iir estra | 5 WHOTH WAS | | | | |
| Effects | | s an increase in mean cycle length in D | 04- exposed animals consisting of | an increase in number of days | in diestrus D4-exposed anir | mals showed an | | | | |
| 2.10010 | increase in | n body weight. Estradiol concentrations | s were elevated on the morning of | estrus and there was an incre | ase in the number of large fo | llicles in D4-treated | | | | |
| | | here were no D4-associated changes | | | and have a surface of large to | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | | |
| 1. Chemical well char | | Highly purified, well characterized | | | | | | | | |
| including presence | | 5 71 / | | | | | | | | |
| contaminants | | | | | | | | | | |
| 2. General experimer | ntal | | Adequate number of animals | | | | | | | |
| design (number of an | | | but no positive controls. | | | | | | | |
| dose group, controls, | | | Inhalation exposure duration | | | | | | | |
| of study duration, hou | | | was determined after the | | | | | | | |
| conditions) | 3 | | beginning of the study based | | | | | | | |
| , | | | on the response observed. | | | | | | | |
| 3. Assessment of pos | ssible | Appropriate handling of control | · | | | | | | | |
| interference from stre | ess due to | animals to reproduce possible | | | | | | | | |
| restraint, toxicity | | stressful conditions; body weight | | | | | | | | |
| | | assessed. | | | | | | | | |
| 4. Mode of applicatio | | Used effective concentration from | | | | | | | | |
| item to animals (stab | | other reproductive studies, 700 | | | | | | | | |
| vehicle used, route of | | ppm, air concentrations monitored | | | | | | | | |
| administration, dosing | g | | | | | | | | | |
| intervals) | | | | | | | | | | |
| 5. Appropriate anima | l model | | Fischer 344 rats were used in | | | | | | | |
| and strain selection | | | other studies, including the 2- | | | | | | | |
| | | | year studies, but Sprague | | | | | | | |
| | | | Dawley rats were used in the | | | | | | | |
| 0.0 % 1.00% | | 0 11 11 11 11 11 | reproductive studies. | | | | | | | |
| 6. Suitability of samp | | Sampling time was justified and the | | | | | | | | |
| measurements, samp | | attempt to standardize cycle phase | | | | | | | | |
| times, and procedure | | was appropriate. | December were to | | | | | | | |
| 7. Suitability of bioch | | | Based on manufacturers | | | | | | | |
| measurements include | arng | | specifications. | | | | | | | |

| | | | 1 | | T | T | |
|--------------------------------------|-------------|-------------------------|-----------------------|---|-----------------------------------|------------------------------|-----------|
| quality control | | | | | | | |
| Suitability of pathological/ | | | Blinding not reported | | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | | | Some but not all ra | w data | | | |
| · | | | available in report | | | | |
| 10. Statistical analysis | Appropriate | e methods with checks | | | | | |
| , | for normal | | | | | | |
| Relevance and strength of effe | ects | 3 | | 2 | | 1 | |
| Concentrations applied and their | relevance | Concentration was the s | same as in the | | | | |
| to dose/tissue concentrations of | chemical of | reproductive studies. | | | | | |
| interest resulting in adverse effect | cts in | | | | | | |
| animals | | | | | | | |
| Relevance of model system and | endpoint | | | The use of Fischer 344 rats precludes the | | | |
| assessed to key events occurring | | | | assessment of strain-dependence of the | | | |
| animals | 9 | | | | rved in the reproductive studies; | | |
| | | | | | her 344 rats is more appropriate | | |
| | | | | for assessment of the 2-year bioassay. | | | |
| Strength of effects | | | | | , | Changes in endpoint or b | oiomarker |
| 3 | | | | | | observed, but no dose of | |
| | | | | | | and limited statistical sign | |
| Scoring | | | | | | | |
| Quality of methods total | | 35 | | | | | |
| Score/domain | | 35/10 = 3.5 | | | | | |
| Strength and relevance total | | 6 | | | | | |
| Total score | | 21.0 | | | · | | |

| Study | Stump DG | 6. 2001. An inhalation study of the effe | cts of octamethylcyclotetrasiloxan | e (D4) exposure on the preovulato | ory LH surge in ovariectomi: | zed female rats | | | | | |
|--|---|--|--------------------------------------|------------------------------------|------------------------------|-------------------|--|--|--|--|--|
| Identification | | ing Corporation Health & Environmen | | | , =. roango m ovanootomi | | | | | | |
| Design | | tudied: D4 99.78% pure | | | | | | | | | |
| | Type/Strain of animals: Sprague-Dawley rat | | | | | | | | | | |
| | Positive controls: None | | | | | | | | | | |
| | Concurrent controls: Clean air | | | | | | | | | | |
| | Group size | Group sizes: 50/dose group | | | | | | | | | |
| | | Vehicle: None | | | | | | | | | |
| | Feed: PMI Certified Roden LabDiet® 5002 | | | | | | | | | | |
| | Dosing and age at dosing: 66 days old on receipt, acclimated for 6 days the nsubjected to ovariectomy. Implanted with estradiol in silastic tube, exposed three | | | | | | | | | | |
| | days later to D4 at 0, 700, or 900 ppm for 6 hours | | | | | | | | | | |
| | | assessed: Serum prolactin and lutein | | | | | | | | | |
| Effects | No change | e in mean LH concentration 0, 2, 4, 6, | or 8 hours after the end of the exp | posure period; however 4 of 10 ani | imals at 700 ppm and 7 of 1 | 10 animals at 900 | | | | | |
| | ppm had l | H concentrations below the lowest co | ntrol concentration at 6 pm (the til | me of peak LH). Prolactin was dec | reased at the end of the D4 | exposure period | | | | | |
| | | se levels. Eight hours later, serum pro | lactin in the 700 ppm group as inc | creased. Estradiol was decreased | in serum at both D4 levels a | at 0 and 2 hours | | | | | |
| | after the e | nd of the exposure period. | | | 1. | | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | | | |
| 1. Chemical well cha | racterized | Well characterized and monitored | | | | | | | | | |
| including presence | | | | | | | | | | | |
| contaminants | | | | | | | | | | | |
| 2. General experimer | | Sufficient animals to have | | | | | | | | | |
| design (number of an | | | | | | | | | | | |
| dose group, controls, | | | | | | | | | | | |
| of study duration, hou | using | | | | | | | | | | |
| conditions) | | | | | | | | | | | |
| 3. Assessment of pos | | Adequate control group, weights | | | | | | | | | |
| interference from stre | ess due to | assessed, prolactin measured. | | | | | | | | | |
| restraint, toxicity | | | | | | | | | | | |
| 4. Mode of application | n of test | Route and dose levels appropriate | | | | | | | | | |
| item to animals (stab | | for assessment of the mode of | | | | | | | | | |
| vehicle used, route of | | action. | | | | | | | | | |
| administration, dosing | g | | | | | | | | | | |
| intervals) | | | | | | | | | | | |
| Appropriate anima | l model | Sprague-Dawley rat is the strain of | | | | | | | | | |
| and strain selection | | interest in reproductive studies. | | | | | | | | | |
| 6. Suitability of samp | | Wide range of sampling times; | | | | | | | | | |
| measurements, samp | | adequate comparison of collection | | | | | | | | | |
| times, and procedure | es | from the vena cava compared to | | | | | | | | | |
| 7 Cuitobility of bissel | omica! | decapitation Well described | | | | | | | | | |
| 7. Suitability of bioch | | vveii described | | | | | | | | | |
| measurements includ | airig | | | | | | | | | | |
| quality control | logica!/ | | Dlinding not mentioned | | | | | | | | |
| 8. Suitability of patho functional assessmen | | | Blinding not mentioned | | | | | | | | |
| | | Available in study report | | | | | | | | | |
| 9. Accessibility of raw | ง น่อเล | Available in study report | | | | 1 | | | | | |

| 10. Statistical analysis | | luation of normality or ances not addressed. | | | |
|--|---|--|-------------------------------|---|--|
| Relevance and strength of effects | 3 | 2 | | 1 | |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | Relevant route and dose levels assessment of adverse effect | | | | |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | Relevant | | | | |
| Strength of effects | | Limited eff hypothesis | ect, although consistent with | | |
| Scoring | | | | | |
| Quality of methods total | 38 | | | | |
| Score/domain | 38/10 = 3.8 | | | | |
| Strength and relevance total | 18 | | | | |
| Total score | 68.4 | | | | |

| Study | Ouinn Al | , Dalu A, Meeker LS, Jean PA, Meeks | PG Crissman IW Gallavan PH | Ir Plotzka KD 2007a Effects | of actamethylovelatetr | rasilovana (D.) on the | | | | |
|--|---|---|------------------------------------|--------------------------------|-------------------------|----------------------------|--|--|--|--|
| Identification | | | | | | | | | | |
| idonanoanon | luteinizing hormone (LH) surge and levels of various reproductive hormones in female Sprague–Dawley rats. Reprod Toxicol 23:532–540. Also: | | | | | | | | | |
| | Quinn AL | Quinn AL. 2002. Effects of octamethylcyclotetrasiloxane (D ₄) on the luteinizing hormone (LH) surge and levels of various reproductive hormones in female | | | | | | | | |
| | | -Dawley rats. Dow Corning Corporation | | | | | | | | |
| Design | Material s | studied: D4 of 99.6 % purity, monitoring | of air concentrations | • | | | | | | |
| | Type/Stra | Type/Strain of animals: female SD rats from Charles River | | | | | | | | |
| | Positive of | Positive controls: not used | | | | | | | | |
| | | nt controls: air only | | | | | | | | |
| | | zes: 138 animal cannulated animals and | | | study, n ranged from 9 | to 26 in phase II | | | | |
| | | whole body inhalation exposure to 0, 70 | 00, and 900 ppm, for 3 days in two | groups of female rats, | | | | | | |
| | | rina 5002 | | | | | | | | |
| | | nd age at dosing: phase I was intact; pl | | | | | | | | |
| | | , animals in phase 1 sacrificed followin | | | mais at 2, 4, 6, 8, and | 10 pm on day of proestrus, | | | | |
| | | ic on the next morning. Animals were 1 | | | li badii walabta and ra | lative weights of | | | | |
| | | s: FSH, estradiol, estrone, progesteron ive organs, histology of ovaries in Phas | | enai biood samples in phase i | i, body weights and re | elative weights of | | | | |
| Effects | | | | and in progestorone only at 00 | O ppm: in phace II th | oro woro docrossos in L U | | | | |
| Ellecis | | In phase I animals, there were in plasma 17β-estradiol at both D4 concentrations and in progesterone only at 900 ppm; in phase II, there were decreases in LH peak values at 900 ppm at 4, 6, and 8 pm and there was reduced AUC for LH. There was increased 17β-estradiol on the morning of estrus, decreased FSH in | | | | | | | | |
| | | D4-exposed animals, reduced prolactin at 2 pm sampling point in both D4 exposed groups. Exposure to D4 decreased the proportion of rats that ovulated from | | | | | | | | |
| | | ontrols to 31% at the 900 ppm exposure | | | reased the proportion | or rate that evalated from | | | | |
| Quality of metho | | 4 | 3 | 2 | 1 | 0 | | | | |
| 1. Chemical well of | characterized | Highly purified, well characterized, | | | | | | | | |
| including presenc | е | the same batch used for all studies | | | | | | | | |
| contaminants | | | | | | | | | | |
| 2. General experi | mental | | Effects of D4 on fertility | | | | | | | |
| design (number o | | | observed after short term | | | | | | | |
| dose group, contr | | | exposures, absence of | | | | | | | |
| of study duration, | | | concurrent positive control | | | | | | | |
| conditions) | · · | | such as 15henobarbital. | | | | | | | |
| , | | | reduces confidence. | | | | | | | |
| 3. Assessment of | possible | Standard experimental design for | | | | | | | | |
| interference from | stress due to | whole body inhalation, concurrent | | | | | | | | |
| restraint, toxicity | | controls. | | | | | | | | |
| 4. Mode of application | | | Used effects concentration | | | | | | | |
| item to animals (s | | | from 2-generation and other | | | | | | | |
| vehicle used, rout | | | reproductive studies, 900 ppm | | | | | | | |
| administration, do | sing | | may produce aerosol | | | | | | | |
| intervals) | | | exposures and associated | | | | | | | |
| | | | issues with dosimetry, air | | | | | | | |
| E Appropriete | imal madal | Caragua Davidov rata ware vas dir | concentrations monitored | | | | | | | |
| Appropriate ani and strain selection | | Sprague-Dawley rats were used in the reproductive studies. | | | | | | | | |
| and strain selections. 6. Suitability of sa | | Based on reproductive physiology | | | | | | | | |
| o. Suitability of sa measurements, sa | | of rat strain, serial sampling to | | | | | | | | |
| neasurements, Sa | ampiing | or rat strain, serial sampling to | | | | | | | | |

| times, and procedures | characterize LH surge based on | | | | | | |
|--------------------------------------|--------------------------------|---|-------------------------|--|---|---|--|
| | known phys | siology. | | | | | |
| 7. Suitability of biochemical | | | Based on manufacturer's | | | | |
| measurements including | | | specifications. | | | | |
| quality control | | | | | | | |
| 8. Suitability of pathological/ | | | Blinding not discuss | sed | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | Report avai | ilable with raw data | | | | | |
| · | (Quinn, 200 | 02) | | | | | |
| 10. Statistical analysis | Appropriate | analyses with | | | | | |
| | evaluation of | of suitability for | | | | | |
| | parametric | testing | | | | | |
| Relevance and strength of effective | cts | 3 | | 2 | | 1 | |
| Concentrations applied and their | relevance | In vivo system identical t | to that used to | | | | |
| to dose/tissue concentrations of c | chemical of | identify the adverse effect, namely reduced | | | | | |
| interest resulting in adverse effect | ts in | ovulation. | | | | | |
| animals | | | | | | | |
| Relevance of model system and e | endpoint | Highly relevant to propos | sed mode of action. | | | | |
| assessed to key events occurring | g in intact | 3 , | | | | | |
| animals | | | | | | | |
| Strength of effects | | | | Only one concentration caused an effect on | | | |
| | | | | LH. | | | |
| Scoring | | | | | | | |
| Quality of methods total 36 | | 36 | | | , | | |
| Score/domain 36/10 = 3.6 | | | | | | | |
| Strength and relevance total 18 | | 18 | | | | | |
| Total score | | 64.8 | | | | | |

| Study | Siddiaui V | VH, Stump DG, Plotzke KP, Holson JF | . Meeks RG. 2007. A two-generat | ion reproductive study of octoemt | :hvlcvclotetrasiloxane (D ₄) ir | n rats exposed by | | | |
|--------------------------------------|--------------------------------|---|---------------------------------------|------------------------------------|---|-------------------|--|--|--|
| Identification | | dy vapor inhalation. Reprod Toxicol 23 | | | , | , | | | |
| | Also: | | | | | | | | |
| | | 2001. A two-generation inhalation reproductive toxicity and developmental neurotoxicity study of octmethylcyclotetracsiloxane (D4) in rats. Dow or portation Environmental Sciences Technical Report number 2001-I0000-50855. | | | | | | | |
| Design | | corporation Environmental Sciences To tudied: D4 at least 99.7% purity, moni- | | 00-50855. | | | | | |
| Design | | in of animals: SD rats from Charles Ri | | | | | | | |
| | | ontrols: none | VCI | | | | | | |
| | | nt controls: air only | | | | | | | |
| | | es: Males and females, 165/sex for the | e F0 generation plus an addition 1 | 65 females used for mating with F | 1 males. | | | | |
| | | hole body inhalation exposure to 0, 70 | | | | | | | |
| | Feed: Pur | | | | | | | | |
| | | nd age at dosing: F0 animals were 29- | | | | | | | |
| | | nating with F1 males were 70-days old | | | | | | | |
| | | nole-body inhalation to 0, 70, 300, 700 | | | | | | | |
| | | except in females from gestation day 2 | | | | | | | |
| | fomales (| tion day 5 after the first litter through on the discussed here). | gestation day 20 of the second little | r. Following the second breeding, | , Fi maies were paired with | unexposed | | | |
| | Endpoints | s assessed: Body weight, mating, vagi | nal smearing for estrous cyclicity | itter data, developmental landma | rks of offspring spermatoge | enic endocints | | | |
| Effects | Reduced | number of pups born at 500 and 700 p | opm. No effect on body weights or | body weight gains in the lactation | period. No effect on number | er of primordial | | | |
| | | the ovaries of F0 or F1 females. Incre | | | | | | | |
| Quality of methods | s | 4 | 3 | 2 | 1 | 0 | | | |
| 1. Chemical well ch | aracterized | Highly purified, well characterized | | | | | | | |
| including presence | | | | | | | | | |
| contaminants | | | | | | | | | |
| General experime | | | Adequate number of animals, | | | | | | |
| design (number of a | | | study well designed to answer | | | | | | |
| dose group, control | | | specific question about | | | | | | |
| of study duration, he | ousing | | sensitive exposure period. No | | | | | | |
| conditions) | | | positive or historical control data | | | | | | |
| 3. Assessment of po | ossible | Monitoring of body weights and | uata | | + | | | | |
| interference from st | | clinical signs, appropriate control | | | | | | | |
| restraint, toxicity | 1000 440 10 | group. | | | | | | | |
| 4. Mode of applicati | on of test | Used effects concentration from | | | | | | | |
| item to animals (sta | | other 17eproductive studies, 700 | | | | | | | |
| vehicle used, route | of | ppm, air concentrations monitored. | | | | | | | |
| administration, dosi | | | | | | | | | |
| intervals) | | | | | | | | | |
| 5. Appropriate anim | | Sprague–Dawley rats were used in | | | | | | | |
| and strain selection | | other reproduction studies. | | | | | | | |
| 6. Suitability of sam | | Standard 2-generation study | | | | | | | |
| measurements, san times, and procedu | measurements, sampling design. | | | | | | | | |
| 7. Suitability of bioc | | Standard measures | | | | | | | |
| 1. Sultability of bloc | Heililicai | Statiuatu IIIEasutes | | | | 1 | | | |

| measurements including quality control | | | N. F. C. CIE | | | |
|--|--------------------------|--|--|-------|---|--|
| 8. Suitability of pathological/ functional assessment | | | No indication of blin | ding. | | |
| Accessibility of raw data | Study repo data (Stum | rt is available with raw p, 2001). | | | | |
| 10. Statistical analysis | | | Appropriate analyse assessment of norm reported. | | | |
| Relevance and strength of effe | ects | 3 | | 2 | 1 | |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | | The exposure levels are highly relevant to those at which the adverse effect is seen in other studies. | | | | |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | | Species and strain are in which the adverse event | | | | |
| Strength of effects | | Consistent time- and dose-related changes in endpoints | | | | |
| Scoring | | | | | | |
| Quality of methods total 37 | | | | | | |
| Score/domain 37/10 = 3.7 | | | | | | |
| Strength and relevance total | | 27 | | | | |
| Total score | | 99.9 | | | | |

Supplemental Material Table 2. Quality assessment of studies supporting D4 effect mediated by estrogenicity

A. In vitro

| A. In vitro | | | | | | | | | |
|---------------------------|------------|--|---------------------------------------|---------------------------------|-------------------------------|---------------------------|--|--|--|
| Study | | nodes-Brower, S., Miller, M.R | | | X.S., Meade, B.J., 2003. Octa | amethylcyclotetrasiloxane | | | |
| identification | | strogenic activity in mice via | ERα. Toxicol Appl Pharmaco | l 192, 254–261. | | | | | |
| Design | | tudied: D4 of unstated purity | | | | | | | |
| | | Model system: Purified human estrogen receptor α and β. | | | | | | | |
| | | Positive controls: 17β-estradiol | | | | | | | |
| | Concurre | oncurrent controls: Not stated. Corn oil was used in the accompanying in vivo study. | | | | | | | |
| | Treatmen | t: D4 added to receptors in 10 | 0 mM Tris, pH 7.5, 10% glyce | erol, 2 mM dithiothreitol, and | 1 mg/ml bovine serum album | nin. | | | |
| | Endpoints | assessed: Competitive displ | lacement of labeled 17β-estra | adiol. | | | | | |
| Effects | D4 compe | eted with estradiol for binding | Era at 4×10^{-5} M and above | . D4 did not compete at Erβ. | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | |
| 1. Chemical well char | racterized | | | | | Not described | | | |
| including presence | | | | | | | | | |
| contaminants Avoida | nce of | | | | | | | | |
| contamination from | | | | | | | | | |
| equipment/dosing sol | lutions. | | | | | | | | |
| Control of adsorption | | | | | | | | | |
| glassware causing | | | | | | | | | |
| interference, appropri | riate | | | | | | | | |
| caution to avoid loss | | | | | | | | | |
| volatiles, limit of solut | | | | | | | | | |
| 2. General experimen | | | | | | Little description | | | |
| design such as numb | | | | | | , | | | |
| assays per dose/cond | | | | | | | | | |
| controls, suitability of | | | | | | | | | |
| duration. | | | | | | | | | |
| Mode of application | n of test | | | | | Not addressed | | | |
| item system (stability | | | | | | 1101 444.0004 | | | |
| used, route of applica | | | | | | | | | |
| dosing intervals, estir | | | | | | | | | |
| actual concentration | | | | | | | | | |
| chemical of interest in | | | | | | | | | |
| 4. All assessments in | | | N | ot applicable – cell-free syste | em | 1 | | | |
| determination of toxic | | | | | | | | | |
| model organism | , | | | | | | | | |
| Suitability of sample | ling | | | | | Not addressed | | | |
| method, sampling tim | | | | | | | | | |
| procedures. | ioo ana | | | | | | | | |
| 6. Suitability of bioche | emical | | | | No description of quality | 1 | | | |
| measurements include | | | | | control. | | | | |
| quality control | g | | | | 33.1.1.31. | | | | |
| quality contitor | | | | | <u>l</u> | | | | |

| 7. System used for biotransformation/cells have capacity to simulate relevant reactions that occur with chemical of interest in animals | | | No relevant information |
|--|------------|--|---|
| Accessibility of raw data | | Summary data only | У |
| 9. Statistical analysis | | No adjustment for multiple comparisons, no characterization of doseresponse relationship | |
| Relevance and strength of effects | 3 | 2 | 1 |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | | | No information about concentration that is active in vivo; no reliable assessment of actual concentration used. |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | | | Receptor binding without characterization of post-binding activity is inadequate. |
| Strength of effects | | Characterization of dose-response relationship was incomplete. | |
| Scoring | | | |
| Quality of methods total | 4 | | |
| Score/domain | 4/9 = 0.44 | | |
| Strength and relevance total | 2 | | |
| Total score | 0.9 | | |

| Study identification | vitro and Also: Plotzke K Report N | Plotzke KP. 2000. Evaluation of potential estrogenic properties of octamethylcyclotetrasiloxane (D4) using the MCF-7 cell Line: Amendment to the Final Report No. 2000-I0000-48477. | | | | | | |
|--|---|---|--|---|---|-------------------------|--|--|
| Design | Model sy Positive of Concurre (Dulbecon Treatmen Transcrip Endpoint reporter of | studied: D4 of >99% purity stem: (1) Human estrogen recontrols: 17β-estradiol, diethy ant controls: (1) Not specified o's MEM with 10% fetal boving: (1) Receptor binding: D4 detional activation assay: D4 activation activation assay: D4 activation activation assay: D4 activation a | Istilbestrol, bisphenol A for receptor binding assay, be serum) for transcriptional a elivered to culture system as ded to media at 0.1 nM, 1 nl ding: Displacement of radiola | ut possibly room air; (2) Etha activation assay a vapor at 900 ppm, resultin M, 10 nM, 100 nM, 1 μM, or 1 beled 17β-estradiol; (2) Tran | nol at concentration g in media concer 10 µM. scriptional activati | on not excentrations of | 0.45 μM. (2) transfected licoferase | |
| Lifeois | | assay, only the 10 µM conce | | | as estimated at 20 | 370. III tile t | itanisonphonal receptor | |
| Quality of methods | | 4 | 3 | 2 | | 1 | 0 | |
| Chemical well chincluding presence contaminants Avoid contamination from equipment/dosing s Control of adsorption glassware causing interference, appropaution to avoid los volatiles, limit of solesign such as num | dance of solutions. on onto oriate s of ubility) ental | | Suitable for purpose but some potentially | Reliance on supplier for information on identity and purity | | | | |
| assays per dose/concentration, suitability of exposu duration. | ire | | significant limitations identified. Lower number of repeats (run in triplicate). Dose-response only for transcriptional activation assay. | | | | | |
| 3. Mode of applicati item system (stabilii used, route of appli dosing intervals, es actual concentration chemical of interest | ty, vehicle cation, timation of n of | Concentrations of chemical of interest in medium over time determined by analytical procedures, stability of chemical of interest well assessed and solubility well characterized, vehicle controls. | | | | | | |

| All assessments include determination of toxicity to model organism | | | | | | | Assessment of cytotoxicity not reported | |
|---|--------------------------------|---|--|---|---|-------------|---|--|
| Suitability of sampling method, sampling times and procedures. | | | | | Only few sample collected, no just for sampling plant | stification | | |
| Suitability of biochemical measurements including quality control | Complies v practice | | | | | | | |
| 7. System used for biotransformation/cells have capacity to simulate relevant reactions that occur with chemical of interest in animals | (MCF-7 cel | ndard system lls) for nal activation | | | | | | |
| 8. Accessibility of raw data | | | Limitations in access to data to identify details of methodology used or results; study report has data from individual replicates. | | | | | |
| 9. Statistical analysis | | | | | Limited informa statistical appro | | | |
| Relevance and strength of effe | | 3 | | 2 | | 1 | | |
| Concentrations applied and their to dose/tissue concentrations of of interest resulting in adverse eranimals | chemical | | | | | concentra | te information; ations not selected based on I concentrations in <i>in vivo</i> | |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | | Endpoint or biomarker is clearly compatible with key event in vivo in mode-of-action, model system applied is relevant. | | | | | | |
| Strength of effects | | | | Changes in endpoint or biomarker observed, but significant changes have limited dose or time-dependency | | | | |
| Scoring | | | | | | | | |
| Quality of methods total | | | | | | | | |
| | Score/domain 21/9 = 2.3 | | | | | | | |
| Strength and relevance total | Strength and relevance total 6 | | · | | | | | |
| Total score | | 13.8 | | | | | | |

| Study identification | Lee D, Ahn C, An B-S, Jeung E-B. 2015. Induction of the estrogenic marker calbindin-D9k by octamethylcyclotetrasiloxane. Int J Environ Res Public Health. 12:14610–14625 | | | | | | | | |
|---|--|---|--|---|---|--------------------|--|--|--|
| Design | Material s Model sys Positive of Concurrer Treatment Endpoints | Material studied: D4 of unspecified purity from Sigma-Aldrich Model system: GH3 cells (a rat pituitary tumor) Positive controls: 17β-estradiol Concurrent controls: Not stated Treatment: D4 added to medium at 1 × 10 ⁻⁵ M with or without ICI 182 780 for 1 day after confluence reached Endpoints assessed: Calbindin-D9k, progesterone receptor, and estrogen receptor-α mRNA and protein | | | | | | | |
| Effects | | calbindin-D9k and progesterone receptor mRNA and protein increased by D4 treatment; the increased was blocked by ICI 182 780. Estrogen receptor xpression was down-regulated by D4, an effect that was blocked by ICI 182 780. | | | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | |
| 1. Chemical well char including presence contaminants Avoidar contamination from equipment/dosing sol Control of adsorption glassware causing interference, appropri caution to avoid loss volatiles, limit of solut 2. General experimer design such as numb assays per dose/concontrols, suitability of duration. | nce of lutions. onto iate of bility) ntal per of centration, | | Performed in triplicate. No dose-response information. | | | Not described | | | |
| 3. Mode of application item system (stability, used, route of application dosing intervals, estimactual concentration of chemical of interest in 4. All assessments in | , vehicle ation, mation of of n medium | | | | No information on measured concentration and fate of D4 in the medium | No information | | | |
| determination of toxic model organism | | | | | | INO IIIIOIIIIauoii | | | |
| 5. Suitability of sampl method, sampling tim procedures. | nes and | | | | One sample, no justification of sampling plan. | | | | |
| 6. Suitability of bioche measurements includ quality control | | Standard assessments | | | | | | | |
| 7. System used for biotransformation/cell capacity to simulate r | | | | No information about biotransformation of D4 in this system | | | | | |

| reactions that occur with | | | | | | | | |
|--|------------|---|--|---|-------|----------------|---------|---|
| chemical of interest in animals | | | | | Cum | many data anly | | |
| 8. Accessibility of raw data 9. Statistical analysis | | | Statistical methods not | | Sulli | mary data only | | |
| 3. Statistical arialysis | | | optimal, normal distribution not checked | | | | | |
| Relevance and strength of effe | ects | 3 | | 2 | | 1 | 1 | |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | | | | | | | concent | mation about relevant rations, only one ration used in this study |
| Relevance of model system and assessed to key events occurring animals | | Endpoints are compatible with key events in vivo | | | | | | |
| Strength of effects | | Changes observed but dose and time dependence not evaluated | | | | | | |
| Scoring | | | | | | | | |
| Quality of methods total | | 15 | | | | | | |
| Score/domain | 15/9 = 1.7 | | | | | | | |
| Strength and relevance total 9 | | 9 | | | | | | |
| Total score | | 15.3 | | | | | | |

| Study identification | Meeker LS 10877-102 | | ethylcyclotetracyloxane (D ₄) | to modulate hypothalamic Gn | RH release in vitro. Dow Co | orning HES Study Numnrt: |
|--|---|--|--|---|-------------------------------|------------------------------------|
| Design | Model sys Positive c Concurrer Treatmen Endpoints | ontrols: Norepinephrine (to set to controls: Krebs-Ringer/HC t: Rapidly dissected hypothats assessed: | stimulate GnRH release) Il medium alamic from decapitated anim | nypothalamic from virgin, non | I, 10, 100, or 1000 μM with o | |
| Effects | | | | retreatment at any concentrat 00 ppm but greater than in bl | | n hypothalamic explants was osure. |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 |
| 1. Chemical well char including presence contaminants Avoidal contamination from equipment/dosing soll Control of adsorption glassware causing interference, appropricaution to avoid loss volatiles, limit of solut 2. General experimer design such as numb assays per dose/controls, suitability of duration. | nce of lutions. onto iate of bility) ntal per of centration, exposure | Well characterized | No historical control data available, positive control for suppression of GnRH was not used. | | | |
| Mode of application item system (stability used, route of applications dosing intervals, estimactual concentration chemical of interest in 4. All assessments in determination of toxic model organism | , vehicle ation, mation of of medium clude | D4 concentration in tissue was assessed. | | Cytotoxicity not directly assessed, but tissue maintained its ability to respond to norepinephrine. | | |
| 5. Suitability of sampl method, sampling tim procedures. | | | Limited time course, but preliminary studies were used to justify the method. | | _ | |
| 6. Suitability of bioche measurements include quality control | | Carefully described in additional documents | | | | |
| 7. System used for | | | | | | N relevant information |

| biotransformation/cells have capacity to simulate relevant reactions that occur with chemical of interest in animals 8. Accessibility of raw data | | | | | Summary data only | <i>V</i> | |
|--|--|---------------------------|----------|---|-------------------|----------|---|
| 9. Statistical analysis App inclu | Appropriate methods including evaluation of normality of distributions | | | | | | |
| Relevance and strength of effects | 3 | | | 2 | | 1 | |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | | | | | | were us | ange of concentrations sed, but tissue trations failed to reach encountered in relevant in udies. |
| Relevance of model system and endposessed to key events occurring in in animals | | | | | | uncerta | n vitro system with some ninty about how well <i>in vivo</i> n is modeled |
| Strength of effects | | Convincing lack of effect | | | | | |
| Scoring | | | | | | | |
| Quality of methods total | | | | | | | |
| Score/domain | | 25/9 = 2.8 | | | | | |
| Strength and relevance total | | | <u> </u> | | | | |
| Total score | 8.4 | | | | | | |

B.In vivo

| Study Identification | He B, Rhodes-Brower S, Miller MR, Munson AE, Germolec DR, Walker VR, Korach KS, Meade BJ. 2003. Octamethylcyclotetrasiloxane exhibits estrogenic activity in mice via ERα. Toxicol Appl Pharmacol 192, 254–261. | | | | | | |
|--|---|--|--|--|--|--|--|
| Design | Material studied: D4 of unstated purity, dosing solutions not monitored | | | | | | |
| | Type/Strain of animals: female wild-type and Erα knock-out mice on a B6C3F1 background. Some animals ovariectomized, some animals ovariectomized. | | | | | | |
| | Positive controls: For uterotrophic assay, 17β-estradiol 10 μg/kg/day subcutaneously for 3 days | | | | | | |
| | Concurrent controls: Corn oil | | | | | | |
| Group sizes: For measurement of 17β-estradiol, 6 mice per dose group (0, 1, 10, 50, 100, 250, 500, 1000 mg/kg); for uterotrophic | | | | | | | |
| | given corn oil or D4 1000 mg/kg/day. | | | | | | |
| | Vehicle: Corn oil 10 mL/kg body weight, presumably daily, oral route (not otherwise specified). | | | | | | |
| | Feed: "Standard diet" | | | | | | |
| | Dosing and age at dosing: Animals were 6–7 weeks old at the start of the experiment. (1) Animals given D4 1, 10, 50, 100, 250, 500, or 1000 mg/kg orally (presumably daily, route not otherwise specified). (2) Uterotrophic assay: dosing with D4 1000 mg/kg/day for 3 days. | | | | | | |
| | Endpoints assessed: Serum 17β-estradiol in intact and adrenalectomized mice, uterine weight and peroxidase. | | | | | | |
| Effects | Serum 17β-estradiol was decreased by about 50% after oral exposure to D4 1000 mg/kg (presumably per day). Adrenalectomy did not alter the | | | | | | |
| | response. D4 250-1000 mg/kg/day increased uterine weight, an effect blocked by the estrogen receptor antagonist ICI 182,780. D4 1000 mg/kg/day | | | | | | |
| | increased uterine peroxidase. Uteri from Erα knock-out mice did not respond. | | | | | | |

Quality of methods 3 2 1. Chemical well No information about characterized including purity, used as obtained presence contaminants from sponsor. No justification of sample 2. General experimental design (number of animals size or duration of per dose group, controls, treatment suitability of study duration, housing conditions) 3. Assessment of possible Corticosterone did not interference from stress explain the effects of D4. due to restraint, toxicity 4. Mode of application of Standard feed not defined. test item to animals Possible estrogenic effects (stability, vehicle used, of feed or bedding not route of administration, considered. Oral dosing dosing intervals) not defined (gavage?) 5. Appropriate animal Mice have not been used model and strain selection in reproductive studies and appear to have been selected because knockouts for estrogen receptor are available. 6. Suitability of sampling Single time point measurements, sampling

| times, and procedures | | | | | | |
|--|-----------|--|---|-----------------|---------|--|
| 7. Suitability of | | Based on manufacturer | | | | |
| biochemical | | specifications, quality | | | | |
| measurements including | | control not addressed | | | | |
| quality control | | | | | | |
| 8. Suitability of | | Blinding not discussed | | | | |
| pathological/ functional | | | | | | |
| assessment O Assessibility of row data | | | | Summan, data | | |
| 9. Accessibility of raw data | | Normality not abacked | | Summary data of | Jrily | |
| 10. Statistical analysis | | Normality not checked, otherwise acceptable. | | | | |
| Relevance and strength of effects | 3 | Otherwise acceptable. | 2 | | 1 | |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical interest resulting in adverse effects in animals Relevance of model system and endpoint assessed to key events occurring in intact | Standard | assessment of estrogenicity; te for assessment of key event. | | | exposu | evels not related to human ires or exposures in species uctive effects |
| animals | арргорпа | no for addeddiment of key event. | | | | |
| Strength of effects | | | | | relevan | at a high dose level with un nee for exposure of humans omparison of ED50 to that co ol was not provided. |
| Scoring | | | | | | |
| Quality of methods total | 22 | | | | | |
| Score3/domain | 22/10 = 2 | 2.2 | | | | |
| Strength and relevance total | 3 | | | <u> </u> | | |
| Total score | 6.6 | | | | | |

| Study identification | Quinn AL, Regan JM, Tobin JM, Marinik BJ, McMahon JM, McNett DA, Sushynski CM, Crofoot SD, Jean PA, Plotzke KP. 2007b. In vitro and in vivo evaluation of the estrogenic, androgenic, and progestagenic potential of two cyclic siloxanes. Toxicol Sci 96, 145–153. Also: Quinn AL. 2003. Evaluation of octamethylcyclotetrasiloxane (D4) with rat uterotrophic assay using ovariectomized adult Sprague–Dawley rats. Dow |
|----------------------|---|
| | Corning Corporation Health & Environmental Sciences Non-Regulated Technical Report No. 2003-0000-53144. |
| Design | Material studied: D4 >99% purity, inhaled concentration monitored at least once |
| | Type/Strain of animals: Ovariectomized female Sprague-Dawley and Fischer 344 rats |
| | Positive controls: Ethinyl estradiol, genistein for uterotrophic assay, ICI 182,780 for estrogen antagonist assay |
| | Concurrent controls: Filtered air |
| | Group sizes: 6 or 10 females per strain per dose group |
| | Vehicle: Filtered air for D4, corn oil for the positive controls |
| | Feed: Not indicated |
| | Dosing and age at dosing: Age of animals not indicated except adult. D4 inhalation at 700 ppm for 16 hours/day for 3 days. Positive controls injected |
| | subcutaneously each day for 3 days. |
| | Endpoints assessed: Uterine wet and blotted weight, uterine epithelial cell height. |
| Effects | D4 had estrogenic and antiestrogenic activity in the uterotrophic assay as assessed by uterine weight and epithelial cell height. |

| Quality of methods | 4 | 3 | 2 | 1 | 0 |
|-----------------------------------|-----------------------------|---------------------------|---|---|---|
| Chemical well characterized | | Well characterized and | | | |
| including presence | | monitored. Estrogenicity | | | |
| contaminants | | of feed or bedding not | | | |
| | | considered. | | | |
| General experimental | Sample size = 12/group, | | | | |
| design (number of animals per | design and sampling plan | | | | |
| dose group, controls, suitability | typical for uterotrophic | | | | |
| of study duration, housing | assay. | | | | |
| conditions) | | | | | |
| 3. Assessment of possible | | Body weights assessed | | | |
| interference from stress due to | | but not considered in the | | | |
| restraint, toxicity | | analysis | | | |
| 4. Mode of application of test | Dose level and route | | | | |
| item to animals (stability, | were appropriate | | | | |
| vehicle used, route of | considering the | | | | |
| administration, dosing | reproductive data under | | | | |
| intervals) | evaluation | | | | |
| 5. Appropriate animal model | Use of Sprague Dawley | | | | |
| and strain selection | and Fischer 344 strains | | | | |
| | permits comparison | | | | |
| | between strains used | | | | |
| | traditionally in studies of | | | | |
| | this compound. | | | | |
| 6. Suitability of sampling | Standard sampling | | | | |
| measurements, sampling | methods for this assay | | | | |
| times, and procedures | | | | | |
| 7. Suitability of biochemical | | Quality control not | | | |

| measurements including quality control | | | addressed | | | | |
|---|--------------------------------------|-------------|---|---|---|---------|--|
| Suitability of pathological/ functional assessment | | | Blinding not addressed | | | | |
| Accessibility of raw data | | | Summary data available | | | | |
| 10. Statistical analysis | | | Comparisons of relative potency to standard estrogens were missing. Normality was not checked prior to applying parametric tests. | | | | |
| Relevance and strength of effect | cts | 3 | | 2 | | 1 | |
| Concentrations applied and their is to dose/tissue concentrations of content interest resulting in adverse effect animals | chemical of | | | | | exposur | vels not related to human res or exposures in species productive effects |
| Relevance of model system and e assessed to key events occurring animals | | | compatible with key event in vivo n, model system applied is highly | | | | |
| Strength of effects | Strength of effects Consistent and t | | ime- and dose-related change in ints, several measurements changes | | | | |
| Scoring | | | | | | | |
| Quality of methods total | | 32 | | | | | |
| Score/domain | · | 32/10 = 3.2 | <u> </u> | | · | | |
| Strength and relevance total | | 9 | | | | | |
| Total score | | 28.8 | | | | | |

| Study identification | octamethy 63:37–46 Also: Turck PA. using a ut | McKim JM Jr, Wilga PC, Breslin WJ, Plotzke KP, Gallavan RH, Meeks RG. 2001. Potential estrogenic and antiestrogenic activity of the cyclic siloxane octamethylcyclotetrasiloxane (D4) and the linear siloxane hexamethyldisiloxane (HMDS) in immature rats using the uterotrophic assay. Toxicol Sci 63:37–46. Also: Turck PA. 1998. Estrogenic and antiestrogenic activity of octamethylcyclotetrasiloxane (D4) in Sprague–Dawley and Fischer 344 immature female rats using a uterotrophic assay. Dow Corning report no. 1998-10000-45425. | | | | | | | |
|--|---|--|---|---|---|----------------------------|--|--|--|
| Design | Type/Stra Positive of Concurred Group siz Vehicle: S Feed: Not Dosing an Endpoints | nt controls: Sesame oil les: 12 females per strain per Sesame oil for the positive con indicated ad age at dosing: Sprague—D s assessed: Uterine weight (p | ale Sprague-Dawley and Fisc chylstilbestrol, coumestrol for dose group ntrols awley rats began study at 18 probably wet weight), uterine | cher 344 rats uterotrophic assay; ICI 182, days of age, Fischer 344 ra epithelial cell height. | 780 for estrogen antagonist a | age. | | | |
| Effects | million tim rats. Com | es less potent than ethinyl es | stradiol in Sprague-Dawley r | ats amd 143,000 to 25-millio | weight and epithelial cell heig n times less potent than ethin day decreased the uterotroph | yl estradiol in Fisher 344 | | | |
| Quality of method | | 4 | 3 | 2 | 1 | 0 | | | |
| Chemical well chemical including presence contaminants | | | Well characterized and monitored. Estrogenicity of feed or bedding not considered. | | | | | | |
| 2. General experim design (number of dose group, control of study duration, h conditions) | animals per ls, suitability | Sample size = 12/group, design and sampling plan typical for uterotrophic assay. | | | | | | | |
| 3. Assessment of possible interference from stress due to restraint, toxicity Clinical signs were not related to D4 treatment but body weights were affected; not considered in the analysis. | | | | | | | | | |
| 4. Mode of application of test item to animals (stability, vehicle used, route of Route and dos dose level in the route of Route and dose level in the route of Route and dose level in the route of Route and dose level in the r | | | | | Route and dose level not related to the route and dose level in the reproductive studies of interest. | | | | |
| 5. Appropriate anim and strain selection | | Use of Sprague Dawley and Fischer 344 strains permits comparison between strains used traditionally in studies of | | | | | | | |

| | this compou | und. | | | | |
|---|-------------|------------------------------------|--------------------------------------|----|---------|--|
| 6. Suitability of sampling | Standard sa | | | | | |
| measurements, sampling | methods fo | r this assay | | | | |
| times, and procedures | 0 | 411-1 | | | | |
| 7. Suitability of biochemical measurements including | Quality con | trol addressed | | | | |
| quality control | | | | | | |
| Suitability of pathological/ | | | Blinding not addressed | | | |
| functional assessment | | | · · | | | |
| Accessibility of raw data | Raw data a | | | | | |
| 10. Statistical analysis | Appropriate | analysis and | | | | |
| | comparison | | | | | |
| Delevere and strength of offe | | rd estogens | | 10 | 4 | |
| Relevance and strength of effe | | 3 | | 2 | Dana la | rala mat valata dita birrasan |
| Concentrations applied and their to dose/tissue concentrations of | | | | | | vels not related to human es or exposures in species |
| interest resulting in adverse effect | | | | | | roductive effects |
| animals | 213 111 | | | | | |
| Relevance of model system and | endpoint | | compatible with key event in vivo | | | |
| assessed to key events occurring | g in intact | | n, model system applied is highly | | | |
| animals | | relevant | | | | |
| Strength of effects | | Consistent and t assessed endpo | ime- and dose-related change in ints | | | |
| Scoring | | accessed on apa | | | | |
| Quality of methods total | | 34 | | | | |
| Score/domain | | 34/10 = 3.4 | | | | |
| Strength and relevance total | | 9 | | | | |
| Total score | | 30.6 | | | | <u> </u> |

| Study Identification | | Lee D, Ahn C, An B-S, Jeung E-B. 2015. Induction of the estrogenic marker calbindin-D _{9k} by octamethylcyclotetrasiloxane. Int J Environ Res Public Health. 12:14610–14625 | | | | | | | | |
|--|--|--|---|---|--|---------------------|--|--|--|--|
| Design | Type/Stra Positive of Concurre Group siz Vehicle: U Feed: AIN Dosing au Endpoints | etudied: D4 of unstated purity from Signation of animals: female Sprague-Dawley controls: Ethinyl estradiol 3 µg/kg/day sont controls: Unspecified vehicle less: 5 pups from the same dam per dost Jaspecified Jas | rats c for 4 days se group g/day sc for 4 days, treatment be pression of calbindin-D _{9k} , estroger | n receptor-α, and progesterone rec | | by both dose levels | | | | |
| | of D4. | | | | • | | | | | |
| Quality of method | ds | 4 | 3 | 2 | 1 | 0 | | | | |
| Chemical well chemical including presence contaminants | • | | | | | Not described | | | | |
| 2. General experim design (number of dose group, contro of study duration, h conditions) | animals per ls, suitability | | | Only 5 animals per dose group, all from the same dam. Feed and housing conditions were appropriate. | | | | | | |
| 3. Assessment of printerference from s restraint, toxicity | | | | | | Not described | | | | |
| 4. Mode of applicative to animals (stavehicle used, route administration, dos intervals) | ability, e of | | | | Dose route not appropriate, dose levels were high. | | | | | |
| 5. Appropriate anin and strain selection | | Sprague-Dawley rat is the strain of interest for reproduction studies. | | | | | | | | |
| 6. Suitability of sam measurements, san times, and procedu | mpling ures | Standard ,methods for uterotrophic assay. | | | | | | | | |
| 7. Suitability of biod measurements including quality control | luding | | No mention of quality control | | | | | | | |
| 8. Suitability of path functional assessm | nent | | Acceptable methods but blinding not addressed | | | | | | | |
| 9. Accessibility of r | | | | | Summary data only | | | | | |
| 10. Statistical analy | ysis | | Appropriate methods were used, but no assessment of normality of distributions | | | | | | | |

| Relevance and strength of effects | 3 | 2 | 1 |
|--|---------------------|---|--|
| Concentrations applied and their relevance | | | Doses and route not relevant to evaluation |
| to dose/tissue concentrations of chemical of | | | of the adverse effect under consideration |
| interest resulting in adverse effects in | | | |
| animals | | | |
| Relevance of model system and endpoint | Relevant end points | | |
| assessed to key events occurring in intact | | | |
| animals | | | |
| Strength of effects | | | Discrepancy between findings in cultured |
| | | | pituitary cells and uterotrophic assay = 0 |
| Scoring | | | |
| Quality of methods total | 21 | | |
| Score/domain | 21/10 = 2.1 | | |
| Strength and relevance total | 0 | | |
| Total score | 0 | | |

Supplemental MaterialTable 3. Quality assessment of studies supporting D4 effect mediated by dopamine agonism **A.** *In vitro*

| A. In vitro | | | | | | | | |
|--|--|---|--|-----------------------|--|--------------------|--|--|
| Study | | | opamine receptor regulation of prola | actin release from ra | t pituitary tumor-derived transformed cell | lines. Dow Corning | | |
| identification | HES Study Number: 9872-102 | | | | | | | |
| Design | Model sys cyclases. Positive c Concurred Treatmen Endpoints | controls: Dopamine nt controls: Not stated t: D4 added to RPMI medium ay 10 s assessed: Blockage of maitotoxin | μΜ stimulated prolactin secretion | | resses functional dopamine D2 receptors a | | | |
| Effects | Maitotoxir | n increased prolactin secretion by co | ells in culture. This increase was pre | evented by dopamine | e and by D4. D4 did not affect cell viability | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | |
| 1. Chemical well chain including presence contaminants Avoida contamination from equipment/dosing so Control of adsorption glassware causing interference, appropricaution to avoid loss volatiles, limit of solul | nce of lutions. onto iate of bility) | D4 was well-characterized | | | | | | |
| General experimer design such as numb assays per dose/concontrols, suitability of duration. | er of centration, | | | | Only 1 D4 concentration used. Number of replicates not given | | | |
| 3. Mode of applicatio item system (stability used, route of applica dosing intervals, estir actual concentration chemical of interest in | y, vehicle ation, mation of of medium | | | | No information on fate of chemical, no measurement of D4 concentration in medium | | | |
| All assessments in determination of toxic model organism | | Cell viability assessed. | | | | | | |
| Suitability of samp method, sampling tim procedures. | nes and | | | | No justification of sampling plan | | | |
| Suitability of biochemeasurements included quality control | | | Commercial kit measurements, quality control not addressed | | | | | |

| 7. System used for | | Well characterized | cell type | | | |
|--|------------|--------------------|-----------|--|---|--|
| biotransformation/cells have | | | | | | |
| capacity to simulate relevant | | | | | | |
| reactions that occur with | | | | | | |
| chemical of interest in animals | | | | | | |
| Accessibility of raw data | | | | | Summary data reported | |
| 9. Statistical analysis | | | | Statistical methods not discussed in detail, no evaluation of normality of distributions | | |
| Relevance and strength of effects | 3 | | 2 | | 1 | |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | | | | | Concentration in culture r with respect to in anticipa concentrations | |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | | | | nodel but it is not clear that nor cell represents in vivo events | | |
| Strength of effects | | | | | Changes observed but do dependence not evaluate | |
| Scoring | | | | | | |
| Quality of methods total | 20 | | | | | |
| Score/domain | 20/9 = 2.2 | | | | | |
| Strength and relevance total | 2 | | | | | |
| Total score | 4.4 | | | | | |

| Otrodo | D | Li IV 0044 In vitra MMO sall based | | | | 1\1 | | | |
|--|---|---|--|---|------------------------------|-----------------|--|--|--|
| Study identification | | ski JY. 2011. <i>In vitro</i> MMQ cell-based of ylcyclopentasiloxane (D5). Study 1125 | | amine receptor activation by octan | netnyicyciotetrasiloxane (d² | i) and | | | |
| Design | | studied: D4 of ~98% purity | 003-102. | | | | | | |
| Design | | | ituitary tumor: this call line secreta | os prolactio and expresses function | nal danamina D2 recentors | and adaptilate | | | |
| | | system: MMQ cells, derived from a rat pituitary tumor; this cell line secretes prolactin and expresses functional dopamine D2 receptors and adenylate | | | | | | | |
| | cyclases. Positive controls: Forskolin (activates adylate cyclase resulting in increased cellular cAMP), quinpirole (D2-receptor agonist, attenuates forskolin-induced increase in cellular cAMP), raclopiride (D2-receptor anatagonist, blocks the activity of quinpirole, permitting the forskolin-induced increase in cellular cAMI | | | | | | | | |
| | | | | | | | | | |
| | | ent controls: Dimethylsulfoxide carrier | eptor anatagonist, blocks the activi | ity of quinpirole, permitting the fors | skolin-induced increase in c | eliulai CAIVIP) | | | |
| | | nt: D4 added to RPMI medium at 0, 25 | 50 or 400 ··M | | | | | | |
| | | | | | | | | | |
| Effects | | ts assessed: Forskolin-induced increas | | of a AMD and distribution in the allege | un DO recenter enemiet. Th | - D4 -#+ | | | |
| Effects | | , 50, and 100 μM decreased forskolin- | | | | | | | |
| | | ccumulation was not altered by treatme | | | | | | | |
| | | The forskolin dose-response curve was | | | suggesting competitive inn | libition by D4. | | | |
| Ovelity of motherin | Pertussis | s toxin did not block the activity of D4 s | | | 4 | | | | |
| Quality of methods | 4 | 4 | 3 | 2 | 1 | 0 | | | |
| 1. Chemical well char | racterized | | | | Reliance on supplier for | | | | |
| including presence | | | | | information on identity | | | | |
| contaminants Avoidar | nce of | | | | and purity. | | | | |
| contamination from | | | | | Concentration of | | | | |
| equipment/dosing sol | | | | | chemical of interest in | | | | |
| Control of adsorption | onto | | | | medium not | | | | |
| glassware causing | | | | | considered, use of | | | | |
| interference, appropri | | | | | media from established | | | | |
| caution to avoid loss | _ | | | | supplier | | | | |
| volatiles, limit of solub | | | | | | | | | |
| General experiment | | | Suitable for purpose but some | | | | | | |
| design such as numb | | | potentially significant | | | | | | |
| assays per dose/cond | | | limitations identified. Lower | | | | | | |
| controls, suitability of | exposure | | number of repeats or no | | | | | | |
| duration. | | | positive or historic control data | | | | | | |
| | | | available. Some aspects of | | | | | | |
| | | | study duration and sampling | | | | | | |
| | | | plan are questionable. Only | | | | | | |
| | _ | | limited dose-response | | | | | | |
| 3. Mode of application | | | | | Limited information on | | | | |
| item system (stability, | | | | | concentration and fate | | | | |
| used, route of applica | | | | | of chemical applied | | | | |
| dosing intervals, estin | | | | | | | | | |
| actual concentration of | _ | | | | | | | | |
| chemical of interest in | | - | | | | | | | |
| 4. All assessments in | | Detailed toxicity assessment by | | | | | | | |
| determination of toxic | city to | appropriate methods in controls | | | | | | | |
| model organism | | and exposed system, cytotoxicity of | | | | | | | |
| | | chemical of interest in system well | | | | | | | |
| | | defined and reported | | | | | | | |
| Suitability of sampl | ling | | | | Only few samples | | | | |

| method, sampling times and procedures. | | | | | | collected, no justification for sampling plan | |
|---|-----------------------|---|--|---|---|---|-------------------------|
| Suitability of biochemical measurements including quality control | | | Selection not compl with best practice lir parameter consister assessed | miting inter | | | |
| 7. System used for biotransformation/cells have capacity to simulate relevant reactions that occur with chemical of interest in animals | | | | | | | No relevant information |
| Accessibility of raw data Statistical analysis | Appropriate | e statistical method analysis of endpoint. | | | | | |
| | Checked fo | r normal distribution | | | | | |
| Relevance and strength of effe | | 3 | | 2 | in a de sur de inferenceire | 1 | |
| Concentrations applied and their to dose/tissue concentrations of interest resulting in adverse effect animals | chemical of cts in | | | Concentratione order of concentration reasonably exposure of effect | cinadequate information. cions required to induce effect are of magnitude above tons of the chemical of interest expected in tissue under conditions causing the adverse | | |
| Relevance of model system and assessed to key events occurring animals | | | | limitations i | d model system, but some regarding relevance of endpoint I for sequence of events resulting effect in vivo | | |
| Strength of effects | | | | Changes in observed, b | n endpoint or biomarker but significant changes have e or time-dependency | | |
| Scoring | | | | | | | |
| Quality of methods total | | 21 | | | | | - |
| Score/domain | | 12/9 = 2.3 | | | | | |
| Strength and relevance total | | 8 | | | | | |
| Total score | | 18.4 | | | | | |

B. In vivo

| Study Identification | Batelle, 2004. Batelle, Toxicology Northwest, Technical Report for Dow Corning Corporation - 24-Month combined chronic toxicity and oncogenicity whole body vapor inhalation study of octamethylcyclotetrasiloxane (D4) in Fischer 344 rats. Report number 2004-I0000-54091, HES/DCC Study number 9106, Batelle Study number N003441A | | | | | | | | |
|---|---|--|---|--|--|--|--|--|--|
| Design | Type/Stra Positive of Concurrer Group size Vehicle: n Feed: Pur Dosing an was an int of exposu after 24 m Endpoints | and age at dosing: Animals were 7–8 we terim kill of 6 animals/sex/dose group are, necropsy of 20 animals/sex/dose grouths of exposure assessed: Body weight, tissue conce | er 344 rats 00 ppm, 6 hours/day for 5 days; verseks old at experiment start. Treating to 6 months for evaluation of tissue proup after 12 months of exposure entrations of D4, clinical pathology, | ehicle for pergolide was ethanol ment was by whole body inhalatic concentrations of D4, necropsy followed by 12 months of recover ophthalmoscopy, gross and hist | on with D4 at 0, 10, 30, 150 of 10 animals/dex/dose grery, and necropsy of 60 animals/dex/dose | oup after 12 months mals/sex/dose group | | | |
| Effects | There were no effects of treatment on survival and only sporadic effects on body weight of females. There were no toxicologically important effects on clinical chemistry or hematology. There were decreases in terminal body weight and increases in relative kidney, iver, and uterine weight in the 700-ppm females. Endometrial stromal polyps were present in 1 of 10 females necropsied after 12 months of exposure. Endometrial adenomas were present in 4 of 60 females necropsied after 24 months of exposure with a statistically significant trend in this lesion. Endometrial hyperplasia occurred in all dose groups, including the control, but there was a statistical increase in endometrial hyperplasia at 700 ppm. There were instances of other proliferative lesions that were not statistically significant or dose-related. | | | | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | |
| Chemical well cha including presence contaminants | racterized | Highly purified, well characterized | | | | | | | |
| 2. General experiment design (number of ar dose group, controls, of study duration, how conditions) | nimals per , suitability | | Adequate number of animals with negative controls. Positive controls were not used. | | | | | | |
| Assessment of posinterference from streetestraint, toxicity | ess due to | Appropriate handling of control animals to reproduce possible stressful conditions | | | | | | | |
| 4. Mode of application of test item to animals (stability, vehicle used, route of administration, dosing intervals) Range of concentrations was used including the maximum vapor concentration achievable for a chronic study. | | | | | | | | | |
| 5. Appropriate anima and strain selection6. Suitability of samp | | Fischer 344 rats were used in the 2-year study, an acceptable strain Reasonable sampling intervals | | | | | | | |
| measurements, samp times, and procedure | pling | Trousonable sampling intervals | | | | | | | |

| 7. Suitability of biochemical | Details of n | neasurements and | | | | | |
|--------------------------------------|--------------|---------------------------------------|-------------------------------|----|--|--------------|--|
| measurements including | quality con | trol were given. | | | | | |
| quality control | | - | | | | | |
| 8. Suitability of pathological/ | | | Blinding not reporte | ed | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | | | | | | Summary data | |
| 10. Statistical analysis | | e methods with checks | | | | | |
| | for normal | | | | | | |
| Relevance and strength of effe | | 3 | | 2 | | 1 | |
| Concentrations applied and their | | Concentrations required | | | | | |
| to dose/tissue concentrations of | | | ected; this study established | | | | |
| interest resulting in adverse effect | cts in | the adverse effect of interest | | | | | |
| animals | | | | | | | |
| Relevance of model system and | | Relevant to the effect being valuated | | | | | |
| assessed to key events occurring | g in intact | | | | | | |
| animals | | | | | | | |
| Strength of effects | | Changes in endpoint or | | | | | |
| | | observed, dose and time | e dependence | | | | |
| | | characterized | | | | | |
| Scoring | | | | | | | |
| Quality of methods total | | 35 | | | | | |
| Score/domain | | 36/10 = 3.5 | | | | | |
| Strength and relevance total | | 27 | | | | | |
| Total score | | 94.5 | | | | | |

| | FI: 00: | 0.5% | (54.0401) 550.07.6 | | 0.10.11 5.11.00.0) | 1.0 1.0 | | |
|---|------------|--|-------------------------------------|------------------------------------|-----------------------------|---------------------|--|--|
| Study | | Effect of octamethylcyclotetrasiloxar | | | CAS No. 541-02-6) on circ | culating prolactin | | |
| Identification | | ne aged female Fischer 344 rat. Study | | No. 2010-10000-62617 | | | | |
| Design | | tudied: D4 of 99.77 % purity, monitorin in of animals: female Fischer 344 rats f | | | | | | |
| | | ontrols: Pergolide 0.2 mg/kg by gavage | | | | | | |
| | | ontrols: Pergolide 0.2 mg/kg by gavage of controls: air only | e on days I and 5 | | | | | |
| | | <i>n controls.</i> all only es: 31 animal cannulated animals, n ra | nged from 8 to 10 per group; addi | tional animals exposed to D5 are i | not considered here | | | |
| | | ose-only inhalation exposure to 0 or 70 | | | | | | |
| | Feed: Pur | | bo ppin, o nodis/day for 5 days, ve | chiefe for pergonae was enfanorm | COTT OIL | | | |
| | | nd age at dosing: Animals were ≥20 mo | onths old at experiment start. Trea | tment was by nose-only inhalation | for D4 and by gayage for a | pergolide. | | |
| | | assessed: Body weight, daily vaginal | | | | | | |
| | | posure, after at least 6 hours of exposi | | | · | | | |
| Effects | | ve control (pergolide) increased prolact | | | not altered, progesterone w | as decreased on | | |
| 1 | | ere was an increase in estrogenic vagir | | | | n after 1 or 5 days | | |
| | of exposur | re to D4. Venous prolactin concentration | on was increased 4 and 8 hours af | ter the end of the D4 exposure pe | riod on day 5. | · | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | |
| 1. Chemical well char | racterized | Highly purified, well characterized, | | | | | | |
| including presence | | same batch used for all studies | | | | | | |
| contaminants | | | | | | | | |
| 2. General experimer | ntal | | Adequate number of animals | | | | | |
| design (number of an | imals per | | with positive and negative | | | | | |
| dose group, controls, | | | controls. Inhalation exposure | | | | | |
| of study duration, hou | ısing | | duration not well justified, | | | | | |
| conditions) | | | sampling intervals not well | | | | | |
| | | | justified. | | | | | |
| Assessment of pos | | Appropriate handling of control | | | | | | |
| interference from stre | ess due to | animals to reproduce possible | | | | | | |
| restraint, toxicity | | stressful conditions | | | | | | |
| 4. Mode of application | | Used effective concentration from | | | | | | |
| item to animals (stabi vehicle used, route of | | other reproductive studies, 700 ppm, air concentrations monitored | | | | | | |
| administration, dosing | | ppm, air concentrations monitored | | | | | | |
| intervals) | 9 | | | | | | | |
| 5. Appropriate animal | l model | Fischer 344 rats were used in the | | | | | | |
| and strain selection | imodoi | 2-year studies. The use of the aged | | | | | | |
| and ottain oblocuon | | animal is useful for evaluation of | | | | | | |
| | | the findings of the 24-month | | | | | | |
| | bioassay. | | | | | | | |
| 6. Suitability of sampl | | | | | | | | |
| measurements, samp | | | | | | | | |
| times, and procedure | res | | | | | | | |
| 7. Suitability of bioche | | | Based on manufacturers | | | | | |
| measurements includ | ling | | specifications. | | | | | |
| quality control | | | | | | | | |
| 8. Suitability of pathol | logical/ | | Blinding not reported | | | | | |

| functional assessment | | | | | | | |
|---|-------------|--|-----------------|---|------|-----------------------------|-----------|
| Accessibility of raw data | Raw data a | vailable in report | | | | | |
| 10. Statistical analysis | Appropriate | methods with checks | | | | | |
| - | for normal | or normal distribution | | | | | |
| Relevance and strength of effe | ects | 3 | | 2 | | 1 | |
| Concentrations applied and their to dose/tissue concentrations of | | Concentrations required in the range of concentrations | | | | | |
| interest resulting in adverse effect | | chemical of interest reas | | | | | |
| animals | 013 111 | tissue under exposure co | | | | | |
| | | the adverse effect | onamono cadomig | | | | |
| Relevance of model system and | | Relevant to the effect be | ing valuated | | | | |
| assessed to key events occurring animals | g in intact | | | | | | |
| Strength of effects | | | | | | Changes in endpoint or | hiomarkar |
| Strength of effects | | | | | | observed, but no dose of | |
| | | | | | | and limited statistical sig | |
| Scoring | | | | | | | |
| Quality of methods total | | 36 | | | | | |
| Score/domain | | 36/10 = 3.6 | | | | | |
| Strength and relevance total | | 9 | | | | | |
| Total score | | 32.4 | | | | | |

| Ctudy | Llomos LT | Γ. 2010. <i>In vivo</i> evaluation of the impac | at of expension and point explication | timing on the notantial for actom | othylovolototropilovono ond | | | | | | | |
|-------------------------------------|-----------|---|--|-------------------------------------|----------------------------------|----------------------|--|--|--|--|--|--|
| Study Identification | | ylcyclopentasiloxane to affect circulatir | | | | | | | | | | |
| luerillication | | Technical Report. Study number 1125 | | -tieated leffiale Fischer F344 fat. | Dow Corning Fleatin and E | Invitorimental | | | | | | |
| Design | | tudied: D4 of 99.77 % purity, monitorin | | | | | | | | | | |
| Doolgii | | in of animals: female Fischer 344 rats | | | | | | | | | | |
| | | | s: Pergolide 0.2 mg/kg by oral gavage | | | | | | | | | |
| | | nt controls: air only | ago | | | | | | | | | |
| | | | 2 ovariectomized animals and 72 intact animals in negative control and treatment groups; 8 animals in positive control group and 32 animals in | | | | | | | | | |
| | | e treatment groups. | | | | | | | | | | |
| | | ose-only inhalation exposure to 0 or 70 | 00 ppm for 6 days in 4 groups of fe | emale rats (a D5-exposed group i | s not included in this summ | nary). The vehicle | | | | | | |
| | | de was a solution containing 10% etha | | | | | | | | | | |
| | osmosis v | vater. | | · | | | | | | | | |
| | Feed: Pur | ina 5002 | | | | | | | | | | |
| | Dosing an | nd age at dosing: Animals were 9 week | s old at the start of the experimen | t. All animals except negative cor | trol group received reserpi | ne 12.5 mg/kg by | | | | | | |
| | | 8 am, repeated 24 hours later. Negati | | | | | | | | | | |
| | | ose of reserpine or vehicle, the positive | | | | de. Immediately | | | | | | |
| | | olide or vehicle administration, animals | | | | | | | | | | |
| | | assessed: Body weight gain, mortality | | | | | | | | | | |
| Effects | | s no mortality in the negative control gr | | | | | | | | | | |
| | | in two of the groups receiving reserping | | | | | | | | | | |
| | | ot exposed to D4. Prolactin concentration | | | | | | | | | | |
| | | rolactin as expected. Reserpine treatm | | | | | | | | | | |
| | | ad a blood prolactin concentration siming prolactin. Eight hours after the end of | | | | | | | | | | |
| | | nours after the end of the inhalation pe | | | | | | | | | | |
| | _ | in air-treated animals. | nod, rescripine was wearing on, ar | ia protactiri was acciming. In D- | ireated ariiriais at triis tirie | point, protactin was | | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | | | | |
| 1. Chemical well cha | | Highly purified, well characterized, | | | | | | | | | | |
| including presence | | same batch used for all studies | | | | | | | | | | |
| contaminants | | | | | | | | | | | | |
| 2. General experime | ntal | | Adequate number of animals | | | | | | | | | |
| design (number of ar | | | with positive and negative | | | | | | | | | |
| dose group, controls | | | controls. Inhalation exposure | | | | | | | | | |
| of study duration, ho | | | duration not well justified, | | | | | | | | | |
| conditions) | 9 | | sampling intervals not well | | | | | | | | | |
| | | | justified | | | | | | | | | |
| 3. Assessment of pos | ssible | Appropriate use of controls to | - | | | | | | | | | |
| interference from stre | | neutralize possible effects of stress | | | | | | | | | | |
| restraint, toxicity | | | | | | | | | | | | |
| 4. Mode of application | | Standard experimental design for | | | | | | | | | | |
| item to animals (stab | | nose-only exposure, sampling | | | | | | | | | | |
| vehicle used, route o | | documented lack of systemic | | | | | | | | | | |
| administration, dosin | g | exposure of unexposed groups | | | | | | | | | | |
| intervals) | | | | | | | | | | | | |
| Appropriate anima | ıl model | Fischer 344 rats were used in the | | | | 1 | | | | | | |

| and strain selection | 2-year bioa | issay. | | | | | |
|--------------------------------------|-------------|-------------------------|---------------------|-----------|--------------------------------|----------------------------|--------------|
| 6. Suitability of sampling | | | | | Serial sampling intervals were | | |
| measurements, sampling | | | | | not justified. | | |
| times, and procedures | | | | | | | |
| 7. Suitability of biochemical | | | Based on manufact | turers | | | |
| measurements including | | | specifications | | | | |
| quality control | | | | | | | |
| 8. Suitability of pathological/ | | | Blinding was not re | ported. | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | | ilable with raw data | | | | | |
| 10. Statistical analysis | Appropriate | e methods were used | | _ | | | |
| Relevance and strength of effe | | 3 | | 2 | | 1 | |
| Concentrations applied and their | | Exposure level was base | | | | | |
| to dose/tissue concentrations of | | findings and was approp | oriate. | | | | |
| interest resulting in adverse effect | cts in | | | | | | |
| animals | | | | | | | |
| Relevance of model system and | | | | | erpinized animals is of | | |
| assessed to key events occurring | g in intact | | | | le relevance to human | | |
| animals | | | | assessmer | nt. | | |
| Strength of effects | | | | | | Effects of treatment docu | |
| | | | | | | 18 hours after exposure, | |
| | | | | | | directions, and not at 0 a | and 8 hours. |
| Scoring | | | | | | | |
| Quality of methods total 36 | | | | | | | |
| Score/domain 3. | | 3.6/10 = 3.6 | | | | | |
| Strength and relevance total | | 6 | | | | | |
| Total score | | 21.6 | | | | | |

| Otrodo | Martin D | 2. Otrono DO Otableo (MIL Halana II | District ICD Description (1997) | A i.e.la. al. ati | | -t! |
|-------------------------|----------------|--|-------------------------------------|---|------------------------------|-----------------------------------|
| Study Identification | | G, Stump DG, Siddiqui WH, Holson JF, | | | study of octamethylcyclot | etrasiloxane (D ₄) in |
| identification | Also: | ts using multiple and single day exposi | are regimens. Reprod Toxicol 23. i | 192–201. | | |
| | | LE. 1998. An inhalation reproductive to | vicity study of octamethylcycloteti | rasilovana (DA) in female rats usi | na multinle evnosure regim | ens Dow Corning |
| | | on Health and Environmental Sciences | | | ig maniple exposure regin | iens. Dow Conning |
| Design | | studied: D4 of >99% purity, monitoring | | 7 11100. | | |
| Doolgii | | nin of animals: SD rats from Charles Riv | | | | |
| | | controls: none | . •. | | | |
| | | nt controls: air only | | | | |
| | | es: 24 females per dose group expose | d 28 days prior to mating, during r | mating, and until gestation day 19 | . An additional 60, 60, and | 24 animals, |
| | | ely, were exposed to the highest dose I | | | | |
| | | day 3, and only on gestation days 2-5 | | | | |
| | inhalation | at 700 ppm. | · | | · | |
| | Vehicle: v | whole body inhalation exposure to 0, 70 |), 300, 500, or 700 ppm, 6 hours/d | lay for 5 days | | |
| | Feed: Pui | | | | | |
| | | nd age at dosing: Animals were 59-71 | days old on receipt and were accli | imated for 14 days prior to the sta | irt of the experiment. Treat | ment was by whole- |
| | body inha | | | | | |
| | Endpoints | s assessed: Body weight, gravid uterine | e weight, number of corpora lutea, | number and location of fetuses, i | mplantation sites. | |
| Effects | | d mean body weight in 700 ppm dams. | | | | |
| | | tation loss, and decreased viable fetus | | sitive period for reduction of corpo | ora lutea and implantation s | sites and for |
| | preimplan | tation loss was from 3 days before ma | ting to gestation day 3. | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 |
| 1. Chemical well cha | aracterized | Highly purified, well characterized, | | | | |
| including presence | | same batch used for all studies | | | | |
| contaminants | | | | | | |
| 2. General experime | ental | | Adequate number of animals, | | | |
| design (number of a | nimals per | | study well designed to answer | | | |
| dose group, controls | s, suitability | | specific question about | | | |
| of study duration, ho | ousing | | sensitive exposure period. No | | | |
| conditions) | | | positive or historical control | | | |
| | | | data | | | |
| 3. Assessment of po | ssible | Stress was evaluated using animal | | | | |
| interference from str | ess due to | weight changes and adrenal | | | | |
| restraint, toxicity | | weights; appropriate controls used. | | | | |
| 4. Mode of application | | | | D4 contamination was not | | |
| item to animals (stat | | | | assessed, possible effects of | | |
| vehicle used, route | | | | estrogen feed, bedding not | | |
| administration, dosir | ng | | | assessed. Dose level and | | |
| intervals) | | | | route (whole body inhalation) were appropriate. | | |
| 5. Appropriate anima | al model | | Sprague Dawley rats might not | | | |
| and strain selection | | 1 | be appropriate for evaluation | Í | | |
| and ottain obloction | | | | | | |
| and otrain colociton | | | of the 24-month bioassay in | | | |
| 6. Suitability of samp | | Sampling intervals were justified | | | | |

| measurements, sampling | and sampli | ng methods were | | | | | |
|--------------------------------------|--------------|---------------------------|------------------------|-----------|------|----------|--|
| times, and procedures | adequate. | | | | | | |
| 7. Suitability of biochemical | Based on n | nanufacturer's | | | | | |
| measurements including | specificatio | ns | | | | | |
| quality control | | | | | | | |
| 8. Suitability of pathological/ | | | Blinding was not rep | oorted. | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | Accessible | in Kaufman (1998) | | | | | |
| 10. Statistical analysis | | | Adequate methods | but no | | | |
| | | | mention of evaluation | on of | | | |
| | | | normality of distribu | tions for | | | |
| | | | parametric testing. | | | | |
| Relevance and strength of effe | ects | 3 | | 2 | | 1 | |
| Concentrations applied and their | relevance | Concentrations required | I to induce effect are | | | | |
| to dose/tissue concentrations of | chemical of | in the range of concentra | | | | | |
| interest resulting in adverse effect | cts in | chemical of interest reas | | | | | |
| animals | | tissue under exposure c | conditions causing | | | | |
| | | the adverse effect | J | | | | |
| Relevance of model system and | endpoint | Endpoint or biomarker is | s clearly compatible | | | | |
| assessed to key events occurring | | with key event in vivo in | | | | | |
| animals | 9 | model system applied is | | | | | |
| Strength of effects | | Consistent time- and do | | | | | |
| | | in assessed endpoints, | | | | | |
| | | measurements show sig | | | | | |
| Scoring | | · | , J | | | | |
| Quality of methods total | | 34 | | | | | |
| Score/domain | | 34/10 = 3.4 | | | | | |
| Strength and relevance total | | 27 | | | | <u> </u> | |
| Total score | | 91.8 | · | | | | |

| Study | Quinn AL | . 2005. Effects of octamethylcyclotetras | siloxane (D4) on estrous cyclicity, | estradiol levels and ovarian end | points in the female Fis | scher 344 rats. Dow | | | | |
|---|--|--|--------------------------------------|----------------------------------|--------------------------|----------------------------|--|--|--|--|
| Identification | | Corporation Health and Environmental S | | | • | | | | | |
| Design | | tudied: D4 of 99.77 % purity, monitorin | | | | | | | | |
| · · | | in of animals: female Fischer 344 rats | | | | | | | | |
| | 7. | ontrols: None | | | | | | | | |
| | Concurrer | nt controls: air only | | | | | | | | |
| | | es: 20 female rats per dose group | | | | | | | | |
| | Vehicle: A | | | | | | | | | |
| | Feed: Purina Certified Rodent Chow #5002 Dosing and age at dosing: Animals were13 weeks old at delivery and approximately 16 weeks old at the start of exposure. Treatment was by whole-body | | | | | | | | | |
| | | | | | | | | | | |
| | inhalation | of D4 at 0 or 700 ppm. The duration of | exposure was 35 days. | | | • | | | | |
| | Endpoints | assessed: Body weight, daily vaginal | lavage, trunk blood estradiol, ova | counts, ovarian histopathology; | animals were killed in e | estrus when it was | | | | |
| | possible t | o predict cycle phase. | - | | | | | | | |
| Effects | There was | s an increase in mean cycle length in D | 4- exposed animals consisting of | an increase in number of days | in diestrus. D4-exposed | d animals showed an | | | | |
| | increase i | n body weight. Estradiol concentrations | s were elevated on the morning of | estrus and there was an increa | ise in the number of lar | ge follicles in D4-treated | | | | |
| | | There were no D4-associated changes | in the number of oocytes in the over | viducts. | | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | | |
| Chemical well cha | aracterized | Highly purified, well characterized | | | | | | | | |
| including presence | | | | | | | | | | |
| contaminants | | | | | | | | | | |
| 2. General experime | ental | | Adequate number of animals | | | | | | | |
| design (number of a | | | but no positive controls. | | | | | | | |
| dose group, controls | | | Inhalation exposure duration | | | | | | | |
| of study duration, ho | | | was determined after the | | | | | | | |
| conditions) | Ü | | beginning of the study based | | | | | | | |
| , | | | on the response observed. | | | | | | | |
| 3. Assessment of po | ossible | Appropriate handling of control | • | | | | | | | |
| interference from str | ress due to | animals to reproduce possible | | | | | | | | |
| restraint, toxicity | | stressful conditions; body weight | | | | | | | | |
| | | assessed. | | | | | | | | |
| Mode of application | | Used effective concentration from | | | | | | | | |
| tem to animals (stal | | other reproductive studies, 700 | | | | | | | | |
| vehicle used, route | | ppm, air concentrations monitored | | | | | | | | |
| administration, dosir | ng | | | | | | | | | |
| intervals) | | | | | | | | | | |
| Appropriate anima | | Fischer 344 rats were used in the | | | | | | | | |
| and strain selection | | 2-year bioassay. | | | | | | | | |
| 6. Suitability of samp | | Sampling time was justified and the | | | | | | | | |
| measurements, sam | | attempt to standardize cycle phase | | | | | | | | |
| imes, and procedur | | was appropriate. | | | | | | | | |
| Suitability of bioch | hemical | | Based on manufacturers | | | | | | | |
| measurements inclu | ıding | | specifications. | | | | | | | |
| quality control | | | | | | | | | | |
| 3. Suitability of path | • | | Blinding not reported | | | | | | | |
| unctional assessme | ent | | | | | | | | | |

| Accessibility of raw data | | | Some but not all rav | w data | | | |
|--|------------------------|---|----------------------|--------|--|---|--------------------|
| 10. Statistical analysis | Appropriate for normal | e methods with checks distribution | | | | | |
| Relevance and strength of effe | ects | 3 | | 2 | | 1 | |
| Concentrations applied and their to dose/tissue concentrations of interest resulting in adverse effect animals | chemical of | Concentration was the s reproductive studies. | ame as in the | | | | |
| Relevance of model system and assessed to key events occurring animals | | Fischer 344 rats are appropriate for assessment of the 2-year bioassay. | | | | | |
| Strength of effects | | | | | | Changes in endpoint or observed, but no dose of and limited statistical significant controls. | or time dependence |
| Scoring | | | | | | | |
| Quality of methods total 35 | | 35 | | | | | |
| Score/domain 36/10 = 3.6 | | | | | | | |
| Strength and relevance total 9 | | 9 | | | | | |
| Total score | | 32.4 | • | | | | |

| Study | Stump DG | 6. 2001. An inhalation study of the effe | cts of octamethylcyclotetrasiloxan | e (D4) exposure on the preovulate | ory LH surge in ovariectomia | zed female rats. | | | | | |
|---|--------------|--|------------------------------------|-----------------------------------|---------------------------------|------------------|--|--|--|--|--|
| Identification | | ing Corporation Health & Environmen | | | , oaigo iii ovailooloiliii | | | | | | |
| Design | | tudied: D4 99.78% pure | | | | | | | | | |
| | | in of animals: Sprague-Dawley rat | | | | | | | | | |
| | | ontrols: None | | | | | | | | | |
| | Concurrer | nt controls: Clean air | | | | | | | | | |
| | | es: 50/dose group | | | | | | | | | |
| | Vehicle: N | phicle: None | | | | | | | | | |
| | | I Certified Roden LabDiet® 5002 | | | | | | | | | |
| | | nd age at dosing: 66 days old on receip | | ected to ovariectomy. Implanted w | ith estradiol in silastic tube, | exposed three | | | | | |
| | | to D4 at 0, 700, or 900 ppm for 6 hour | | | | | | | | | |
| | | assessed: Serum prolactin and lutein | | | | | | | | | |
| Effects | | e in mean LH concentration 0, 2, 4, 6, | | | | | | | | | |
| | | H concentrations below the lowest co | | | | | | | | | |
| | | se levels. Eight hours later, serum pro and of the exposure period. | lactin in the 700 ppm group as inc | creased. Estradiol was decreased | in serum at both D4 levels a | at 0 and 2 hours | | | | | |
| Quality of methods | anter the e | 4 | 3 | 2 | 1 | 0 | | | | | |
| Chemical well cha | racterized | Well characterized and monitored | | _ | - | | | | | | |
| including presence | . 401011204 | Tron characterized and memored | | | | | | | | | |
| contaminants | | | | | | | | | | | |
| 2. General experime | ntal | Sufficient number of animals, | | | | | | | | | |
| design (number of ar | | adequate housing | | | | | | | | | |
| dose group, controls | | adoquate nodeg | | | | | | | | | |
| of study duration, ho | | | | | | | | | | | |
| conditions) | 3 | | | | | | | | | | |
| 3. Assessment of pos | ssible | Adequate control group, weights | | | | | | | | | |
| interference from stre | ess due to | assessed, prolactin measured. | | | | | | | | | |
| restraint, toxicity | | | | | | | | | | | |
| 4. Mode of application | n of test | Route and dose levels appropriate | | | | | | | | | |
| item to animals (stab | | for assessment of the mode of | | | | | | | | | |
| vehicle used, route o | | action. | | | | | | | | | |
| administration, dosin | g | | | | | | | | | | |
| intervals) | | | | | | | | | | | |
| 5. Appropriate anima | ı model | | Sprague-Dawley rat not used | | | | | | | | |
| and strain selection | lina | Wide range of compling times: | in the 2-year bioassay | | | | | | | | |
| 6. Suitability of samp measurements, samp | | Wide range of sampling times; adequate comparison of collection | | | | | | | | | |
| times, and procedure | | from the vena cava compared to | | | | | | | | | |
| unico, and procedure | decapitation | | | | | | | | | | |
| 7. Suitability of bioch | | | | | | | | | | | |
| measurements include | | | | | | | | | | | |
| quality control | J | | | | | | | | | | |
| 8. Suitability of patho | logical/ | | Blinding not mentioned | | | | | | | | |
| functional assessmen | | | | | | | | | | | |
| 9. Accessibility of rav | v data | Available in study report | | | | | | | | | |

| 10. Statistical analysis | | on of normality or es not addressed. | | | |
|--|---|--------------------------------------|-------------------------------|---|--|
| Relevance and strength of effects | 3 | 2 | | 1 | |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | Relevant route and dose levels for assessment of adverse effect of in | | | | |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | | Strain of ra | at not ideal | | |
| Strength of effects | | Limited eff hypothesis | ect, although consistent with | | |
| Scoring | | | | | |
| Quality of methods total | 37 | | | | |
| Score/domain | 38/10 = 3.7 | | | | |
| Strength and relevance total | 12 | | | | |
| Total score | 44.4 | | | | |

| Ctudy | Ouinn Al | Dalu A Maakari S Jaan DA Maaka | PC Cricomon IM Colloyon PU | Ir Diotaka KD 2007a Effacts of | a ata mathylayalatatra ail | |
|---|---|--|---|--------------------------------|----------------------------|---|
| Study Identification | | Dalu A, Meeker LS, Jean PA, Meeks | | | | |
| Design | luteinizing hormone (LH) surge and levels of various reproductive hormones in female Sprague–Dawley rats. Reprod Toxicol 23:532–540. Also: | | | | | |
| | Quinn AL. 2002. Effects of octamethylcyclotetrasiloxane (D ₄) on the luteinizing hormone (LH) surge and levels of various reproductive hormones in female | | | | | |
| | Sprague—Dawley rats. Dow Corning Corporation Health & Environmental Sciences Report Number 2002-10000-51695 | | | | | |
| | Material studied: D4 of 99.6 % purity, monitoring of air concentrations | | | | | |
| 3 | Type/Strain of animals: female SD rats from Charles River | | | | | |
| | Positive controls: not used | | | | | |
| | Concurrent controls: air only | | | | | |
| | Group sizes: 138 animal cannulated animals and 73 non-canulated, only animals with regular cycle included in study, n ranged from 9 to 26 in phase II | | | | | |
| | Vehicle: whole body inhalation exposure to 0, 700, and 900 ppm, for 3 days in two groups of female rats, | | | | | |
| | Feed: Purina 5002 | | | | | |
| | Dosing and age at dosing: phase I was intact; phase II 51henobarbi for serial blood samples, exposures on diestrus days 1 and 2(6h/day) and for 2.5 h on day of | | | | | |
| | proestrus, animals in phase 1 sacrificed following last exposure (10 am); blood samples taken from phase II animals at 2, 4, 6, 8, and 10 pm on day of proestrus, | | | | | |
| | animal sac on the next morning. Animals were 13 weeks of age at initiation of exposures | | | | | |
| | Endpoints: FSH, estradiol, estrone, progesterone in phase I; LH and prolactin in serial blood samples in phase II; body weights and relative weights of | | | | | |
| Effects | reproductive organs, histology of ovaries in Phase II | | | | | |
| | In phase I animals, there were in plasma 17β-estradiol at both D4 concentrations and in progesterone only at 900 ppm; in phase II, there were decreases in LH | | | | | |
| | peak values at 900 ppm at 4, 6, and 8 pm and there was reduced AUC for LH. There was increased 17β-estradiol on the morning of estrus, decreased FSH in | | | | | |
| | D4-exposed animals, reduced prolactin at 2 pm sampling point in both D4 exposed groups. Exposure to D4 decreased the proportion of rats that ovulated from 79% in controls to 31% at the 900 ppm exposure group with a reduction in mean number of eggs in the oviduct. | | | | | |
| | | | aroun with a reduction in mean r | number of eags in the aviduct | | |
| O 1:1 1 11 1 | | ntrois to 31% at the 900 ppm exposure | group with a reduction in mean i | | 14 | |
| Quality of method | ls | 4 | 3 | 2 | 1 | 0 |
| 1. Chemical well ch | ls naracterized | 4 Highly purified, well characterized, | 3 | | 1 | 0 |
| Chemical well chincluding presence | ls naracterized | 4 | 3 | | 1 | 0 |
| Chemical well chemical well chemical well chemical well chemical well contaminants | ls naracterized | 4 Highly purified, well characterized, | 3 | | 1 | 0 |
| Chemical well ch including presence contaminants General experim | naracterized | 4 Highly purified, well characterized, | Effects of D4 on fertility | | 1 | 0 |
| Chemical well chincluding presence contaminants General experim design (number of a | naracterized nental animals per | 4 Highly purified, well characterized, | Effects of D4 on fertility observed after short term | | 1 | 0 |
| Chemical well chincluding presence contaminants General experim design (number of a dose group, control) | naracterized nental animals per ls, suitability | 4 Highly purified, well characterized, | Effects of D4 on fertility observed after short term exposures, absence of | | 1 | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h | naracterized nental animals per ls, suitability | 4 Highly purified, well characterized, | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control | | 1 | 0 |
| Chemical well chincluding presence contaminants General experim design (number of a dose group, control) | naracterized nental animals per ls, suitability | 4 Highly purified, well characterized, | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. | | 1 | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) | naracterized nental animals per ls, suitability nousing | Highly purified, well characterized, the same batch used for all studies | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control | | 1 | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p | naracterized nental animals per ls, suitability nousing | Highly purified, well characterized, the same batch used for all studies Standard experimental design for | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. | | 1 | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from st | naracterized nental animals per ls, suitability nousing | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. | | 1 | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from st restraint, toxicity | nental animals per ls, suitability nousing | Highly purified, well characterized, the same batch used for all studies Standard experimental design for | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. | | 1 | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from strestraint, toxicity 4. Mode of applicati | naracterized nental animals per ls, suitability nousing possible tress due to | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. Used effects concentration | | 1 | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from strestraint, toxicity 4. Mode of applicatitem to animals (sta | nental animals per ls, suitability nousing cossible tress due to tion of test ability, | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. Used effects concentration from 2-generation and other | | 1 | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from st restraint, toxicity 4. Mode of application item to animals (stavehicle used, route | nental animals per ls, suitability nousing cossible tress due to tion of test ability, of | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. Used effects concentration from 2-generation and other reproductive studies, 900 ppm | | 1 | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from st restraint, toxicity 4. Mode of application to animals (stavehicle used, route administration, dosi | nental animals per ls, suitability nousing cossible tress due to tion of test ability, of | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. Used effects concentration from 2-generation and other reproductive studies, 900 ppm may produce aerosol | | | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from st restraint, toxicity 4. Mode of application item to animals (stavehicle used, route | nental animals per ls, suitability nousing cossible tress due to tion of test ability, of | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. Used effects concentration from 2-generation and other reproductive studies, 900 ppm may produce aerosol exposures and associated | | | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from st restraint, toxicity 4. Mode of application to animals (stavehicle used, route administration, dosi | nental animals per ls, suitability nousing cossible tress due to tion of test ability, of | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. Used effects concentration from 2-generation and other reproductive studies, 900 ppm may produce aerosol exposures and associated issues with dosimetry, air | | | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from st restraint, toxicity 4. Mode of applicatifiem to animals (stavehicle used, route administration, dosi intervals) | nental animals per ls, suitability nousing possible tress due to tion of test ability, of ing | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. Used effects concentration from 2-generation and other reproductive studies, 900 ppm may produce aerosol exposures and associated issues with dosimetry, air concentrations monitored | | | 0 |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from strestraint, toxicity 4. Mode of applicatifiem to animals (stavehicle used, route administration, dosi intervals) 5. Appropriate anim | nental animals per ls, suitability nousing possible tress due to tion of test ability, e of ining | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. Used effects concentration from 2-generation and other reproductive studies, 900 ppm may produce aerosol exposures and associated issues with dosimetry, air concentrations monitored Sprague-Dawley rats are not | | | |
| 1. Chemical well chincluding presence contaminants 2. General experim design (number of a dose group, control of study duration, h conditions) 3. Assessment of p interference from st restraint, toxicity 4. Mode of applicatifiem to animals (stavehicle used, route administration, dosi intervals) | nental animals per ls, suitability nousing possible tress due to tion of test ability, e of ining | Highly purified, well characterized, the same batch used for all studies Standard experimental design for whole body inhalation, concurrent | Effects of D4 on fertility observed after short term exposures, absence of concurrent positive control such as phenobarbital. reduces confidence. Used effects concentration from 2-generation and other reproductive studies, 900 ppm may produce aerosol exposures and associated issues with dosimetry, air concentrations monitored | | | 0 |

| measurements, sampling | of rat strain | , serial sampling to | | | | | |
|--|---------------------------|---|-------------------------|--|--|---|--|
| times, and procedures | | e LH surge based on | | | | | |
| , , | known phys | | | | | | |
| 7. Suitability of biochemical | | | Based on manufacturer's | | | | |
| measurements including | | | specifications. | | | | |
| quality control | | | | | | | |
| 8. Suitability of pathological/ | | | Blinding not discuss | sed | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | Report ava (Quinn, 200 | ilable with raw data 02) | | | | | |
| 10. Statistical analysis | Appropriate | e analyses with of suitability for | | | | | |
| Relevance and strength of effe | | 3 | | 2 | | 1 | |
| Concentrations applied and their to dose/tissue concentrations of interest resulting in adverse effect animals | relevance chemical of | In vivo system identical identify the adverse efferovulation. | | | | | |
| Relevance of model system and assessed to key events occurring animals | | Highly relevant to proposed mode of action. | | | | | |
| Strength of effects | Strength of effects | | | Only one concentration caused an effect on LH. | | | |
| Scoring | | | | | | | |
| Quality of methods total 36 | | 36 | - | | | | |
| Score/domain 36/10 = 3 | | 36/10 = 3.6 | <u>-</u> | | | | |
| Strength and relevance total | | 18 | | | | | |
| Total score | | 64.8 | | | | | |

| Chudu | C:dd:au.:V | VIII Charan DC District KD Helega I | T. Maraka D.C. 2007. A true manage | tion wormen describes of early of outcome | | (D) is rate as a said by | | | | |
|-------------------------|--|---|---------------------------------------|---|---------------------------|--|--|--|--|--|
| Study | | VH, Stump DG, Plotzke KP, Holson J | | tion reproductive study of octoer | ntnyicyciotetrasiioxane | e (D ₄) in rats exposed by | | | | |
| Identification | Also: | dy vapor inhalation. Reprod Toxicol 2 | 3.202–215. | | | | | | | |
| | | G. 2001. A two-generation inhalation reproductive toxicity and developmental neurotoxicity study of octmethylcyclotetracsiloxane (D4) in rats. Dow | | | | | | | | |
| | | ing Corporation Environmental Sciences Technical Report number 2001-10000-50855. | | | | | | | | |
| Design | | studied: D4 at least 99.7% purity, mon | | 00-30633. | | | | | | |
| Design | | | | | | | | | | |
| | Type/Strain of animals: SD rats from Charles River Positive controls: none Concurrent controls: air only | | | | | | | | | |
| | | | | | | | | | | |
| | | res: Males and females, 165/sex for th | o E0 gonoration plus an addition 1 | 65 famales used for mating with | E1 malos | | | | | |
| | | whole body inhalation exposure to 0, 7 | | | iri illales. | | | | | |
| | Feed: Pur | | 0, 300, 300, 01 700 ppm, 6 flours/0 | lay lot 5 days | | | | | | |
| | | nd age at dosing: F0 animals were 29 | 30 days old on receipt and were a | ecclimated for 15 days prior to th | a start of the experime | ant. The additional females | | | | |
| | | nating with F1 males were 70-days of | | | | | | | | |
| | | hole-body inhalation to 0, 70, 300, 70 | | | | | | | | |
| | | except in females from gestation day | | | | | | | | |
| | | ition day 5 after the first litter through | | | | | | | | |
| | | not discussed here). | gestation day 20 of the second little | i. I ollowing the second breedin | g, i i iliales wele palit | ed with unexposed | | | | |
| | | s assessed: Body weight, mating, vag | inal emparing for estrous cyclicity | litter data developmental landm | arke of offenring energ | matagenic endocints | | | | |
| Effects | Peduced | number of nume born at 500 and 700 | nom. No effect on body weights or | hody weight gains in the lactation | on period. No effect on | number of primordial | | | | |
| LIIGOIS | | sed number of pups born at 500 and 700 ppm. No effect on body weights or body weight gains in the lactation period. No effect on number of primordial s in the ovaries of F0 or F1 females. Increase in estrous cycle length (prolonged diestrus) and decreased mating index in F1a females at 700 ppm. | | | | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | | |
| Chemical well cha | | Highly purified, well characterized | 3 | 2 | | U | | | | |
| including presence | aracterized | Highly purified, well characterized | | | | | | | | |
| contaminants | | | | | | | | | | |
| | | | | | | | | | | |
| 2. General experime | | | Adequate number of animals, | | | | | | | |
| design (number of a | | | study well designed to answer | | | | | | | |
| dose group, controls | | | specific question about | | | | | | | |
| of study duration, ho | busing | | sensitive exposure period. No | | | | | | | |
| conditions) | | | positive or historical control | | | | | | | |
| | | | data | | | | | | | |
| 3. Assessment of po | | Monitoring of body weights and | | | | | | | | |
| interference from str | ess due to | clinical signs, appropriate control | | | | | | | | |
| restraint, toxicity | • • • | group. | | | | | | | | |
| 4. Mode of application | | Used effects concentration from | | | | | | | | |
| item to animals (stat | | other reproductive studies, 700 | | | | | | | | |
| vehicle used, route of | | ppm, air concentrations monitored. | | | | | | | | |
| administration, dosir | ıg | Stability addressed. | | | | | | | | |
| intervals) | | | | | | | | | | |
| | Appropriate animal model Sprague—Dawley rats were not | | | | | | | | | |
| and strain selection | - 1' | Oten deed O managetica etc. | used in the 2-year bioassay | <u> </u> | | | | | | |
| 6. Suitability of samp | | Standard 2-generation study | | | | | | | | |
| measurements, sam | | design. | | | | | | | | |
| times, and procedure | es | | | | | | | | | |
| 7. Suitability of bioch | | Standard measures | | | | | | | | |

| | 1 | | 1 | | I | 1 | |
|---|--|--|--|--|---------------------------------|---|--|
| measurements including quality control | | | | | | | |
| Suitability of pathological/ functional assessment | | | No indication of blir | nding. | | | |
| Accessibility of raw data | | Study report is available with raw data (Stump, 2001). | | | | | |
| 10. Statistical analysis | | | Appropriate analyse assessment of norm reported. | | | | |
| Relevance and strength of eff | ects | 3 | | 2 | | 1 | |
| to dose/tissue concentrations of interest resulting in adverse effe animals | Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | | The exposure levels are highly relevant to those at which the adverse effect is seen in other studies. | | detain d'étand fann tha atabair | | |
| Relevance of model system and assessed to key events occurring animals | | | | Species and strain differed from the study in which the adverse events were documented | | | |
| Strength of effects | | Consistent time- and do in endpoints | se-related changes | | | | |
| Scoring | | | | | | | |
| Quality of methods total 36 | | 36 | | | | | |
| Score/domain 36/10 = 3.6 | | 36/10 = 3.6 | | | | | |
| Strength and relevance total | | 18 | | | | | |
| Total score | | 64.8 | | | | | |

| Study | Sloter ED. | 2015. A dietary and inhalation vagina | cytology study of chronically adn | ninistered pergolide, octamethylcy | clotetrasiloxane (D4) or | | | | | |
|---|---|---|-----------------------------------|--------------------------------------|------------------------------|-------------------|--|--|--|--|
| Identification | | ylcyclopentasiloxane (D5) in aging Fisc | | | (= 1, 1 | | | | | |
| Design | Material s | tudied: D4 ≥99.5% purity, monitoring o | f air concentrations | | | | | | | |
| | | in of animals: female Fischer-344 rats | | | | | | | | |
| | | ontrols: Pergolide in the diet at 0.0045, | 0.045, or 4.5 ppm | | | | | | | |
| | Concurrent controls: air only Group sizes: 50 animals/dose group Vehicle: whole body inhalation exposure to 0 or 700 ppm, for 58 weeks, | | | | | | | | | |
| | | | | | | | | | | |
| | | | | | | | | | | |
| | Feed: Purina Certified Rodent LabDiet® 5002 | | | | | | | | | |
| | Dosing an | nd age at dosing: Animals were 49–50 | weeks old at onset of exposure. | | | | | | | |
| | Endpoints | : Clinical observations, body weight, fo | od consumption, daily vaginal lav | age, serum prolactin, estradiol, pro | ogesterone, corticosterone | | | | | |
| Effects | The positive | ve control pergolide at all dose levels p | roduced an increase in number o | f days with estrogenic vaginal lava | ige (proestrus/estrus). D4 e | exposure was | | | | |
| | associated | d with an increase in total number of es | trogenic days. Serum estradiol co | oncentration and estrogen:progest | erone ratio were elevated b | by D4 compared to | | | | |
| | control. Pr | olactin concentrations were similar in I | 04-exposed animals and controls | . Corticosterone was decreased by | / D4. Histopathology showe | ed an increase in | | | | |
| | vaginal thi | ckness and an increase in cystic endo | metrial hyperplasia. | | | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | | |
| 1. Chemical well char | acterized | Highly purified, well characterized | | | | | | | | |
| including presence | | | | | | | | | | |
| contaminants | | | | | | | | | | |
| 2. General experimen | ntal | Adequate number of animals, use | | | | | | | | |
| design (number of an | | of aged animals may be highly | | | | | | | | |
| dose group, controls, | | relevant to the adverse effect under | | | | | | | | |
| of study duration, hou | | evaluation. Study duration and | | | | | | | | |
| conditions) | 9 | housing were adequate. | | | | | | | | |
| 3. Assessment of pos | sible | Standard experimental design for | | | | | | | | |
| interference from stre | | whole body inhalation, concurrent | | | | | | | | |
| restraint, toxicity | | controls. Corticosterone was not | | | | | | | | |
| ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,, | | increased by D4 inhalation. | | | | | | | | |
| 4. Mode of application | n of test | Used effects concentration from 2- | | | | | | | | |
| item to animals (stabi | | generation and other reproductive | | | | | | | | |
| vehicle used, route of | f | studies, appropriate dose regimen. | | | | | | | | |
| administration, dosing | g | | | | | | | | | |
| intervals) | | | | | | | | | | |
| 5. Appropriate animal | model | Fischer-344 rats are the strain of | | | | | | | | |
| and strain selection | | interest in evaluation of the adverse | | | | | | | | |
| | | effect. | | | | | | | | |
| 6. Suitability of sampl | ing | Suitable serial assessments of | | | | | | | | |
| measurements, samp | | hormones and vaginal cytology | | | | | | | | |
| times, and procedure | S | | | | | | | | | |
| 7. Suitability of bioche | | Detailed procedures and quality | | | | | | | | |
| measurements includ | ling | control | | | | | | | | |
| quality control | | | | | | | | | | |
| 8. Suitability of pathol | logical/ | | Blinding not discussed | | | | | | | |
| functional assessmen | | | | | | | | | | |
| 9. Accessibility of raw | / data | Report available with raw data | | | | | | | | |

| evalua | opriate analyses with ation of suitability for netric testing | | | |
|---|---|---|-----------------|--|
| Relevance and strength of effects | 3 | 2 | 1 | |
| Concentrations applied and their relevar to dose/tissue concentrations of chemica interest resulting in adverse effects in animals | | 0 | | |
| Relevance of model system and endpoir assessed to key events occurring in inta animals | | action | | |
| Strength of effects | | Only one concentration of D precluding dose-response e the effects were clearly dem | evaluation, but | |
| Scoring | | | | |
| Quality of methods total | 38 | | | |
| Score/domain | 35/10 = 3.8 | | | |
| Strength and relevance total | 18 | | | |
| Total score | 68.4 | | | |

Supplemental Material
Table 4. Quality assessment of studies supporting D4 effect mediated by estrogenicity

A. In vitro

| A. III VILIO | | | | | | | | | |
|---------------------------|--|--|---|---------------------------------|----------------------------|--------------------|--|--|--|
| Study | He, B., Rhodes-Brower, S., Miller, M.R., Munson, A.E., Germolec, D.R., Walker, V.R., Korach, K.S., Meade, B.J., 2003. Octamethylcyclotetrasiloxane | | | | | | | | |
| identification | | exhibits estrogenic activity in mice via ERα. Toxicol Appl Pharmacol 192, 254–261. | | | | | | | |
| Design | | aterial studied: D4 of unstated purity | | | | | | | |
| | | odel system: Purified human estrogen receptor α and β. | | | | | | | |
| | | ive controls: 17β-estradiol | | | | | | | |
| | | rrent controls: Not stated. Corn oil was used in the accompanying in vivo study. | | | | | | | |
| | | | 0 mM Tris, pH 7.5, 10% glyce | | 1 mg/ml bovine serum album | nin. | | | |
| | | | lacement of labeled 17β-estra | | | | | | |
| Effects | D4 compe | eted with estradiol for binding | $_{ m I}$ Erα at 4 $	imes$ 10 $^{	ext{-}5}$ M and above | . D4 did not compete at Erβ. | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | |
| 1. Chemical well char | racterized | | | | | Not described | | | |
| including presence | | | | | | | | | |
| contaminants Avoida | nce of | | | | | | | | |
| contamination from | | | | | | | | | |
| equipment/dosing sol | | | | | | | | | |
| Control of adsorption | onto | | | | | | | | |
| glassware causing | | | | | | | | | |
| interference, appropri | iate | | | | | | | | |
| caution to avoid loss | of | | | | | | | | |
| volatiles, limit of solub | bility) | | | | | | | | |
| 2. General experimen | ntal | | | | | Little description | | | |
| design such as numb | | | | | | | | | |
| assays per dose/cond | centration, | | | | | | | | |
| controls, suitability of | exposure | | | | | | | | |
| duration. | | | | | | | | | |
| 3. Mode of application | | | | | | Not addressed | | | |
| item system (stability, | , vehicle | | | | | | | | |
| used, route of applica | | | | | | | | | |
| dosing intervals, estin | mation of | | | | | | | | |
| actual concentration | of | | | | | | | | |
| chemical of interest in | | | | | | | | | |
| 4. All assessments in | clude | | N | ot applicable – cell-free syste | m | | | | |
| determination of toxic | city to | | | • | | | | | |
| model organism | | | | | | | | | |
| 5. Suitability of sampl | ling | | | | | Not addressed | | | |
| method, sampling tim | | | | | | | | | |
| procedures. | | | | | | | | | |
| 6. Suitability of bioche | emical | | | | No description of quality | | | | |
| measurements includ | | | | | control. | | | | |
| quality control | J | | | | | | | | |
| | | | | | | | | | |

| 7. System used for biotransformation/cells have capacity to simulate relevant reactions that occur with chemical of interest in animals | | | No relevant information |
|--|------------|--|---|
| Accessibility of raw data | | Summary data only | у |
| 9. Statistical analysis | | No adjustment for multiple comparisons, no characterization of doseresponse relationship | |
| Relevance and strength of effects | 3 | 2 | 1 |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | | | No information about concentration that is active in vivo; no reliable assessment of actual concentration used. |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | | | Receptor binding without characterization of post-binding activity is inadequate. |
| Strength of effects | | Characterization of dose-response relationship was incomplete. | |
| Scoring | | | |
| Quality of methods total | 4 | | |
| Score/domain | 4/9 = 0.44 | | |
| Strength and relevance total | 2 | | |
| Total score | 0.9 | | |

| Study identification | vitro and Also: Plotzke K Report N | Plotzke KP. 2000. Evaluation of potential estrogenic properties of octamethylcyclotetrasiloxane (D4) using the MCF-7 cell Line: Amendment to the Final Report No. 2000-10000-48477. | | | | | | | |
|--|---|--|--|---|--------------------|---|---|--|--|
| Design Effects | Model sy Positive of Concurre (Dulbecon Treatmen Transcrip Endpoint reporter of | Material studied: D4 of >99% purity Model system: (1) Human estrogen receptor α and β; (2) Transfected MCF-7 (human epithelial breast cancer cells). Positive controls: 17β-estradiol, diethylstilbestrol, bisphenol A Concurrent controls: (1) Not specified for receptor binding assay, but possibly room air; (2) Ethanol at concentration not exceeding 0.25% in media (Dulbecco's MEM with 10% fetal bovine serum) for transcriptional activation assay Treatment: (1) Receptor binding: D4 delivered to culture system as a vapor at 900 ppm, resulting in media concentrations of 0.45 μΜ. (2) Transcriptional activation assay: D4 added to media at 0.1 nM, 1 nM, 10 nM, 100 nM, 1 μM, or 10 μΜ. Endpoints assessed: (1) Receptor binding: Displacement of radiolabeled 17β-estradiol; (2) Transcriptional activation assay: transfected licoferase reporter gene activation D4 atmospheric exposure of 900 ppm bound to ERα but not Erβ. The extent of binding to Erα was estimated at 20%. In the transcriptional receptor | | | | | | | |
| | activation | assay, only the 10 µM conce | | | ao ootimatoa at 2t | ,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,, | | | |
| Quality of methods | | 4 | 3 | 2 | | 1 | 0 | | |
| Chemical well chincluding presence contaminants Avoid contamination from equipment/dosing s Control of adsorptio glassware causing interference, appropaution to avoid loss volatiles, limit of sol General experime | dance of solutions. on onto oriate s of ubility) | | Suitable for purpose but | Reliance on supplier for information on identity and purity | | | | | |
| design such as num assays per dose/concentration, suitability of exposu duration. | nber of , controls, ire | | some potentially significant limitations identified. Lower number of repeats (run in triplicate). Dose-response only for transcriptional activation assay. | | | | | | |
| 3. Mode of applicati item system (stabilit used, route of applic dosing intervals, estactual concentration chemical of interest | ty, vehicle cation, timation of n of | Concentrations of chemical of interest in medium over time determined by analytical procedures, stability of chemical of interest well assessed and solubility well characterized, vehicle controls. | | | | | | | |

| All assessments include determination of toxicity to model organism | | | | | | | Assessment of cytotoxicity not reported |
|---|---|------------|--|--|---|-------------|---|
| Suitability of sampling method, sampling times and procedures. | | | | | Only few sample collected, no just for sampling plant | stification | |
| Suitability of biochemical measurements including quality control | Complies v practice | | | | | | |
| 7. System used for biotransformation/cells have capacity to simulate relevant reactions that occur with chemical of interest in animals | Use of standard system (MCF-7 cells) for transcriptional activation assay | | | | | | |
| 8. Accessibility of raw data | | | Limitations in access to data to identify details of methodology used or results; study report has data from individual replicates. | | | | |
| 9. Statistical analysis | | | | | Limited informa statistical appro | | |
| Relevance and strength of effe | | 3 | | 2 | | 1 | |
| Concentrations applied and their to dose/tissue concentrations of of interest resulting in adverse eranimals | chemical | | | | | concentra | te information; ations not selected based on I concentrations in <i>in vivo</i> |
| assessed to key events occurrin animals | Relevance of model system and endpoint assessed to key events occurring in intact | | narker is clearly compatible with in mode-of-action, model s relevant. | | | | |
| Strength of effects | | | | Changes in endpoint or bi but significant changes ha time-dependency | | | |
| Scoring | | | | | | | |
| Quality of methods total | | 21 | | | | | |
| Score/domain | | 21/9 = 2.3 | | | | | |
| Strength and relevance total | | 6 | | | | | |
| Total score | | 13.8 | | | | | |

| Study identification | Lee D, Ahn C, An B-S, Jeung E-B. 2015. Induction of the estrogenic marker calbindin-D9k by octamethylcyclotetrasiloxane. Int J Environ Res Public Health. 12:14610–14625 | | | | | | | | |
|--|--|--|-------------------------------|---|---|---------------------------|--|--|--|
| Design | Model sys Positive c Concurred Treatmen Endpoints | Material studied: D4 of unspecified purity from Sigma-Aldrich Model system: GH3 cells (a rat pituitary tumor) Positive controls: 17β-estradiol Concurrent controls: Not stated Treatment: D4 added to medium at 1 x 10 ⁻⁵ M with or without ICI 182 780 for 1 day after confluence reached Endpoints assessed: Calbindin-D9k, progesterone receptor, and estrogen receptor-α mRNA and protein | | | | | | | |
| Effects | | -D9k and progesterone recept n was down-regulated by D4 | | ased by D4 treatment; the inc by ICI 182 780. | reased was blocked by ICI 1 | 82 780. Estrogen receptor | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | |
| 1. Chemical well char including presence contaminants Avoidal contamination from equipment/dosing sol Control of adsorption glassware causing interference, appropricaution to avoid loss volatiles, limit of solut 2. General experimer | nce of lutions. onto iate of billity) | | Performed in triplicate. | | | Not described | | | |
| design such as numb assays per dose/cond controls, suitability of duration. | er of centration, exposure | | No dose-response information. | | | | | | |
| 3. Mode of application item system (stability used, route of applications dosing intervals, estimactual concentration chemical of interest in | , vehicle ation, mation of of | | | | No information on measured concentration and fate of D4 in the medium | | | | |
| All assessments in determination of toxic model organism | city to | | | | | No information | | | |
| Suitability of sampl method, sampling tim procedures. | nes and | | | | One sample, no justification of sampling plan. | | | | |
| Suitability of bioche measurements include quality control | | Standard assessments | | | | | | | |
| 7. System used for biotransformation/cell capacity to simulate r | | | | No information about biotransformation of D4 in this system | | | | | |

| e a | | | | | | | | |
|--|-----|---|--|---|------|---------------|---|-----|
| reactions that occur with | | | | | | | | |
| chemical of interest in animals | | | | | | | | |
| Accessibility of raw data | | | | | Summ | ary data only | | |
| 9. Statistical analysis | | | Statistical methods not optimal, normal distribution not checked | | | | | |
| Relevance and strength of effe | cts | 3 | | 2 | 2 | | 1 | |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | | | | | | С | No information about concentrations, only concentration used in | one |
| Relevance of model system and assessed to key events occurring animals | | Endpoints are vivo | compatible with key events in | | | | | |
| Strength of effects | | Changes observed but dose and time dependence not evaluated | | | | | | |
| Scoring | | | | | | | | |
| Quality of methods total | | 15 | | | | | | |
| Score/domain | | 15/9 = 1.7 | | | | | | |
| Strength and relevance total | | 9 | | | | | | |
| Total score | | 15.3 | | | | | | |

| Study identification | Meeker L 10877-10 | | ethylcyclotetracyloxane (D ₄) t | o modulate hypothalamic Gr | RH release in vitro. Dow Co | orning HES Study Numnrt: |
|--|--|--|--|--|-----------------------------|--------------------------|
| Design | Model sys Positive o Concurre Treatmen Endpoints | controls: Norepinephrine (to int controls: Krebs-Ringer/H0 it: Rapidly dissected hypothe is assessed: | CI medium alamic from decapitated anim | als were cultured with D4 at | 1, 10, 100, or 1000 μM with | |
| Effects | | | nRH was not altered by D4 privo studies after inhalation o 7 | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 |
| 1. Chemical well chaincluding presence contaminants Avoid contamination from equipment/dosing s Control of adsorptio glassware causing interference, appropaution to avoid loss volatiles, limit of soli | lance of olutions. on onto oriate s of ubility) | Well characterized | No historical control data | | | |
| 2. General experime design such as num assays per dose/cor controls, suitability of duration. | nber of ncentration, | | No historical control data available, positive control for suppression of GnRH was not used. | | | |
| 3. Mode of applicative item system (stability used, route of application dosing intervals, est actual concentration chemical of interest 4. All assessments determination of tox | ty, vehicle cation, timation of of in medium include | D4 concentration in tissue was assessed. | | Cytotoxicity not directly assessed, but tissue | | |
| model organism | • | | | maintained its ability to respond to norepinephrine. | | |
| 5. Suitability of sam method, sampling ti procedures. | | | Limited time course, but preliminary studies were used to justify the method. | | | |
| Suitability of biocl measurements inclu quality control | | Carefully described in additional documents | | | | |
| 7. System used for | | | | | | N relevant information |

| biotransformation/cells have capacity to simulate relevant reactions that occur with chemical of interest in animals 8. Accessibility of raw data | | Summary data on | lv |
|--|--|-----------------|--|
| 9. Statistical analysis Approprincluding | ate methods gevaluation of gof distributions | | |
| Relevance and strength of effects | 3 | 2 | 1 |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical interest resulting in adverse effects in animals | | | Large range of concentrations were used, but tissue concentrations failed to reach levels encountered in relevant in vivo studies. |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | | | Novel in vitro system with some uncertainty about how well <i>in vivo</i> situation is modeled |
| Strength of effects | Convincing lack of effect | | |
| Scoring | | | |
| Quality of methods total | 25 | | |
| Score/domain | 25/9 = 2.8 | | |
| Strength and relevance total | 3 | | |
| Total score | 8.4 | | |

B.In vivo

| D.III VIVO | | | | | | | | | |
|---|---|--|---|--|---|---|--|--|--|
| Study | He B, Rhodes-Brower S, Miller MR, Munson AE, Germolec DR, Walker VR, Korach KS, Meade BJ. 2003. Octamethylcyclotetrasiloxane exhibits | | | | | | | | |
| Identification | | | Toxicol Appl Pharmacol 192, 2 | | | | | | |
| Design | Type/S Positiv Concu Group corn of Vehicle | Material studied: D4 of unstated purity, dosing solutions not monitored Type/Strain of animals: female wild-type and Erα knock-out mice on a B6C3F1 background. Some animals ovariectomized, some animals ovariectomized. Positive controls: For uterotrophic assay, 17β-estradiol 10 μg/kg/day subcutaneously for 3 days Concurrent controls: Corn oil Group sizes: For measurement of 17β-estradiol, 6 mice per dose group (0, 1, 10, 50, 100, 250, 500, 1000 mg/kg); for uterotrophic assay, 5 animals given corn oil or D4 1000 mg/kg/day. Vehicle: Corn oil 10 mL/kg body weight, presumably daily, oral route (not otherwise specified). Feed: "Standard diet" | | | | | | | |
| | Dosing (presu Endpo | g and age at dosing: Animals ν mably daily, route not otherwis ints assessed: Serum 17β-est | se specified). (2) Uterotrophic radiol in intact and adrenalect | rt of the experiment. (1) Anima assay: dosing with D4 1000 m omized mice, uterine weight a | ng/kg/day for 3 days. nd peroxidase. | | | | |
| Effects | D4 250 uterine |)–1000 mg/kg/day increased ເ | | sure to D4 1000 mg/kg (presuled by the estrogen receptor ar | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | |
| | | | | | No information about purity, used as obtained from sponsor. | | | | |
| 2. General experime design (number of a per dose group, con suitability of study duration, housing conditions) | nimals | | | No justification of sample size or duration of treatment | | | | | |
| Assessment of pointerference from str due to restraint, toxic | ess | Corticosterone did not explain the effects of D4. | | | | | | | |
| 4. Mode of application test item to animals (stability, vehicle use route of administration dosing intervals) | of application of to animals Possible estrogenic effects of feed or bedding not considered. Oral dosing | | | | | | | | |
| 5. Appropriate anima model and strain sel | nimal Mice have not been used | | | | | | | | |
| 6. Suitability of samp measurements, sam times, and procedure | pling | | | Single time point | | | | | |

| 7. Suitability of biochemical measurements including quality control 8. Suitability of pathological/ functional assessment | | | Based on manufacturer specifications, quality control not addressed Blinding not discussed | | | | | |
|--|--|-----|---|----------|--|-------------------|---------------------|---|
| 9. Accessibility of raw data | | | | | | Summary data only | | |
| 10. Statistical analysis | | | Normality not checked, otherwise acceptable. | | | | | |
| Relevance and strength of | effects | 3 | | 2 | | | 1 | |
| to dose/tissue concentration interest resulting in adverse animals Relevance of model system assessed to key events occurrence. | Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in | | assessment of estrogenicity; e for assessment of key event. | | | | exposu | evels not related to human res or exposures in species with active effects |
| Strength of effects | | | | | | | relevan rats. Co | at a high dose level with unclear ce for exposure of humans or omparison of ED50 to that of ol was not provided. |
| Scoring | | | | | | | | |
| Quality of methods total 22 | | | | | | | | |
| Score3/domain 22/10 = 2.2 | | 2 | | <u> </u> | | | · | |
| Strength and relevance total | | 3 | | | | | | |
| Total score | | 6.6 | | | | | | |

| Study identification | Quinn AL, Regan JM, Tobin JM, Marinik BJ, McMahon JM, McNett DA, Sushynski CM, Crofoot SD, Jean PA, Plotzke KP. 2007b. In vitro and in vivo evaluation of the estrogenic, androgenic, and progestagenic potential of two cyclic siloxanes. Toxicol Sci 96, 145–153. Also: Output AL, 2003. Evaluation of actomethylogolatetrasilogona (DA) with return returns the acceptance of actomethylogolatetrasilogona (DA) with return returns the acceptance of actomethylogolatetrasilogona (DA) with return returns a construction of actomethylogonal construction of actomethyl |
|----------------------|--|
| | Quinn AL. 2003. Evaluation of octamethylcyclotetrasiloxane (D4) with rat uterotrophic assay using ovariectomized adult Sprague–Dawley rats. Dow Corning Corporation Health & Environmental Sciences Non-Regulated Technical Report No. 2003-0000-53144. |
| Design | Material studied: D4 >99% purity, inhaled concentration monitored at least once |
| _ | Type/Strain of animals: Ovariectomized female Sprague-Dawley and Fischer 344 rats |
| | Positive controls: Ethinyl estradiol, genistein for uterotrophic assay, ICI 182,780 for estrogen antagonist assay |
| | Concurrent controls: Filtered air |
| | Group sizes: 6 or 10 females per strain per dose group |
| | Vehicle: Filtered air for D4, corn oil for the positive controls |
| | Feed: Not indicated |
| | Dosing and age at dosing: Age of animals not indicated except adult. D4 inhalation at 700 ppm for 16 hours/day for 3 days. Positive controls injected |
| | subcutaneously each day for 3 days. |
| | Endpoints assessed: Uterine wet and blotted weight, uterine epithelial cell height. |
| Effects | D4 had estrogenic and antiestrogenic activity in the uterotrophic assay as assessed by uterine weight and epithelial cell height. |

| Quality of methods | 4 | 3 | 2 | 1 | 0 |
|-----------------------------------|-----------------------------|---------------------------|---|---|---|
| Chemical well characterized | | Well characterized and | | | |
| including presence | | monitored. Estrogenicity | | | |
| contaminants | | of feed or bedding not | | | |
| | | considered. | | | |
| General experimental | Sample size = 12/group, | | | | |
| design (number of animals per | design and sampling plan | | | | |
| dose group, controls, suitability | typical for uterotrophic | | | | |
| of study duration, housing | assay. | | | | |
| conditions) | | | | | |
| Assessment of possible | | Body weights assessed | | | |
| interference from stress due to | | but not considered in the | | | |
| restraint, toxicity | | analysis | | | |
| 4. Mode of application of test | Dose level and route | | | | |
| item to animals (stability, | were appropriate | | | | |
| vehicle used, route of | considering the | | | | |
| administration, dosing | reproductive data under | | | | |
| intervals) | evaluation | | | | |
| 5. Appropriate animal model | Use of Sprague Dawley | | | | |
| and strain selection | and Fischer 344 strains | | | | |
| | permits comparison | | | | |
| | between strains used | | | | |
| | traditionally in studies of | | | | |
| 0.0 % 1.3% | this compound. | | | | |
| 6. Suitability of sampling | Standard sampling | | | | |
| measurements, sampling | methods for this assay | | | | |
| times, and procedures | | | | | |
| 7. Suitability of biochemical | | Quality control not | | | |

| measurements including quality control | | | addressed | | | | |
|--|--|--|---|----------|--|---------|---|
| 8. Suitability of pathological/ functional assessment | | | Blinding not addressed | | | | |
| Accessibility of raw data | | | Summary data available | | | | |
| 10. Statistical analysis | | | Comparisons of relative potency to standard estrogens were missing. Normality was not checked prior to applying parametric tests. | | | | |
| Relevance and strength of effect | ets | 3 | | 2 | | 1 | |
| to dose/tissue concentrations of cl | Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in | | | | | exposur | vels not related to human res or exposures in species roductive effects |
| Relevance of model system and e assessed to key events occurring animals | | | compatible with key event in vivo n, model system applied is highly | | | | |
| Strength of effects Consistent and t | | ime- and dose-related change in ints, several measurements changes | | | | | |
| Scoring | | | | | | | |
| Quality of methods total | · · · · · · · · · · · · · · · · · · · | 32 | | | | | |
| Score/domain $32/10 = 3.2$ | | | | <u>-</u> | | | |
| Strength and relevance total | | 9 | | | | | |
| Total score | | 28.8 | | | | | |

| Study identification | octameth 63:37–46 Also: Turck PA using a ut | McKim JM Jr, Wilga PC, Breslin WJ, Plotzke KP, Gallavan RH, Meeks RG. 2001. Potential estrogenic and antiestrogenic activity of the cyclic siloxane octamethylcyclotetrasiloxane (D4) and the linear siloxane hexamethyldisiloxane (HMDS) in immature rats using the uterotrophic assay. Toxicol Sci 63:37–46. Also: Turck PA. 1998. Estrogenic and antiestrogenic activity of octamethylcyclotetrasiloxane (D4) in Sprague–Dawley and Fischer 344 immature female rats using a uterotrophic assay. Dow Corning report no. 1998-I0000-45425. | | | | | | | |
|---|---|--|---|---|---|---|--|--|--|
| Design | Type/Stra Positive of Concurred Group size Vehicle: Streed: Not Dosing an Endpoints | Material studied: D4 >99% purity, homogeneity of dosing solutions assessed Type/Strain of animals: Immature female Sprague-Dawley and Fischer 344 rats Positive controls: Ethinyl estradiol, diethylstilbestrol, coumestrol for uterotrophic assay; ICI 182,780 for estrogen antagonist assay Concurrent controls: Sesame oil Group sizes: 12 females per strain per dose group Vehicle: Sesame oil for the positive controls Feed: Not indicated Dosing and age at dosing: Sprague—Dawley rats began study at 18 days of age, Fischer 344 rats began study at 21 days of age. Endpoints assessed: Uterine weight (probably wet weight), uterine epithelial cell height. | | | | | | | |
| Effects | million tim rats. Com | D4 had estrogenic and antiestrogenic activity in the uterotrophic assay as assessed by uterine weight and epithelial cell height. D4 was 77,000 to 1.5 million times less potent than ethinyl estradiol in Sprague-Dawley rats amd 143,000 to 25-million times less potent than ethinyl estradiol in Fisher 344 rats. Comparisons to diethylstilbestrol resulted in similar orders of magnitude. D4 at 500 mg/kg/day decreased the uterotrophic response to ethinyl estradiol by a half order of magnitude. | | | | | | | |
| Quality of method | | 4 | 3 | 2 | 1 | 0 | | | |
| including presence contaminants | Chemical well characterized including presence contaminants | | Well characterized and monitored. Estrogenicity of feed or bedding not considered. | | | | | | |
| 2. General experim design (number of dose group, control of study duration, h conditions) | animals per ls, suitability | Sample size = 12/group, design and sampling plan typical for uterotrophic assay. | | | | | | | |
| 3. Assessment of possible interference from stress due to restraint, toxicity | | | Clinical signs were not related to D4 treatment but body weights were affected; not considered in the analysis. | | | | | | |
| 4. Mode of applicatitem to animals (stavehicle used, route administration, dosintervals) | ability, of ing | | | | Route and dose level not related to the route and dose level in the reproductive studies of interest. | | | | |
| 5. Appropriate anim and strain selection | | Use of Sprague Dawley and Fischer 344 strains permits comparison between strains used traditionally in studies of | | | | | | | |

| | this compou | und. | | | | |
|---|------------------------------------|------------------|-----------------------------------|---|---|--|
| 6. Suitability of sampling | Standard sa | | | | | |
| measurements, sampling | methods fo | r this assay | | | | |
| times, and procedures 7. Suitability of biochemical | Quality con | trol addressed | | | | |
| measurements including | Quality Con | iioi addiessed | | | | |
| quality control | | | | | | |
| 8. Suitability of pathological/ | | | Blinding not addressed | | | |
| functional assessment | | | | | | |
| Accessibility of raw data | Raw data a | | | | | |
| 10. Statistical analysis | | analysis and | | | | |
| | | of potency | | | | |
| | | rd estrogens | | | | |
| Relevance and strength of effe | | 3 | | 2 | 1 | |
| Concentrations applied and their | | | | | | vels not related to human es or exposures in species |
| to dose/tissue concentrations of | | | | | | roductive effects |
| interest resulting in adverse effect animals | as in | | | | | |
| Relevance of model system and | endpoint | Endpoint clearly | compatible with key event in vivo | | | |
| assessed to key events occurring | | in mode-of-actio | n, model system applied is highly | | | |
| animals | . | relevant | | | | |
| Strength of effects | Strength of effects Consistent and | | ime- and dose-related change in | | | |
| assessed endpo | | ints | | | | |
| Scoring | | | | | | |
| Quality of methods total 34 | | | | | | |
| Score/domain $34/10 = 3.4$ | | | | | | |
| Strength and relevance total | | 9 | | | | |
| Total score | | 30.6 | | | | |

| Study Identification | Lee D, Ahn C, An B-S, Jeu | ung E-B. 2015. Induction of the e | strogenic marker calbindin-D _{9k} l | oy octamethylcyclotetrasi | loxane. Int J Environ Res | | | |
|--|---|--|---|--|---------------------------|--|--|--|
| • | Public Health. 12:14610-1 | 4625 | | <u> </u> | | | | |
| Design Effects | Type/Strain of animals: fer Positive controls: Ethinyl e Concurrent controls: Unsp Group sizes: 5 pups from Vehicle: Unspecified Feed: AIN-76A (xenoestro Dosing and age at dosing: Endpoints assessed: Uteri There was no effect of D4 | Material studied: D4 of unstated purity from Sigma-Aldrich Type/Strain of animals: female Sprague-Dawley rats Positive controls: Ethinyl estradiol 3 µg/kg/day sc for 4 days Concurrent controls: Unspecified vehicle Group sizes: 5 pups from the same dam per dose group | | | | | | |
| | both dose levels of D4. | | | | | | | |
| Quality of methods | 4 | 3 | 2 | 1 | 0 | | | |
| Chemical well characterized including presence contaminants | | | | | Not described | | | |
| 2. General experimental design (number of animals per dose group, controls, suitability of study duration, housing conditions) | | | Only 5 animals per dose group, all from the same dam. Feed and housing conditions were appropriate. | | | | | |
| 3. Assessment of possible interference from stress due to restraint, toxicity | | | | | Not described | | | |
| 4. Mode of application of test item to animals (stability, vehicle used, route of administration, dosing intervals) | | | | Dose route not appropriate, dose levels were high. | | | | |
| 5. Appropriate animal model and strain selection | Sprague-Dawley rat is the strain of interest for reproduction studies. | | | | | | | |
| 6. Suitability of sampling measurements, sampling times, and procedures | Standard, methods for uterotrophic assay. | | | | | | | |
| 7. Suitability of biochemical measurements including quality control | | No mention of quality control | | | | | | |
| 8. Suitability of pathological/ functional | | Acceptable methods but blinding not addressed | | | | | | |

| assessment | | | | | |
|-------------------------------|-------------|----------------------|----------|--------------------------|------------------------------|
| Accessibility of raw | | | | Summary data only | |
| data | | | | | |
| 10. Statistical analysis | | Appropriate metho | ods were | | |
| | | used, but no asses | | | |
| | | normality of distrib | outions | | |
| Relevance and strength o | of 3 | | 2 | 1 | |
| effects | | | | | |
| Concentrations applied and | | | | | evant to evaluation of the |
| their relevance to dose/tissu | | | | adverse effect under co | nsideration |
| concentrations of chemical | | | | | |
| interest resulting in adverse | ; | | | | |
| effects in animals | | | | | |
| Relevance of model system | | | | | |
| and endpoint assessed to k | rey | | | | |
| events occurring in intact | | | | | |
| animals | | | | | |
| Strength of effects | | | | | ndings in cultured pituitary |
| | | | | cells and uterotrophic a | ssay = 0 |
| Scoring | | | | | |
| Quality of methods total | 21 | | | | |
| Score/domain | 21/10 = 2.1 | | | | |
| Strength and relevance total | al O | | | | |
| Total score | 0 | | | | |

Supplemental Material

Table 5. Quality assessment of studies supporting anti-androgenic effects of DEHP

A. In vitro

| A. In vitro | | | | | | |
|--|--|--|---|---|-----------------------------------|--------------------|
| Study | Svechniko | ov K, Svechnikova I, Söder O. 200 | 08. Inhibitory effects of mno-ethylhexyl | phthalate on steroidogenesis in in | nmature and adult rat Leyd | ig cells in vitro. |
| identification | Reprod T | oxicol 25:485–490 | | | · | |
| Design | Model sys Positive of Concurred Treatment | controls: cyclic AMP, glutethimide ont controls: 0 µM DEHP ot: MEHP in medium at 10, 100, or | cells isolated from testes of Sprague-D | | | |
| Effects | In immatu decrease | ire Leydig cells, cholesterol transp | port, testosterone, and DHT production | are inhibited by MEHP 250 μM ar | nd StAR and 5α-reductase | activity are |
| Quality of metho | ods | 4 | 3 | 2 | 1 | 0 |
| 1. Chemical well including present contaminants Avo contamination fro equipment/dosing Control of adsorp glassware causin interference, approximation to avoid levolatiles, limit of second controls, suitabilit duration. | ce oidance of om g solutions. otion onto ng ropriate oss of solubility) imental umber of /concentration, | | Small number of repeats (4 cultures for each concentration) | Reliance on supplier for information on identity and purity | | |
| 3. Mode of applic item system (stab used, route of application of applications) dosing intervals, actual concentrations chemical of interest | oility, vehicle plication, estimation of tion of | | | Concentrations in media only calculated based on amount added to system | | |
| All assessment determination of the model organism | ts include toxicity to | | | Very limited information on cytotoxicity | | |
| Suitability of sa method, sampling procedures. | g times and | | | | No justification of sampling plan | |
| 6. Suitability of bi | ochemical | | Complies with best practice for | | | |

| measurements including quality control 7. System used for biotransformation/cells have capacity to simulate relevant reactions that occur with | | | all measurements, control not address | | | | |
|--|-------------|--|---------------------------------------|---|--|---|--|
| chemical of interest in animals | | | | | | | |
| Accessibility of raw data | | | | | | Summary data reported | |
| 9. Statistical analysis | | | | | Statistical methods not discussed in detail, no evaluation of normality of distributions | | |
| Relevance and strength of effe | ects | 3 | | 2 | 1 | | |
| Concentrations applied and their to dose/tissue concentrations of interest resulting in adverse effect animals | chemical of | | | | | Concentration in culture is with respect to in anticipal concentrations | |
| Relevance of model system and assessed to key events occurring animals | | Endpoint or biomarker is with key event in vivo in model system applied is | mode of action, | | | | |
| Strength of effects | | | | | | | |
| Scoring | | | | | | | |
| Quality of methods total | 20 | | | | | | |
| Score/domain | 20/9 = 2.2 | | | | | | |
| Strength and relevance total | | 9 | | | | | |
| Total score | | 19.8 | | | | | |

B. In vivo

| Study | Akingber | ni BT, Youker RT, Sottas CM, Ge R, K | atz F. Klinefelter GR. Zirkin BR. F | lardy MP Modulation of rat Levdi | a cell steroidogenic functio | n by di(2- |
|--------------------------|-------------|---|-------------------------------------|---|-------------------------------|-----------------|
| Identification | | l)phthalate. Biol Reprod 2001; 65:125 | | iaray iiii . iiioaalalloii oi rat 20yal | g con clorolacycline fallolic | by an(<u>-</u> |
| Design | | studied: DEHP, >99% purity | 1200 | | | |
| 2 congin | | nin of animals: Long-Evans rats from 0 | Charles River | | | |
| | | controls: None | manos ravor | | | |
| | | nt controls: Corn oil vehicle | | | | |
| | | res: At least 7 dams per treatment gro | un | | | |
| | Vehicle: 0 | | ч | | | |
| | | t specified | | | | |
| | | nd age at dosing: Gavage treatment o | f pregnant animals with 100 mg/kg | n/day on destation days 12–21. T | here was additional treatm | ent durina |
| | | and additional treatment of postnatal r | | grady on goodation days 12 21. 11 | noro wao additional trodim | one dannig |
| | | s assessed: Serum LH and testostero | | roduction steroidogenic enzymes | | |
| Effects | | in serum testosterone in juvenile Lon | | | | |
| Quality of methods | 1 | 4 | 3 | 2 | 1 | 0 |
| 1. Chemical well char | racterized | | Material adequately | | | |
| including presence | | | characterized based on | | | |
| contaminants | | | supplier information | | | |
| 2. General experimer | ntal | | Suitable for purpose but lower | | | |
| design (number of an | | | number of animals/group, no | | | |
| dose group, controls, | iiiiaio poi | | positive or historic control data | | | |
| suitability of study du | ration | | available | | | |
| housing conditions) | ration, | | available | | | |
| 3. Assessment of pos | ssible | Well established exposure and | | | | |
| interference from stre | | sampling system, experienced | | | | |
| restraint, toxicity | 000 000 10 | facility and staff regarding animal | | | | |
| Toolium, toxiony | | handling | | | | |
| 4. Mode of application | n of test | - nanamig | Mode, dose, route, medium, | | | |
| item to animals (stabi | | | duration appear appropriate | | | |
| vehicle used, route of | | | but no measurements to | | | |
| administration, dosing | | | determine stability | | | |
| intervals) | 9 | | /homogeneity | | | |
| 5. Appropriate anima | l model | Appropriate species and strain | 3 | | | |
| and strain selection | | 11 1 | | | | |
| 6. Suitability of samp | ling | | Single sampling time | | | |
| measurements, samp | | | | | | |
| times, and procedure | | | | | | |
| 7. Suitability of bioche | | | Appropriate measurements, | | | |
| measurements includ | | | quality control not addressed | | | |
| quality control | • | | | | | |
| 8. Suitability of patho | logical/ | | Blinding not addressed. | | | |
| functional assessmer | nt | | | | | |

| Accessibility of raw data | | | | | Summary data reported | |
|--|--------------------------------------|-------------------|---|------------------------|-----------------------|--|
| 10. Statistical analysis | | | | Normality not assessed | | |
| Relevance and strength of effects | 3 | | 2 | | 1 | |
| Concentrations applied and their relevator to dose/tissue concentrations of chemic of interest resulting in adverse effects in animals | al studies | ant to other rat | | | | |
| Relevance of model system and endpo assessed to key events occurring in inta animals | | n mode of action, | | | | |
| Strength of effects | Clear dose-related findi exposure | ngs with juvenile | | | | |
| Scoring | | | | | | |
| Quality of methods total | 29 | | | | | |
| Score/domain | 29/10 = 2.9 | · | | · | | |
| Strength and relevance total | 9 | · | | · | | |
| Total score | 54.8 | • | | | | |

| | T = | | | | | |
|---|------------------|---|--------------------------------|------------------------------------|-------------------------------|------------------|
| Study Identification | | Metzdorff SB, Vinggaard AM, Brokken | L, Dalgaard M. 2006. Mechanism | s underlying the anti-androgenic e | ffects of diethylhexyl phthal | ate in fetal rat |
| Design | Motorial a | xicology 223:144–155. studied: DEHP of 99 % purity | | | | |
| Design | Type/Stra | nin of animals: Time-mated Wistar rats | from Taconic | | | |
| | | controls: None | TOTT TACOTIC | | | |
| | | nt controls: Vehicle | | | | |
| | | res: 8 dams per dose group | | | | |
| | Vehicle: 0 | | | | | |
| | | omin standard chow (No. 1324) | | | | |
| | | nd age at dosing: Adult based on weigh | nt, not otherwise indicated | | | |
| | | s assessed: mRNA and protein for ster | | topathology, ex vivo production of | testosterone | |
| Effects | | JC was at least 2 orders of magnitude | | | | |
| Quality of metho | ds | 4 | 3 | 2 | 1 | 0 |
| 1. Chemical well of | | Highly purified, well characterized | | | | |
| including presence | e | | | | | |
| contaminants | | | | | | |
| 2. General experir | mental | | | Small number of animals per | | |
| design (number of | f animals per | | | condition and sampling time | | |
| dose group, contro | ols, suitability | | | | | |
| of study duration, | housing | | | | | |
| conditions) | | | | | | |
| 3. Assessment of | | Well established exposure and | | | | Not considered |
| interference from | stress due to | sampling system, experienced | | | | |
| restraint, toxicity | | facility and staff regarding animal | | | | |
| | | handling | | | | |
| 4. Mode of applica | | | Mode, dose, route, medium, | | | |
| item to animals (s | | | duration appear appropriate | | | |
| vehicle used, rout | | | but no measurements to | | | |
| administration, do | sing | | determine stability | | | |
| intervals) | | | /homogeneity, intake | | | |
| 5. Appropriate ani | | Appropriate species and strain | | | | |
| and strain selection | | Complian appropriate for seel, f | | | <u> </u> | |
| 6. Suitability of sa | | Sampling appropriate for goals of | | | | |
| measurements, sa | | the study | | | | |
| times, and proced 7. Suitability of bio | | | Selection not complying fully | | | |
| measurements inc | | | with best practice, quality | | | |
| quality control | Juding | | control not discussed | | | |
| 8. Suitability of pa | thological/ | | Selection not complying fully | | | |
| functional assessr | • | | with best practice; e.g., | | | |
| Turicuoriai assessi | HOH | | replicate samples not run or | | | |
| | | | blinding not reported | | | |
| 9. Accessibility of | raw data | | Dimening flot reported | | Summary data reported | |
| z z z z z z z z z z z z z z z z z z z | | I . | 1 | 1 | | 1 |

| suitable for Checked for | e statistical method analysis of endpoint. or normal distribution | | |
|---|---|---|---|
| Relevance and strength of effects | 3 | 2 | 1 |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of interest resulting in adverse effects in animals | Doses used were relevant to other rat studies | | |
| Relevance of model system and endpoint assessed to key events occurring in intact animals | Endpoint or biomarker is clearly compatible with key event in vivo in mode of action, model system applied is highly relevant | | |
| Strength of effects | | Changes in endpoint or biomarker observed, but significant changes have limited dose or time-dependency | |
| Scoring | | | |
| Quality of methods total | 32 | | |
| Score/domain | 32/10 = 3.2 | | |
| Strength and relevance total | 8 | | |
| Total score | 256 | | |

| Study Identification | | huillier R, Li W, Wang Y, Martinez-Arg | | | | |
|---|------------|---|-------------------------------------|------------------------------------|----------------------------------|-------------------|
| | |) phthalate exerts both short-term and | liong-lasting suppressive effects o | n testosterone production in the r | at. Bioi Reprod. 78(6):1018 | -1028 |
| Design | | tudied: DEHP, purity not specified | ovelov roto | | | |
| | | in of animals: Time-mated Sprague-D | awiey rats | | | |
| | | ontrols: None | | | | |
| | | nt controls: Corn oil vehicle | A musica a su timo a maint | | | |
| | Vehicle: C | es: Fetuses from 3 dams per treatmer | it group per time point | | | |
| | | | | | | |
| | Feed: Not | d age at dosing: Gavage treatment of | progrant onimals with E9, 1250 m | valkaldov on acception dovo 14 to | the day of delivery | |
| | | assessed: Testes from GD20 fetuses | | | | o Histopothology |
| | | nometric measurements not discussed | | i lestosterone and DHT productio | in by letal of fleoriatal testes | s. Histopathology |
| Effects | | in fetal testis basal production of teste | | after treatment of prognant Spra | gue Daloy rate with 224 mg | r/ka/day |
| LIICUS | | one) or 117 mg/kg/day (DHT). | erosterone and uniyarotestosterone | e alter treatment of pregnant opra | gue-Daley rats with 254 mg | g/kg/uay |
| Quality of methods | (100100101 | 4 | 3 | 2 | 1 | 0 |
| 1. Chemical well char | racterized | | | | | Purity not |
| including presence | | | | | | described |
| contaminants | | | | | | |
| 2. General experimer | ntal | | Suitable for purpose but lower | | | |
| design (number of an | | | number of animals/group, no | | | |
| dose group, controls, | | | positive or historic control data | | | |
| of study duration, hou | | | available | | | |
| conditions) | Ü | | | | | |
| 3. Assessment of pos | ssible | Well established exposure and | | | | |
| interference from stre | ess due to | sampling system, experienced | | | | |
| restraint, toxicity | | facility and staff regarding animal | | | | |
| • | | handling | | | | |
| 4. Mode of application | n of test | | Mode, dose, route, medium, | | | |
| item to animals (stabi | | | duration appear appropriate | | | |
| vehicle used, route of | | | but no measurements to | | | |
| administration, dosing | g | | determine stability | | | |
| intervals) | | | /homogeneity | | | |
| Appropriate anima | l model | Appropriate species and strain | | | | |
| and strain selection | | | | | | |
| Suitability of sample | | | Two sampling times | | | |
| measurements, samp | | | | | | |
| times, and procedure | | | ļ., | | | |
| 7. Suitability of bioche | | | Appropriate measurements, | | | |
| measurements includ | ding | | quality control not addressed | | | |
| quality control | , | | 15, | | | |
| 8. Suitability of patho | | | Blinding not addressed. | | | |
| functional assessmer | ∩t | | | | | |

| Accessibility of raw data | | | | | Summary data reported | |
|--|----------------------------|-----------------|---|------------------------|-----------------------|--|
| 10. Statistical analysis | | | | Normality not assessed | | |
| Relevance and strength of effects | 3 | | 2 | | 1 | |
| Concentrations applied and their relevance | Doses used were releva | nt to other rat | | | | |
| to dose/tissue concentrations of chemical of | studies | | | | | |
| interest resulting in adverse effects in | | | | | | |
| animals | | | | | | |
| Relevance of model system and endpoint | Endpoint or biomarker is | | | | | |
| assessed to key events occurring in intact | with key event in vivo in | | | | | |
| animals | model system applied is | <u> </u> | | | | |
| Strength of effects | Clear dose-related finding | gs | | | | |
| Scoring | | | | | | |
| Quality of methods total | 26 | | | | | |
| Score/domain | 29/10 = 2.6 | | | | | |
| Strength and relevance total | 9 | | | | | |
| Total score | 23.4 | | | | | |

| Study Identification | | /, Numtip W, Grote K, Csanady GA, C in pregnant and nonpregnant rats and | | | e and its primary metabo | olite mono(2-ethylhexyl) |
|---|---|---|---|---|--------------------------|--------------------------|
| Design | Material s Type/Stra Positive of Concurred Group siz Vehicle: A Feed: Altr Dosing ar | in pregnant and nonpregnant rate and studied: DEHP of >99.5 % purity in of animals: Pregnant Sprague-Daw ontrols: Not applicable int controls: None ees: 2 or 3 rats per treatment and sampaqueous emulsion in Tween 80 omin standard chow (No. 1324) and age at dosing: Adult based on weigs assessed: Concentrations of parent and standard chow (No. 1324) and age at dosing: Adult based on weigs assessed: Concentrations of parent and standard chow (No. 1324) | ley rats from Charles River bling condition ht, not otherwise indicated | 100.142 | | |
| Effects | 1 . | 0 mg/kg/day decreased mRNA and pr | | enic enzymes in fetal Leydig cells | | |
| Quality of method | ls | 4 | 3 | 2 | 1 | 0 |
| Chemical well chemical including presence contaminants | | Highly purified, well characterized | | | | |
| 2. General experim design (number of a dose group, control of study duration, h conditions) | animals per ls, suitability | | | Small number of animals per condition and sampling time | | |
| Assessment of p interference from si restraint, toxicity | | | | | | Not considered |
| 4. Mode of applicat item to animals (sta | | | Mode, dose, route, medium, duration appear appropriate | | | |

| vehicle used, route of administration, dosing intervals) | | | but no measurement determine stability /homogeneity, intak | | | |
|--|----------------------|--|--|---|--------------|----------|
| 5. Appropriate animal model and strain selection | Appropriate | e species and strain | | | | |
| Suitability of sampling measurements, sampling times, and procedures | Sampling a the study | ppropriate for goals of | | | | |
| 7. Suitability of biochemical measurements including quality control | Appropriate control | e technique and quality | | | | |
| 8. Suitability of pathological/ functional assessment | Not applica | ble | | | | |
| Accessibility of raw data | | | | | Summary data | reported |
| 10. Statistical analysis | Not applica | ble | | | | |
| Relevance and strength of effe | | 3 | | 2 | 1 | |
| Concentrations applied and their to dose/tissue concentrations of interest resulting in adverse effect animals | chemical of | Doses used were releva studies | nt to other rat | | | |
| Relevance of model system and assessed to key events occurring animals | | Endpoint or biomarker is with key event in vivo in model system applied is | mode of action, | | | |
| Strength of effects | | Not applicable | Inginy rolovani | | | |
| Scoring | | sppnosero | | | | |
| Quality of methods total | | 23 | | | | |
| Score/domain | | 23/8 = 2.9 | | | | |
| Strength and relevance total | | 6 (of 6) | | | | |
| Total score | | 17.4 | | | | |

| Study | Kariyazono Y, Taura J, Hattori Y, Ishii Y, Narimatsu S, Fujimura M, Takeda T, Yamada H. 2015. Effect of in utero exposure to endocrine disruptors on fetal |
|----------------|--|
| Identification | steroidogenesis governed by the pituitary-gonad axis: a study in rats using different ways of administration. J Toxicol Sci. 40(6):909–916. |
| Design | Material studied: DEHP, purity not indicated |
| | Type/Strain of animals: Pregnant Wistar rats from Kyudo Co. |
| | Positive controls: None |
| | Concurrent controls: Corn oil vehicle |
| | Group sizes: 2 fetuses from each of 3–7 dams |
| | Vehicle: Corn oil |
| | Feed: Not specified |
| | Dosing and age at dosing: Oral dose (not otherwise specified) at 30, 100, or 300 mg/kg on GD 15 |
| | Endpoints assessed: StAR mRNA and protein |
| Effects | DEHP 100 and 300 mg/kg/day decreased StAR mRNA and protein |

| Quality of methods | 4 | 3 | 2 | 1 | 0 |
|-----------------------------------|--------------------------------|---|-----------------------------|-----------------------|----------------|
| Chemical well characterized | | | | | Not described |
| including presence | | | | | |
| contaminants | | | | | |
| General experimental | | | Small number of animals per | | |
| design (number of animals per | | | dose group | | |
| dose group, controls, suitability | | | | | |
| of study duration, housing | | | | | |
| conditions) | | | | | |
| 3. Assessment of possible | | | | | Not considered |
| interference from stress due to | | | | | |
| restraint, toxicity | | | | | |
| 4. Mode of application of test | | Mode, dose, route, medium, | | | |
| item to animals (stability, | | duration appear appropriate | | | |
| vehicle used, route of | | but no measurements to | | | |
| administration, dosing | | determine stability | | | |
| intervals) | | /homogeneity, route not | | | |
| Appropriate animal model | Appropriate species and strain | specified except as oral | | | |
| and strain selection | Appropriate species and strain | | | | |
| 6. Suitability of sampling | | Sampling time not justified | | | |
| measurements, sampling | | Sampling time not justified | | | |
| times, and procedures | | | | | |
| 7. Suitability of biochemical | | Appropriate measurements, | | | |
| measurements including | | quality control not addressed | | | |
| quality control | | 7 | | | |
| 8. Suitability of pathological/ | | Blinding not addressed. | | | |
| functional assessment | | 3 | | | |
| Accessibility of raw data | | | | Summary data reported | |

| 10. Statistical analysis | | | Normality not assesse | ed | |
|---|---------------------------|----------------------|-----------------------|----|--|
| Relevance and strength of effects | 3 | 2 | | 1 | |
| Concentrations applied and their relevance to dose/tissue concentrations of chemical of | Doses used were releva | int to other rat | | | |
| interest resulting in adverse effects in | Studies | | | | |
| animals | | | | | |
| Relevance of model system and endpoint | Endpoint or biomarker is | s clearly compatible | | | |
| assessed to key events occurring in intact | with key event in vivo in | mode of action, | | | |
| animals | model system applied is | highly relevant | | | |
| Strength of effects | Clear dose-related findir | ngs | | | |
| Scoring | | | | | |
| Quality of methods total | 21 | | | | |
| Score/domain | 23/10 = 2.1 | | | | |
| Strength and relevance total | 9 | | | | |
| Total score | 18.9 | | | | |

| Study | Moore RV | /, Rudy TA, Lin TM, Ko K, Peterson RI | E. 2001. Abnormalities of sexual d | evelopment in male rats with in uto | ero and lactational exposur | e to the |
|---|--|---|--|-------------------------------------|-----------------------------|--------------|
| Identification | antiandrog | genic plasticizer di(2-ethylhexyl) phthal | ate. Environ Health Perspect 109 | :229–237 | • | |
| Design | Material s | tudied: DEHP, 99% purity | | | | |
| | Type/Stra | in of animals: Sprague-Dawley rats fro | m Harlan Sprague Dawley | | | |
| | Positive c | ontrols: None | | | | |
| | Concurrer | nt controls: vehicle | | | | |
| | | es: At least 8 dams per treatment grou | р | | | |
| | | copherol-stripped corn oil | | | | |
| | | 5012 rat diet | | | | |
| | | d age at dosing: Gavage treatment of | | | | |
| | | assessed: Anogenital distance, puber | | | | |
| Effects | | oring with reduced anogenital distance, | | | lescended testes, incomple | te preputial |
| | separation | n, and reduced weights of testes and te | estosterone-dependent sex organs | s beginning at 375 mg/kg/day | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 |
| Chemical well char | ractorized | | | | | |
| | acienzeu | | Material adequately | | | |
| including presence | racienzeu | | characterized based on | | | |
| | acienzeu | | | | | |
| including presence | | | characterized based on | | | |
| including presence contaminants | ntal | | characterized based on supplier information | | | |
| including presence contaminants 2. General experimer | ntal iimals per | | characterized based on supplier information Suitable for purpose but no | | | |
| including presence contaminants 2. General experimer design (number of an | ntal nimals per suitability | | characterized based on supplier information Suitable for purpose but no positive or historic control data | | | |
| including presence contaminants 2. General experimer design (number of an dose group, controls, | ntal nimals per suitability | | characterized based on supplier information Suitable for purpose but no positive or historic control data | | | |
| including presence contaminants 2. General experimer design (number of an dose group, controls, of study duration, hou conditions) 3. Assessment of pos | ntal nimals per suitability using ssible | Well established exposure and | characterized based on supplier information Suitable for purpose but no positive or historic control data | | | |
| including presence contaminants 2. General experimer design (number of an dose group, controls, of study duration, hou conditions) | ntal nimals per suitability using ssible | sampling system, experienced | characterized based on supplier information Suitable for purpose but no positive or historic control data | | | |
| including presence contaminants 2. General experimer design (number of an dose group, controls, of study duration, hou conditions) 3. Assessment of pos | ntal nimals per suitability using ssible | | characterized based on supplier information Suitable for purpose but no positive or historic control data | | | |

| | ı | | T | | | 1 | |
|--|--|---|---|-----------|----------|----------------------|----|
| 4. Mode of application of test | | | Mode, dose, route, medium, | | | | |
| item to animals (stability, | | | duration appear appropriate | | | | |
| vehicle used, route of | | | but no measurements to | | | | |
| administration, dosing | ı, dosing | | determine stability | | | | |
| intervals) | | | /homogeneity | | | | |
| 5. Appropriate animal model | Appropriate species and strain | | | | | | |
| and strain selection | | | | | | | |
| 6. Suitability of sampling | | | Single sampling time | | | | |
| measurements, sampling | | | | | | | |
| times, and procedures | | | | | | | |
| 7. Suitability of biochemical | | | Appropriate measurements, | | | | |
| measurements including | | | quality control not a | addressed | | | |
| quality control | | | | | | | |
| 8. Suitability of pathological/ | | | Blinding not addressed. | | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | | | | | | Summary data reporte | ed |
| 10. Statistical analysis | | e statistical method | | | | | |
| | | analysis of endpoint. | | | | | |
| | | or normal distribution | | | | | |
| Relevance and strength of effe | | 3 | | 2 | | 1 | |
| Concentrations applied and their | | Doses used were relevant to other rat | | | | | |
| to dose/tissue concentrations of | chemical of | studies | | | | | |
| interest resulting in adverse effect | cts in | | | | | | |
| animals | | | | | | | |
| Relevance of model system and | Relevance of model system and endpoint | | Endpoint or biomarker is clearly compatible | | | | |
| assessed to key events occurring in intact | | with key event in vivo in mode of action, | | | | | |
| animals | | model system applied is highly relevant | | | | | |
| Strength of effects | | Clear dose-related findings | | | | | |
| Scoring | | | | | | | |
| Quality of methods total | | 31 | | | | | |
| Score/domain | | 29/10 = 3.1 | | | <u> </u> | | |
| Strength and relevance total | | 9 | | | <u> </u> | | |
| Total score | | 27.9 | | | | | |

| Study | NTP (National Toxicology Program). 2004. Diethylhexylphthalate: multigenerational reproductive assessment by continuous breeding when administered to | | | | | | | | | |
|--|---|--|---|---|---|---|--|--|--|--|
| Identification | Sprague–Dawley rats in the diet. Research Triangle Park, NC: National Toxicology Program; | | | | | | | | | |
| | | trl.ntis.gov/NTRL/dashboard/searchResults/titleDetail/PB2005107575.xhtml | | | | | | | | |
| Design | Material studied: DEHP, 99.8% purity | | | | | | | | | |
| | Type/Strain of animals: Sprague-Dawley rats from Charles River | | | | | | | | | |
| | Positive controls: None Concurrent controls: Control feed (1.5 ppm DEHP) | | | | | | | | | |
| | Group sizes: 17 pairs per treatment group | | | | | | | | | |
| | Vehicle: NIH-07 feed | | | | | | | | | |
| | Feed: NIH-07 feed | | | | | | | | | |
| | Dosing and age at dosing: Dietary DEHP at 1.5, 10, 30, 100, 300, 1000, 7500, or 10,000 ppm, corresponding to 0.12, 0.78, 2.4, 7.9, 23, 77, 592, and 77 | | | | | | | | | |
| | mg/kg/day in the F₀ animals; 0.09, 0.48, 1.4, 4.9, 14, 48, 391, and 543 mg/kg/day in the F₁ animals; and 0.1, 0.47, 1.4, 4.8, 14. F0 animals were weeks of age at initiation of dosing. | | | | | | | | | |
| | | | | | | | | | | |
| | 46, 359 mg/kg/day in the F ₂ animals | | | | | | | | | |
| | Endpoints assessed: Pubertal progression, anogenital distance, reproductive organ weights (other mutligeneration endpoints not addressed here) | | | | | | | | | |
| Effects | Reduced male anogenital distance and delayed preputial separation and testis descent. Epididymides and testes were smaller. Effects were seen at a dietary | | | | | | | | | |
| | | 500 ppm and highe | | | | | | | | |
| Quality of methods | | 4 | 3 | 2 | 1 | 0 | | | | |
| 1. Chemical well cha | racterized | Fully characterized by performing | | | | | | | | |
| including presence | | laboratory, source specified, CAS | | | | | | | | |
| contaminants | | given | | | | | | | | |
| 2. General experime | | Well designed for purpose | | | | | | | | |
| design (number of a | | including use of adequate controls. | | | | | | | | |
| dose group, controls of study duration, ho | | Adequate number of animals (n > 8), well justified sampling plan with | | | | | | | | |
| conditions) | using | adequate study duration | | | | | | | | |
| 3. Assessment of po | ssible | Well established exposure and | | | | | | | | |
| interference from stress due to | | sampling system, experienced | | | | | | | | |
| restraint, toxicity | | facility and staff regarding animal | | | | | | | | |
| | | handling | | | | | | | | |
| | 4. Mode of application of test Checks made on levels of test item | | | | | | | | | |
| item to animals (stability, | | in the feed, intake by animals | | | | | | | | |
| vehicle used, route of | | assessed, human relevant route of | | | | | | | | |
| administration, dosing intervals) | | administration, application during | | | | | | | | |
| 5. Appropriate animal model | | sensitivity window Appropriate species and strain | | | | | | | | |
| and strain selection | | Appropriate species and strain | | | | | | | | |
| 6. Suitability of samp | olina | Standard, appropriate evaluation | | | | | | | | |
| measurements, sampling | | times | | | | | | | | |
| times, and procedures | | | | | | | | | | |
| 7. Suitability of biochemical | | N/A: No biochemical | | | | | | | | |

| measurements including | measurements | | | | | | |
|--|--|---|-------------------------|---|---|----------|---|
| quality control | | | | | | | |
| 8. Suitability of pathological/ | Suitability of pathological/ | | Blinding not addressed. | | | | |
| functional assessment | | | | | | | |
| Accessibility of raw data | Access to a | all raw data | | | | | |
| 10. Statistical analysis | Appropriate nonparametric testing | | | | | | |
| | performed. | - | | | | | |
| Relevance and strength of effe | ects | 3 | | 2 | | 1 | |
| Concentrations applied and their | | Doses used were relevant to other rat | | | | | |
| to dose/tissue concentrations of | to dose/tissue concentrations of chemical of | | studies | | | | |
| interest resulting in adverse effects in | | | | | | | |
| animals | | | | | | | |
| Relevance of model system and endpoint | | Endpoint or biomarker is clearly compatible | | | | | |
| assessed to key events occurring in intact | | with key event in vivo in | | | | | |
| animals | | model system applied is | | | | | |
| Strength of effects | | Clear dose-related findings | | | | | |
| Scoring | | | | | | | |
| Quality of methods total | | 35 | | | | <u> </u> | |
| Score/domain | | 35/9 = 3.9 | | | | | |
| Strength and relevance total | | 9 | | | - | | - |
| Total score | | 35.1 | | | · | · | · |